



**ICC Medical Imaging Working Group**

**Boston, MA**

1 Nov 2014 08:30 (EDT)

The meeting was called to order at 08:30 am (EDT) by Craig Revie, chair of MIWG, with the following attendees:

Steve Smiley, SmileyColor/FTA  
Phil Green, Gjovik University College  
Heijo Reinl, CGS  
Markus Barbieri, Barbieri Electronics  
James Chang, Sharp  
Michael Flynn, HFHS, UofM  
Mitchell Marks, Toshiba  
Chaminda Weerasinghe, Toshiba  
Bas Hulsken, Philips  
J.P. Van de Capelle, X-Rite  
Robert Horn, AGFA  
Chris Bai, BenQ  
Max Derhak, Onyx Graphics  
Veronika Lovell, Sun Chemical  
Marti Maria, Hewlett Packard  
Marc Mahy, Agfa Graphics NV  
Kaida Xiao, University of Liverpool  
Hiroyuki Fukuda, Olympus  
Michael Chang, Kyocera  
Jeremie Pescatore, BioMereix  
James Vogh, X-Rite  
William Li, Kodak  
Tom Lianza, Photo Research  
Craig Revie, FFEI, Ltd.  
Po-Chieh Hung, Konica Minolta

Masahiro Yamaguchi, Tokyo Institute of  
Technology  
Andy Masia, X-Rite  
Debbie Orf, NPES  
Pinky Bautista, MGH

On-line attendees:

Allen Olsen, Leica Biosystems  
Glenn Davis  
Brandon Gallus, FDA  
Takashi Matsui, Eizo Corporation  
John Dalrymple  
Kathryn Epsig, Barco  
Yusuke Bamba, Eizo Corporation  
John Penczek, NIST  
Craig Revie, Fujifilm Corporation  
Christye Sisson, Rochester Institute of  
Technology  
John Sweeney, BenQ Corporation  
Dave Wyble, Avian Rochester, LLC  
Kaida Xiao, Technical Consultant  
Albert Xthona, Barco NV  
Masahiro Yamaguchi, Tokyo Institute of  
Technology

After self-introductions and a check of sound quality, Mr. Revie proposed the agenda for the meeting as follows:

## **Whole Slide Imaging**

1. Round robin assessment status, objectives and next steps
2. Review of Leica's assessment
3. Review of Konica Minolta assessment and additional observations
4. Review of FFEI, Leeds and FDA assessment
5. Color Translator tool -possible use in calibration assessment
6. Sierra slide accelerated fading testing and extended slide

## **Multispectral Imaging**

7. Status update
8. DICOM proposal for multispectral extensions
9. Demonstration of MCS connection with actual profiles and images

## **Skin Colour**

10. Spectral measurement project status update

## **Medical displays**

11. mRGB and dRGB update
12. Experiences in building an ICC Profile for an mRGB display
13. Characterisation of displays
14. Preparation of reference images for display testing

## **Medical Photography**

15. Status update and next steps for digital photography workflow white paper

## **Petri plate imaging**

16. A colour calibration method for Petri plate image acquisition
17. The need for standardisation of reading and viewing of Petri plates

FDA Working Group for Whole Slide Imaging

Medical Imaging Working Group activities for 2015

## **Whole Slide Imaging**

### **1. Round robin assessment status, objectives and next steps**

Mr Revie presented an update on the Sierra slide evaluation [see attached]. He described the Sierra calibration assessment slide, which uses a biopolymer to carry biomarker stains. The assessment focused on two areas: determining the transmittance of the patches, and scanning the slide on a calibrated scanner with an ICC profile. These two tasks will help understand how colour is being processed by digital microscopes. Several people have measured the slide, and there is good agreement between measurements. The most significant variation was probably due to fading of the Eosin stain. Five participants have completed the slide scanning, and a further five are in progress and are expected to complete by the end of the year. Participants are discussing how to publish the results, which will probably be anonymized.

Some other issues that need to be considered are:

- The gamut limit of sRGB (Eosin in particular is outside)
- The numerical aperture (NA) of the system (an issue being that measurements are made using collimated light, while scans are usually not)
- The encoding of over-range white (which is clipped in the ICC profile).

It was noted that NA differences were intentional, and the effect needed to be quantified in order to consider how to handle it.

Mr Revie summarised the main goal as a framework for conformance. One option was to agree a standard calibration method, but this could be difficult.

Dr Bas Hulsken noted that idea of 'true colour' was not applicable to whole-slide imaging. Staining is highly variable. More important is to have consistency in scanning – agreement between different scanners was more important than colour accuracy. Dr Allan Olsen agreed, adding that more uniformity between vendors was needed, and a standard viewing experience was the ideal. Mr Revie suggested that an option to show a calibrated image was provided in the scanner interface it would help users to understand what they were looking at, and this calibrated image could be a basis for consistency between scanners. Dr Hulsken also noted that regulatory approval is important, and if viewing options other than the calibrated image are provided, it might be necessary to flag to the user that the resulting image did not meet conformance requirements.

The issue of viewer displays was considered out of scope for this activity, although it was acknowledged that work on displays was needed to ensure consistency of presentation. A colour managed display might be sufficient, and GSDF may not be required for this application. The sRGB colour space is not suited to encoding stain colours as some stains are outside the sRGB gamut.

Mr Revie reported that it had been difficult to encode images in the DICOM format than expected, and in the short term a different way of assessing the results may be needed.

Dr Brendan Gallas stated that he would welcome a baseline calibration procedure.

Mr Revie concluded this discussion, stating that the next steps in the work would be reviewed after presentation of the results of the calibration slide evaluation.

## **2. Review of Leica's assessment**

Dr Olsen reported results of Leica's assessment of the calibration slide [see attached]. He had computed results in Matlab and found there was good agreement between these and the results from the Sierra analysis tool. The visual comparison was also very good. He had built a profile (with help from FFEI) and was happy with the results from this. However, an average profile, using data from multiple scanners, might be desirable.

Dr Olsen identified sources of variability in the process, and summarised recommendations, the principle one being the scanner profiling procedure.

The light source in the Leica scanner is a blue pumped LED. No IR illumination is emitted by this source, so IR filtration is not needed. The scanner sensor is a trilinear array with R, G, B dye filters.

Dr Po-Chieh Hung observed that some white LEDs have low power at certain wavelengths, and can reduce quality depending on the spectral reflectance of the stain. Using the spectral power distribution of the source, and the spectral sensitivities of the sensors and optical system, it was possible to compute a Q factor, which essentially defined the device metamerism. He agreed to document the Q factor calculation for the group, using the manufacturers' published sensor sensitivities. It was noted that there was a standard procedure for measuring sensor sensitivity, and that if possible the sensitivity of the whole system should be measured, including the scanner optics.

## **3. Review of Konica Minolta assessment and additional observations**

Dr Hung presented results from the assessment at Konica Minolta [see attached]. He had tested the slide with a fluorescence microscope. He reported that a diffuse:0 or a diffuse:large-aperture geometry is the best way to illuminate the slide, and noted that dust rings / bubbles were apparent on the scans. He showed an analysis of the illumination quality using the CQS spreadsheet from Dr Ohno.

Dr Hulsken observed that in his experience non-linear correction does not work, and a 3x3 scene analysis matrix gives best results since sensors are linear, and accuracy depends on the degree to which the sensor spectral sensitivities conform to the Luther condition (that sensitivities are a linear transform of colour matching functions). In his work he had tried to make a scanner for a broad range of samples rather than an individual stain. It was reported that approximately 80% of pathology stains at Leeds were eosin, followed by DAB at 15%. The remaining 5% comprised a wide range of stain types.

Mr Revie commented that multiple calibration slides might be needed to cover the different stain types, instead of the current one which is mainly H&E (haematoxylin and eosin), but using such calibration assessment slides might be difficult in practice due to the batch processing frequently performed in clinical practice. Mr Hulsken suggested another approach might be to build a stain detection system, which would have no need for colour management.

Mr Revie noted that a non-linear correction is used in the FFEI profile posted on the Sierra web site, but FFEI attempt to avoid distortion while minimising errors. Mr Revie agreed with Mr Hulsken that this is difficult to achieve but can be done using similar technology to that used in Graphic Arts scanners.

#### **4. Review of FFEI, Leeds and FDA assessment**

Mr Revie presented results of assessments carried out at FFEI, Leeds, Ventana and FDA [see attached]. Slide measurements (FFEI, Ventana, FDA) showed good agreement, the main difference being between the reflectance of eosin. This could be due to fading of eosin, or the use of different light sources.

He described the FFEI calibration method and demonstrated the use of the project Sierra web site. Sample CIELAB values had been obtained by applying profiles to RGB values averaged across each patch. FFEI results were consistent with the measurements, but Leeds and FDA differences were much higher. It was noted that the latter two had used the same model of Leica Aperio scanner. Mr Allen Olson of Leica stated that older scanners have no colour correction, being adjusted according to customer feedback.

It was found that different scanners imaged different portions of the slide, and a revised slide layout is being tested. While operators would normally focus on the comb element on the cover slip, it was noted that this would result in the colour patches being slightly out of focus.

Mr Revie also reported on some tests on the visual assessment method proposed by Yagi and Bautista, whereby the slide image on a display was compared to the original slide using the display backlight. This seemed to work well, and had received positive feedback from the pathologist at Leeds.

#### **5. Color Translator tool -possible use in calibration assessment**

Marti Maria demonstrated his Colour Translator tool (<http://www.littlecms.com/professional/>). This converts images using ICC profiles, and enables a default source profile to be defined for untagged images without having to embed the profile. It recognises abstract profiles and supports many image file formats and encoding precisions, including 8- and 16-bit integer and 16- and 32-bit floating point. The tool is extremely fast in operation, using parallel processing, and is suited to batch processing of images. Mr Maria stated that it was available at a low cost of possibly free for research and academic use.

## **6. Sierra slide accelerated fading testing and extended slide**

Mr Revie reported results from a fading test of the calibration assessment slide [see attached]. Four different exposure protocols had been tested, using a halogen lamp at different intensities and durations. Results showed that in the full exposure phase there were significant changes in the reflectance of eosin, and that the change was larger for slides with a small amount of eosin stain. Mr Revie stated that it was possible to improve the slide manufacturing process by applying DABCO stabiliser as a separate step.

It is planned to employ the fading of eosin in future slides to infer the degree of light exposure undergone by the slide, and it was intended to estimate the number of scans that can be made before a slide changes unacceptably (possibly around 1-2 units of CIELAB 1976 colour difference).

FFEI are also developing a calibration slide, which currently has 55 patches covering the majority of stains used at Leeds. FFEI are evaluating the feasibility of this slide, but would look for another partner to manufacture. The format is currently too large for some scanners and may need adjustment. Dr Bautista suggested IHC as a further stain candidate, if different from DAB. It might also be useful to add tissue-based spectra such as haemoglobin.

## **Multispectral Imaging**

### **7. Status update**

Professor Yamaguchi provided an update of work on multi-spectral imaging [see attached]. Multi-spectral imaging is used in brightfield and fluorescence imaging, primarily in research rather than clinical practice. Yamaguchi-sensei showed a general model for spectral unmixing to infer biomarker quantities. It was desired by the DICOM committee that the original image is retained and a separate unmixing transform is provided. He showed two possible solutions: in the first an ICC v4 profile was used to define a virtual input device, where each channel represents a biomarker, and an ICC device link profile can be used to generate a biomarker image from a real multi-spectral image; while in the second solution an iccMAX Material Connection Space profile was used.

Yamaguchi-sensei described the current status of this work and a planned test implementation. He welcomed contributions.

### **8. DICOM proposal for multispectral extensions**

Dr Bas Hulsken presented an update of his work to bring multispectral extensions to DICOM [see attached]. He had discussed this in WG26, so far with limited progress, and he hoped to get feedback from DICOM experts in this MIWG meeting.

He showed the extensions need to DICOM; some are simple, but adding modules defining needed functionality is more complex. Multispectral images could possibly be stored as multiple DICOM images. It was desirable to support sub-sampling, so it was not appropriate to impose a common resolution limit, although it was noted that variable resolution is typically only seen in WSI imaging. In the case of PET and CT, images have different resolutions but the presentation state has to combine them. For image alignment, fiducials are commonly used to map coordinates, with both linear and non-linear deformation fields defined (e.g. to compare pre- and post-excision images).

As a next step Dr Hulsken wished to work with DICOM experts to develop proposals. There was a need to consider use cases – initially this might just be WSI and possibly Petri dish images. It was suggested that Dr Hulsken contact DICOM WG6.

Professor Yamaguchi also suggested the need to consider hyperspectral imaging, and issues like interleaving, channels per plane etc. – some systems such as remote sensing are line-at-a-time.

### **9. Demonstration of MCS connection with actual profiles and images**

Mr Max Derhak provided a demonstration of how iccMAX Material Connection Space (MCS) profiles can be used to provide quantitative representations of material amounts in an image [see attached]. He showed the workflows and connection rules, and an example of using MCS profiles to connect to material identification and visualization outputs.

The next step would be to develop an Interoperability Conformance Specification (ICS) for one or more medical imaging modalities, which Mr Derhak undertook to do in conjunction with Phil Green and anyone else interested in contributing.

### **Skin Colour**

#### **10. Spectral measurement project status update**

Kaida Xiao presented a summary of work at University of Liverpool to evaluate methods of measuring human skin and to build a database of skin measurements [see attached]. By determining the reflectance properties independently of the illumination, it was possible to connect the results with skin chromophores such as melanin and haemoglobin.

Dr Xiao described the procedures used to measure skin. Calibration was considered very important, and this is performed with the aid of a test chart. A diffuse illumination is used, and a white card is imaged in order to recover the spectral power of the illumination. He had also explored the use of 3D photogrammetry, saving a 3D image.

Reflectances were reconstructed from RGB image data using the skin colour database.

### **Medical displays**

#### **11. mRGB and dRGB update**

Dr Michael Flynn presented a progress report on dRGB (formerly mRGB) [see attached]. He showed the background to this work, including previous display standards, and introduced the report of AAPM Task Group 196 on requirements for medical displays. The neutral luminance scale had a variable  $L_{max}$  (white point) and used the DICOM GSDF to define the relationship between RGB and luminance. The default colour gamut was sRGB, but others could be indicated by the profile description. Issues such as tolerances for the primaries, methods of display calibration and the use of source and destination ICC profiles, will be addressed in the dRGB standard.

#### **12. Experiences in building an ICC Profile for an mRGB display**

Mr Revie presented an analysis of display profiling requirements for dRGB [see attached]. The main issue is that LUT-based profiles are necessary, and few profiling applications support this (as opposed to Matrix/TRC profiles). Those commercial applications that do generate LUT-based profiles tend to modify the graphics card LUT at the calibration stage, which may be undesirable, especially with self-calibrating displays and with mixed-mode presentation. He proposed that ICC MIWG should specify requirements for display profiling software and work with vendors to test and deliver suitable profile creation software.

### **13. Characterisation of displays**

It was noted that display calibration practices vary, and that users have little guidance on how to do this. Dr Green undertook to make information available on the ICC web site.

### **14. Preparation of reference images for display testing**

Dr Hulsken presented an update on the call for reference images for use in testing displays and display calibration [see attached]. The aim was to validate a proposed display architecture that had been presented in previous MIWG meetings, and for this a limited set of representative images are required, together with a specification of the minimum performance required for the visualization. He proposed to continue collecting images and display measurement data, and to hold a conference call in December to select the display systems and reference images in order to proceed with bench testing and simulation in early 2015.

## **Medical Photography**

### **15. Status update and next steps for digital photography workflow white paper**

Dr John Penczek provided a draft "Recommended Image Capture Workflow for Medicine Photography" to the meeting [see attached]. This is a proposed best-practice guideline for medical capture workflow with a focus on colour accuracy. The draft lists the equipment needed, a basic outline of procedure and a summary of colour correction methods to use. The draft document will be posted on the ICC MIWG website.

Some of the areas discussed include:

1. The use of a test chart in the field of capture
2. Use of diffuse illumination would avoid some of the problems seen with directional lighting.
3. Camera RAW format is better for colour accuracy as it avoids the in-camera rendering applied in JPEG images.

## **Petri plate imaging**

### **16. A colour calibration method for Petri plate image acquisition**

Dr Jeremie Pescatore of Biomerieux presented a summary of colour calibration for Petri plate scanners and related issues [see attached]. He gave an overview of automated Petri plate scanning workflow and the use of ICC input profiles to transform from scanner RGB to CIE colorimetry. It is difficult to validate the results on Petri plate colonies since the colonies are too small to measure with the spectrophotometric equipment available in his lab, and he showed the InVivo hyperspectral imaging system which was built for this type of application. His goals were to build a standard setup to measure Petri plates with both specular included and excluded; and to develop an imaging system that could have similar accuracy in reflectance measurement to a spectrophotometer.

### **17. The need for standardisation of reading and viewing of Petri plates**

Dr Pescatore reviewed the need for standardisation in reading and viewing Petri plate images [see attached]. He discussed the main requirements of reading and viewing environments, and identified open issues as:

- How should the effect ambient light be incorporated?
- How should the effect of the displayed background be taken into account?
- How should the display gamut be standardized?
- How should the display profile be attached or embedded with the Petri image?
- Could mRGB or dRGB colour spaces incorporate these considerations for Petri images?

It was agreed to establish an activity area on this topic on the ICC MIWG web site.

#### **FDA Working Group for Whole Slide Imaging**

Dr Brendan Gallas showed information about the FDA WSI group [see attached] and invited ICC MIWG members to participate.

#### **Medical Imaging Working Group activities for 2015**

Craig Revie agreed to prepare a schedule of ICC MIWG meetings for 2015, in conjunction with Debbie Orf and Phil Green.

Mr Revie closed the meeting at 12:00pm.

#### **Action items**

##### **Sierra project**

- Report results of round robin test to MIWG (Craig)
- Review next steps after presentation of results (Craig)
- Provide documentation on Q factor for assessing image quality (Po-Chieh)

##### **Accelerated fading**

- Modify manufacturing process to apply DABCO stabiliser as separate step (Craig)
- Estimate no of scans that can be made before slide fades unacceptably (Craig)

##### **FFEI calibration slide**

- Consider adding tissue-based spectra (e.g. haemoglobin) (Craig)
- Consider adding IHC stain (Craig)

##### **Multi-spectral**

- Provide contributions on proposed workflow and test implementations to Yamaguchi-san (all interested)

##### **DICOM**

- Contact DICOM WG6 to discuss extending DICOM format for colour support (Bas Hulsken)
- Develop ICS for medical imaging modalities (Derhak, Green)

##### **Display calibration for WSI**

- Define requirements for display profiling software and work with vendors to test and deliver suitable profiling software
- Add notes on display calibration to ICC web site (Green)

##### **Petri dish imaging**

- Establish activity page for Petri dish imaging on ICC MIWG web site (Green,

##### **Next meetings**

- Set up schedule of 2015 meetings (Revie, Green, Orf)

A full recording of the meeting is available at <http://www.npes.org/Portals/0/standards/2014-11-01%2008.33%20ICC%20Regular%20Meetings.wmv>



# ICC Medical Imaging Working Group

**Face-to-face meeting in Boston, USA**

**1<sup>st</sup> November 2014**

# First of all some breaking news from 'The Beautiful Game'



## Manchester United topped Liverpool in final; **Did it raise ICC profile?**

Nicholas Mendola | Aug 5, 2014, 10:38 AM EDT

 3 Comments

A trophy was kissed after a championship match played between Liverpool and Manchester United on Sunday in Miami, a title tilt played as part of the two clubs' preseason tours of the United States. United claimed a 3-1 decision in the final game of the 2014 Guinness International Champions Cup (Wayne Rooney, Juan Mata and Jesse Lingard scored for Manchester, while Steven Gerrard scored for Liverpool).

And despite 51,000 fans in attendance at Sun Life Stadium, the question remains: how far away is the tournament from having relevance outside of big names and big crowds?



## ICC: Wayne Rooney player of the tournament



There's no question the tournament itself was a success in terms of attendance and promotion of the clubs branding in America. And many of the matches were a good watch, as fans got to check out superstars plying their trade for major clubs.

And, as mentioned, on the one hand, there were the championship photos and celebrations...



 Manchester United   
@ManUtd



Manchester United: ICC champions! #mutour

3:27 AM - 5 Aug 2014

Clearly ICC Profiles are starting to be used in areas we had not anticipated – are there other sports that could benefit from our help?

# ICC MIWG Agenda for Saturday 1<sup>st</sup> November 2014

- **Whole Slide Imaging**
  - 1. Round robin assessment status, objectives and next steps Craig Revie
  - 2. Review of Leica's assessment Allen Olson
  - 3. Review of Konica Minolta assessment and additional observations Po-Chieh Hung
  - 4. Review of FFEI, Leeds and FDA assessment Craig Revie
  - 5. Color Translator tool – possible use in calibration assessment Marti Maria
  - 6. Sierra slide accelerated fading testing and extended slide Craig Revie
- **Multispectral Imaging**
  - 7. Status update Masahiro Yamaguchi
  - 8. DICOM proposal for multispectral extensions Bas Hulsken
  - 9. Demonstration of MCS connection with actual profiles and images Max Derhak
- **Skin Colour**
  - 10. Spectral measurement project status update Kaida Xiao

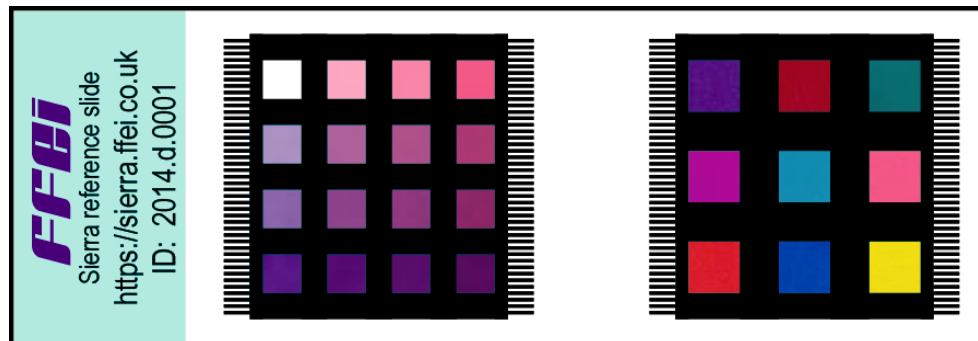
## ICC MIWG Agenda for Saturday 1<sup>st</sup> November 2014

- **Medical Displays**
  - 11. mRGB and dRGB update Mike Flynn
  - 12. Experiences in building an ICC Profile for an mRGB display Craig Revie
  - 13. Characterisation of displays Mike Flynn / Tom Kimpe
  - 13. Preparation of reference images for display testing Mike Flynn / Tom Kimpe
- **Medical Photography**
  - 14. Status update and next steps for digital photography workflow white paper Phil Green / John Penczek
- **Petri plate imaging**
  - 15. A color calibration method for Petri plate image acquisition Jérémie Pescatore
  - 16. The need for standardisation of reading and viewing of Petri plates Jérémie Pescatore
- **17. FDA Working Group for Whole Slide Imaging** Brandon Gallas
- **Medical Imaging Working Group activities for 2015** Craig Revie

# Sierra calibration assessment slide

## *Round-robin status, objectives and next steps*

*1<sup>st</sup> November 2014*



# Round robin assessment **status**, objectives and next steps

- **Slide evaluation**
  - Completed: FFEI, Leeds, FDA, Leica, MGH
  - In progress: Omnyx, Datacolor, Ventana, Philips, Konica Minolta
  - Not yet started: Tokyo Institute of Technology, Ghent University
- **Some initial results have been presented**
  - Measurements of slide are in reasonably good agreement
  - Analysis of slide images has been difficult in some cases as it has been difficult for some vendors to create DICOM images
- **Publication of results**
  - Round-robin participants are discussing how to publish the comparative results and whether they should be anonymised
- **Other feedback**
  - sRGB gamut limitation; effect of system NA; encoding over-white

# Round robin assessment status, **objectives** and next steps

- **Objectives**
  - make and compare measurements of slides
  - scan slides on a range of digital microscopes and provide feedback
  - **provide feedback on framework for digital microscope evaluation**
- **Framework for digital microscope evaluation options**
  - agree and document an assessment method
  - alternatively agree a standard calibration method
- **Publish results**
  - objective could be to show the current colour behaviour for digital microscopes in use today
  - an alternative approach could be for vendors to improve their calibration before measuring the colour result



# Round robin assessment status, objectives and **next steps**

- **DICOM format has proved more difficult than expected**
  - Is this still a good long-term solution?
  - In the short term we need an alternative assessment method
- **Possible alternative assessment methods**
  - Export low-resolution image with embedded ICC Profile
  - Provide RGB image values for each patch along with an ICC Profile
  - MATLAB or Image J + R or some other tools
- **Possibly document a baseline calibration procedure**
- ***Review next steps following presentation of results***

# **Calibration Assessment Slide Results for ScanScope AT2**

**Allen H. Olson, PhD  
Leica Biosystems**

**ICC Medical Imaging Working Group – 01 Nov 2014**

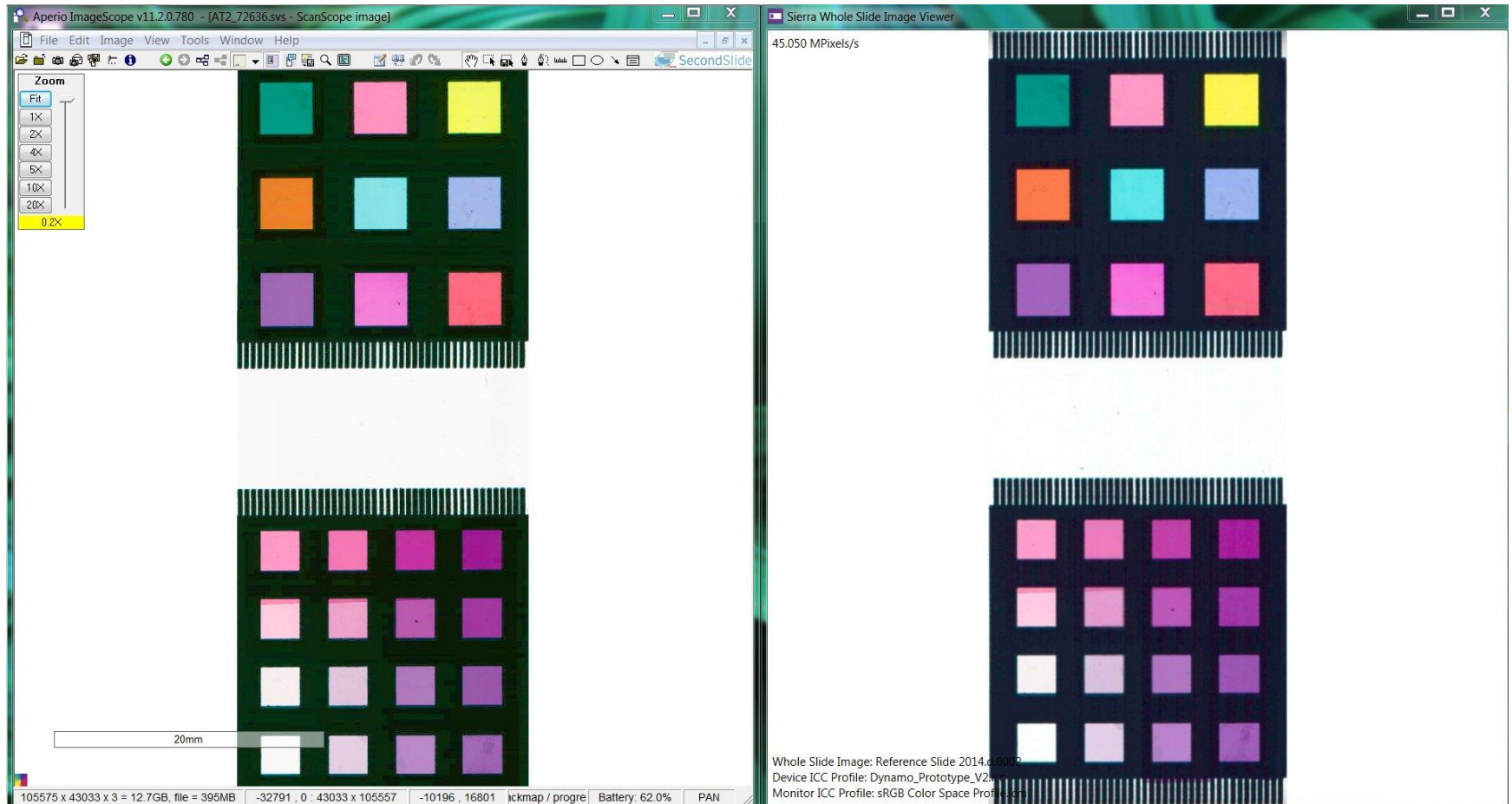
## Overview

- Experience with Sierra Website
- Scanning of the test slide
- Visual comparison – Leica/FFEI
- Processing with Matlab
- Results
- Sources of variability/error

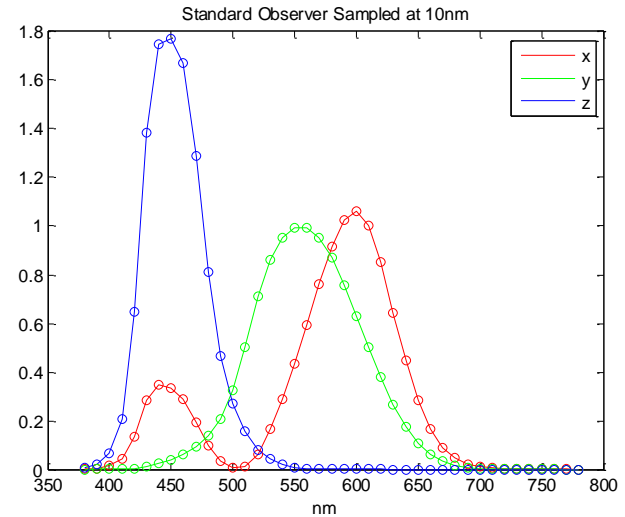
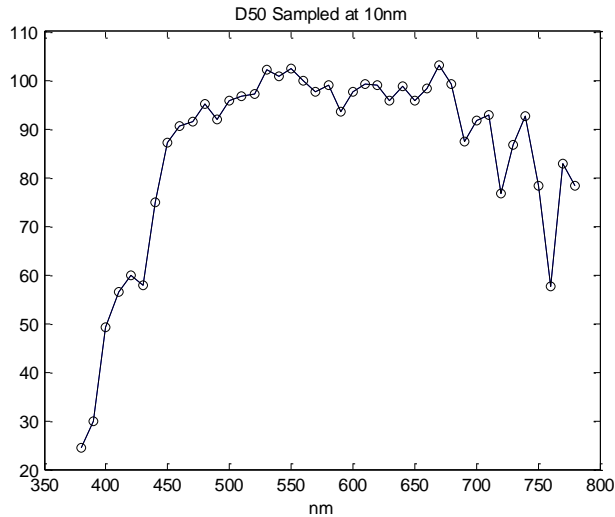
- Downloaded Sierra WSI Viewer
  - This was used to view FFEI WSI image for test slide
- Downloaded Sierra DICOM converter
  - This worked without issue on Leica ScanScope WSI image
- Downloaded spectral data file for my particular test slide
  - 2014.d.0002.csv, opened in Excel
  - The spectral data for each patch was extracted and utilized to calculate reference XYZ and Lab values for each patch
- Sierra Analysis Tool (web based)
  - Had some difficulty.
  - Profile was too large. Thanks to Simon Davidson (FFEI) for website software fix. Afterwards, it worked just fine.
  - Produced csv files with ReferenceLab and Sample Lab & RGB for WSI of target slide.
  - Sierra results agreed with independent Matlab calculations

- Target scanned on Leica AT2 scanner
- Manual scanning mode
  - Selected scan area to include target patches
  - Manual pre-focus on each target patch
  - Calibration taken on clear area between blocks
- Magnification - 20X
- Compression – JPEG, default setting
- File format – TIFF (svs)

- Aperio Imagescope to view AT2 image
- Sierra WSI Viewer to view FFEI image
- Color management using vendor ICC and sRGB monitor profiles

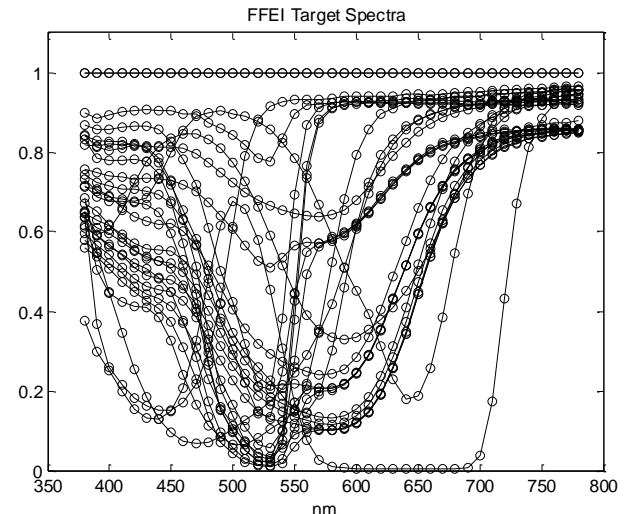


- Calculate Reference XYZ, Lab values
  - Define Illuminant(D50)
  - Integrate target transmittances against standard observer curves
- Estimate WSI RGB values
  - Down-sample WSI to manageable size
  - Average central  $\frac{1}{4}$  of each target patch
- Transform WSI/RGB to D50 PCS values
  - Use vendor calibrated ICC profile (eg, AT2.icc)
  - Matlab functions to convert RGB to XYZ, Lab, sRGB
- Compare Reference to WSI Color values



$$\langle X_j, Y_j, Z_j \rangle = \frac{1}{W_y} \sum_{i=1}^{41} \langle x_i, y_i, z_i \rangle D50_i T_{ij}$$

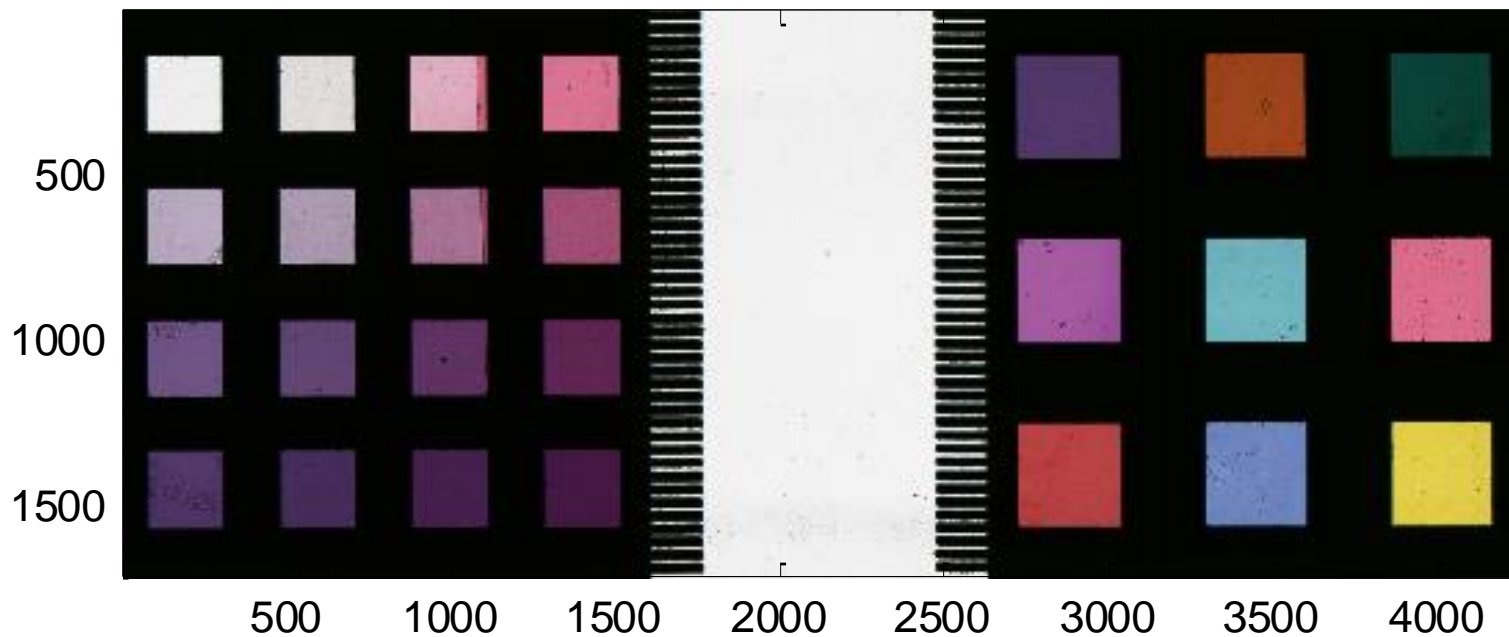
- Integral is approximated by summation
- $W_y$  is normalization ( $T=0$ )
- XYZ to Lab transform using library



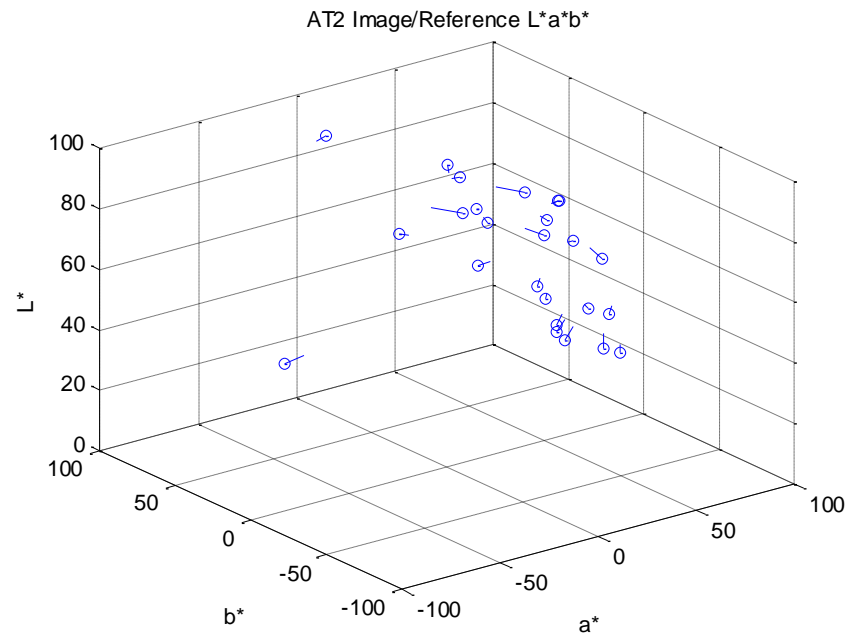
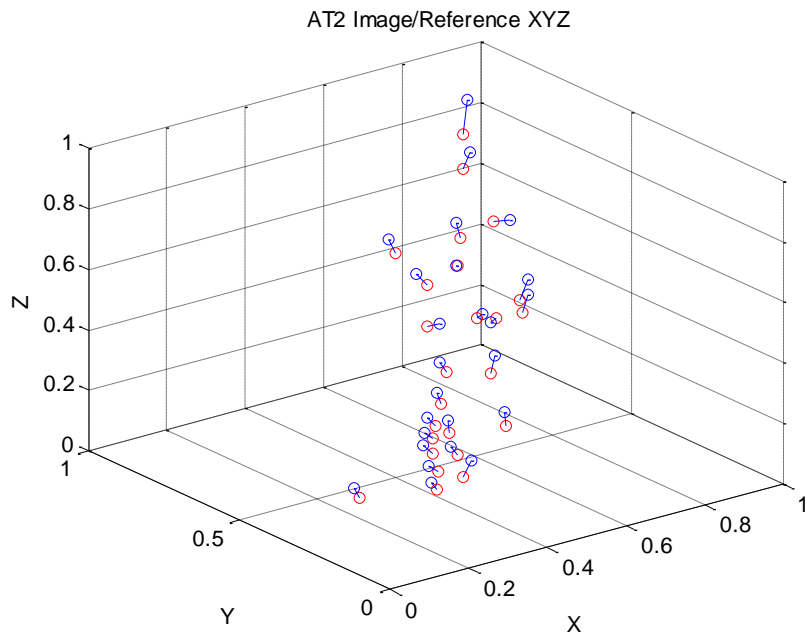


- Down-sample WSI to manageable size (1700x4000)
- Click on centers of corner patches for each block
- Average central  $\frac{1}{4}$  of each target patch

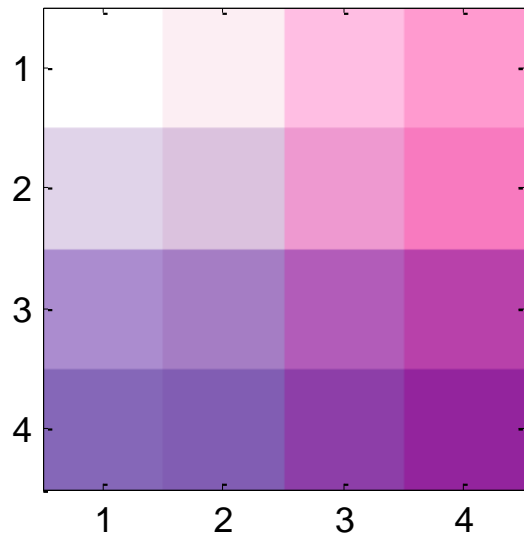
AT2 Input Image File



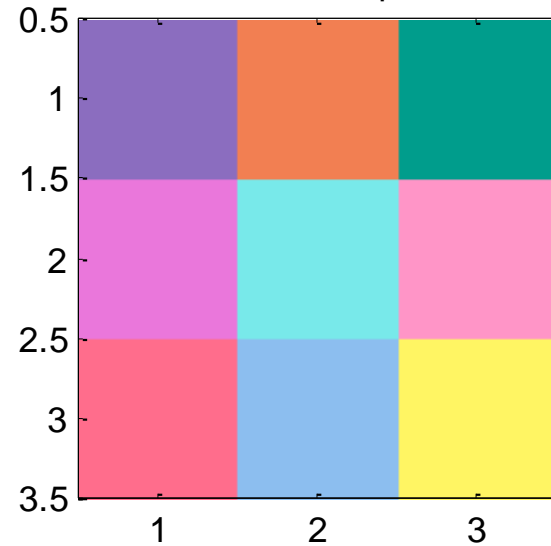
- Matlab used to perform transformations
- Average RGB values transformed to XYZ using ICC profile
- XYZ transformed to Lab using library function
- Values compared to Reference target values



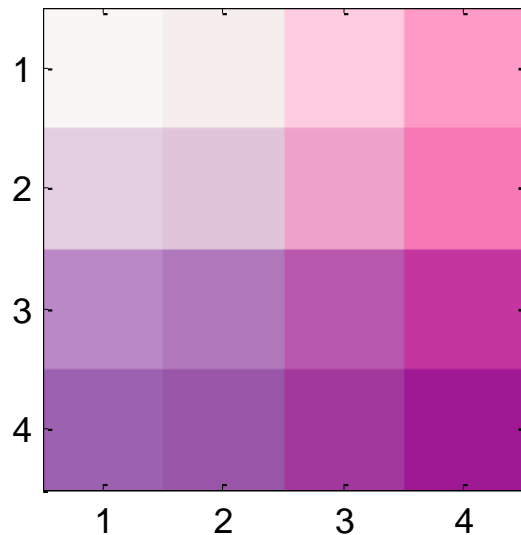
Reference Block 1 Spectral sRGB



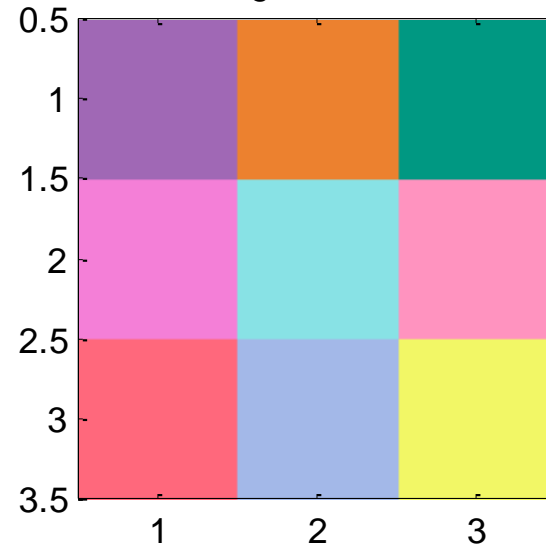
Reference Block 2 Spectral sRGB



AT2 Image Block 1 sRGB



AT2 Image Block 2 sRGB



- Scanner profile
  - general model, based on component specs (light, camera)
  - scanner-specific profile could improve accuracy
  - method used and sample spectra (regression, LUT, spectral)
- Illuminant specified for profile:
  - halogen, D50, E, other?
  - chromatic adaptation to D50 PCS
- Rendering Intent
  - perceptual (default) or absolute
- Intensity of clear/white patch (255, 250, other)
- Noise in target image
  - actual target noise, system noise
- Spectral calibration of target slide
  - measurement errors
  - target fading

- Recommendation
  - Use the initial testing to identify approaches that work best for creating a standardized viewing experience.
  - Make recommendation for profile generation method and apply it to each system. Provide software and assistance to do this.
  - Publish recommended approach along with results from the various systems.
  - Importantly, this will allow us to establish acceptable variation.
- Recommend against publishing existing system results
  - This could unfairly make some systems look bad.
  - Not likely to get participation by all.

**END**



KONICA MINOLTA

# **A Trial at Konica Minolta**

**Nov. 1, 2014**

**Shuji Ichiatani and Po-Chieh Hung**



# Summary

- 1) **We tested as a user (we are not a manufacturer)**
- 2) **We scan the slide with Fluorescence Microscope “Keyence BZ-X710”**
- 3) **We measure the SPD of light source**

## **We found:**

- **Slide image has a lot of dusts and bubbles**
- **The light source is not good in terms of color rendering**



# Keyence BZ-X710 Fluorescence Microscope



- 3 CCDs for transparent color image
- Dichroic filter for color (?)

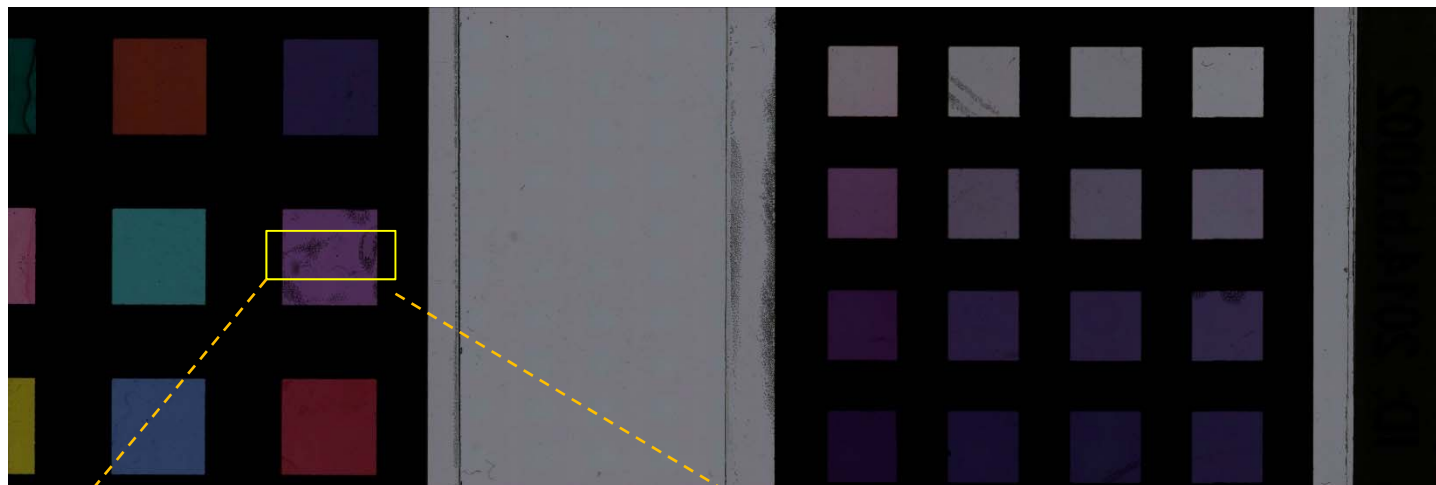


Copied from its catalogue  
<http://www.keyence.co.jp/frontend/documentdownload/documentdownload.do?fileKbn=3>



# Image Obtained

- Issue: the viewing (capturing) area is limited.



- Bubbles and dusts

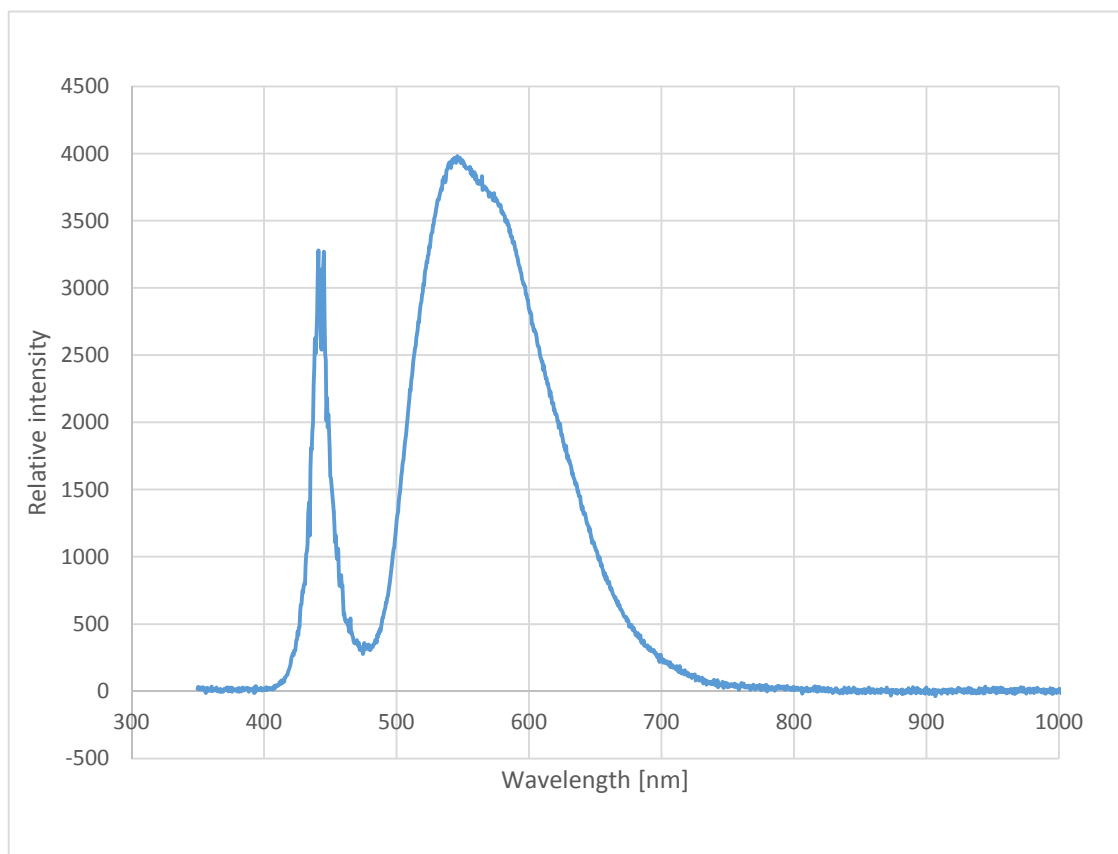


➡ **Geometry issue?**  
**Because it is designed for fluorescence?**



# SPD of Illumination

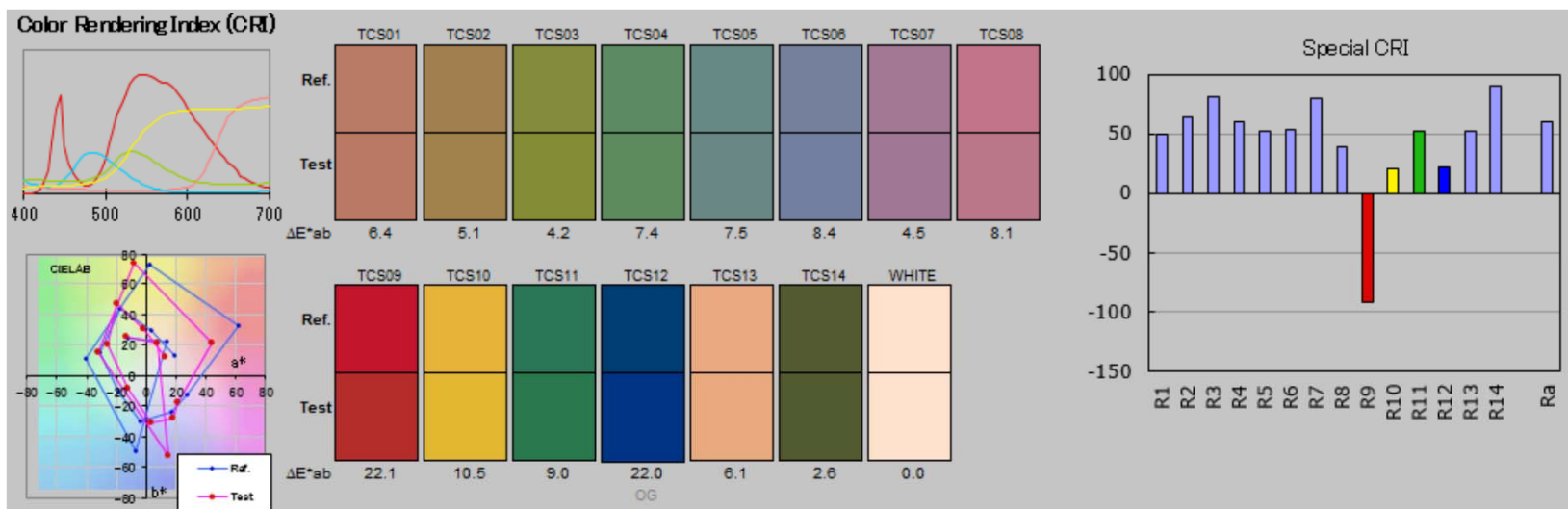
- White LED (blue-pumped phosphor)





# Color Characteristics of Illumination

- CCT: 4527K
- Duv: 0.032
- CRI Ra: 60
- R(9-12): 1
- R9: -92



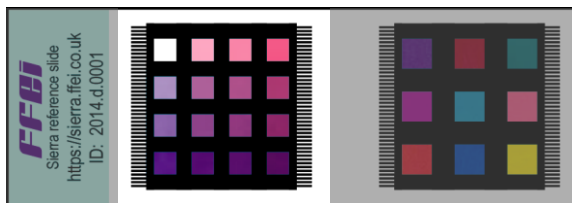
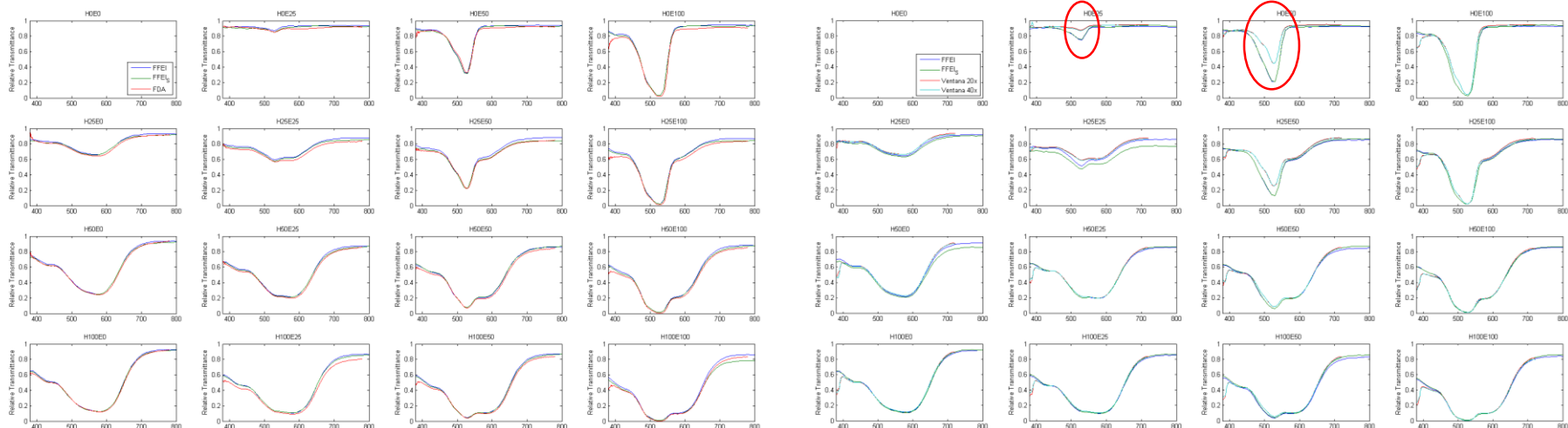
By "CQS 7.5a.xls" courtesy of Dr. Y. Ohno, NIST.

➡ **Not good for optical observations**  
 -> **It is likely to require non-linear color matrix**

# Measurements comparison and FFEI, Leeds and FDA image analysis results

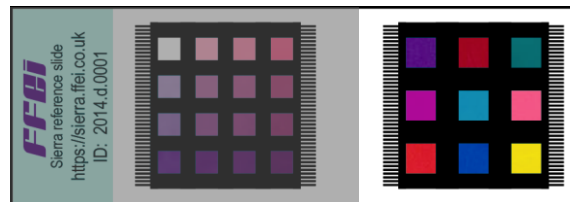
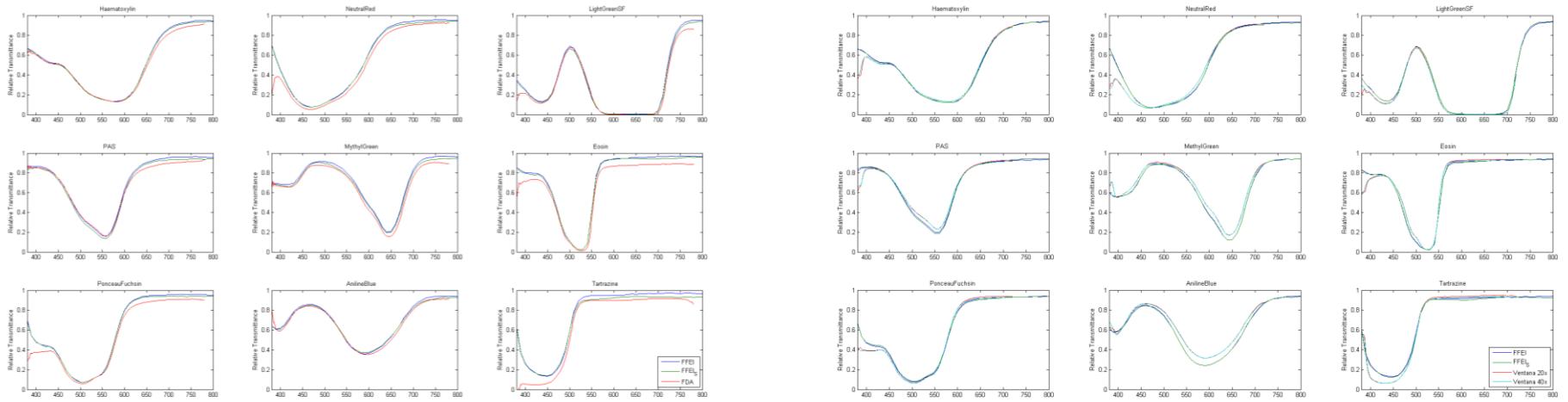
**1<sup>st</sup> November 2014**

# Slide measurements (FFEI / FDA / Ventana)



Possible problem with fading of light Eosin patches may indicate problem with application of anti-fading agent (DABCO)

# Slide measurements (FFEI / FDA / Ventana)



# FFEI calibration method

- Uses a model of the scanner based on ‘generic’ sensor sensitivities
- Training data was large set of measurements of stained pathology slides
- Measurements of Sierra calibration assessment slide were specifically not included in the training set
- Calibration process slightly biased in favour of H&E stains

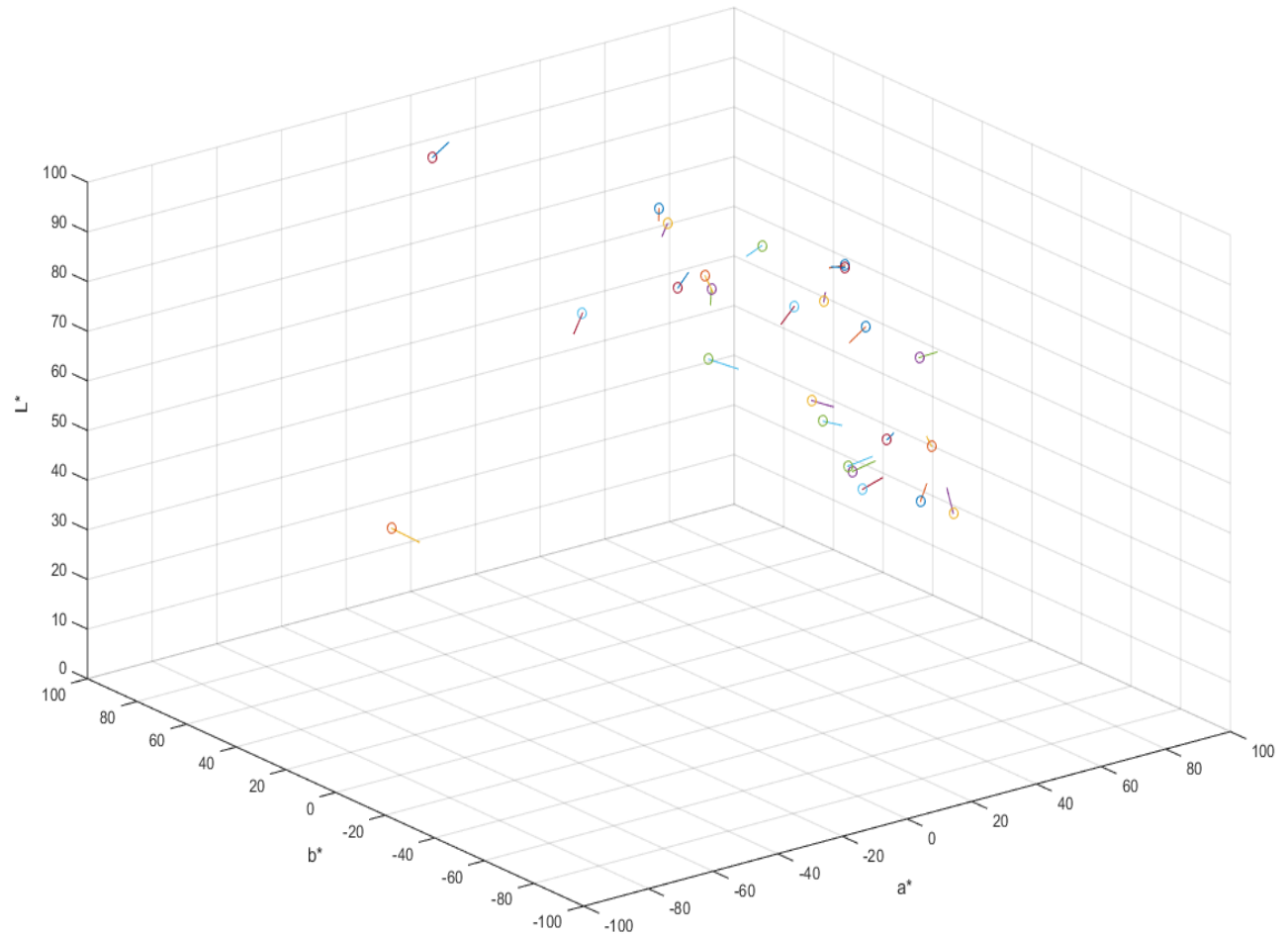
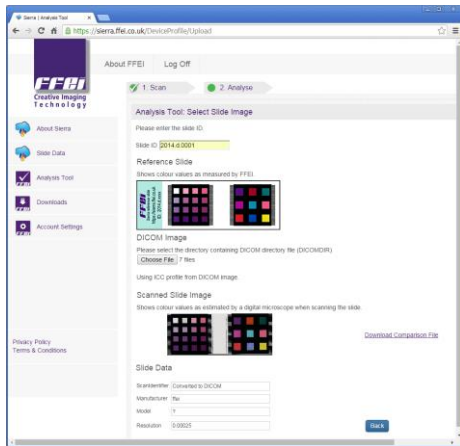
***Note that these results were from FFEI’s development platform and should not be considered to be representative of any OEM product***



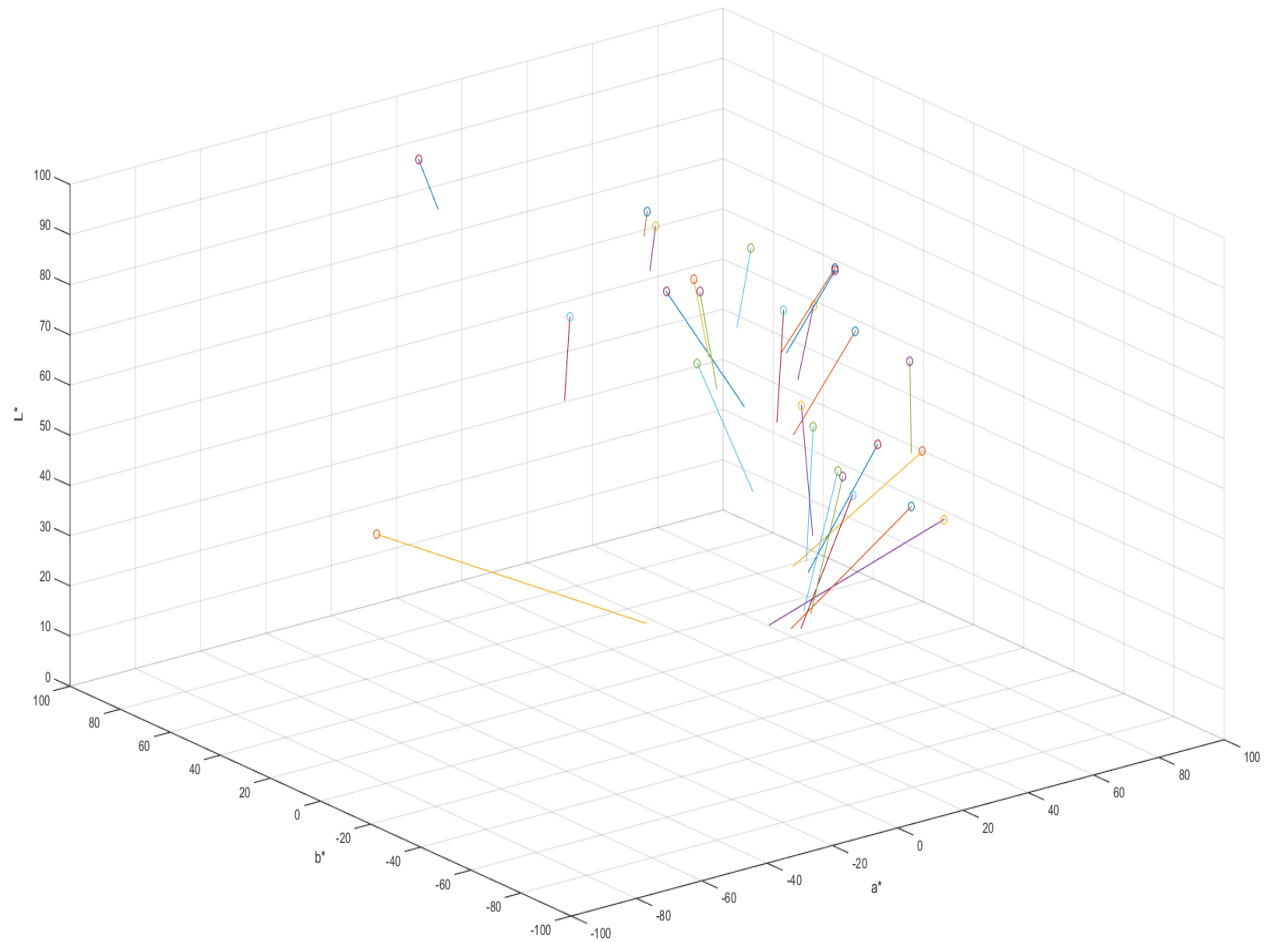
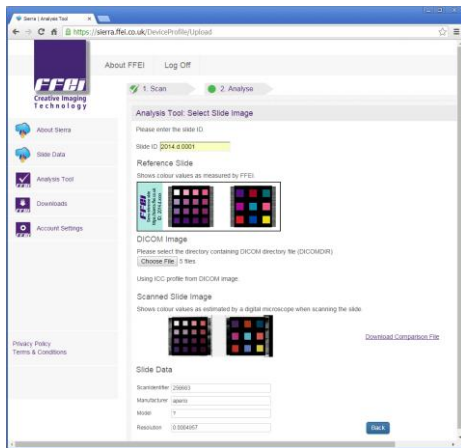
# Calibration assessment method

- **Step 1: calibrate scanner and create ICC Profile**
  - **Step 2: scan slide and create image of slide in DICOM format**
  - **Step 3: use the Sierra calibration assessment tool**  
— <https://sierra.ffei.co.uk>
  - **Step 4: save the analysis file created on the web site**
- 
- **Demonstration**

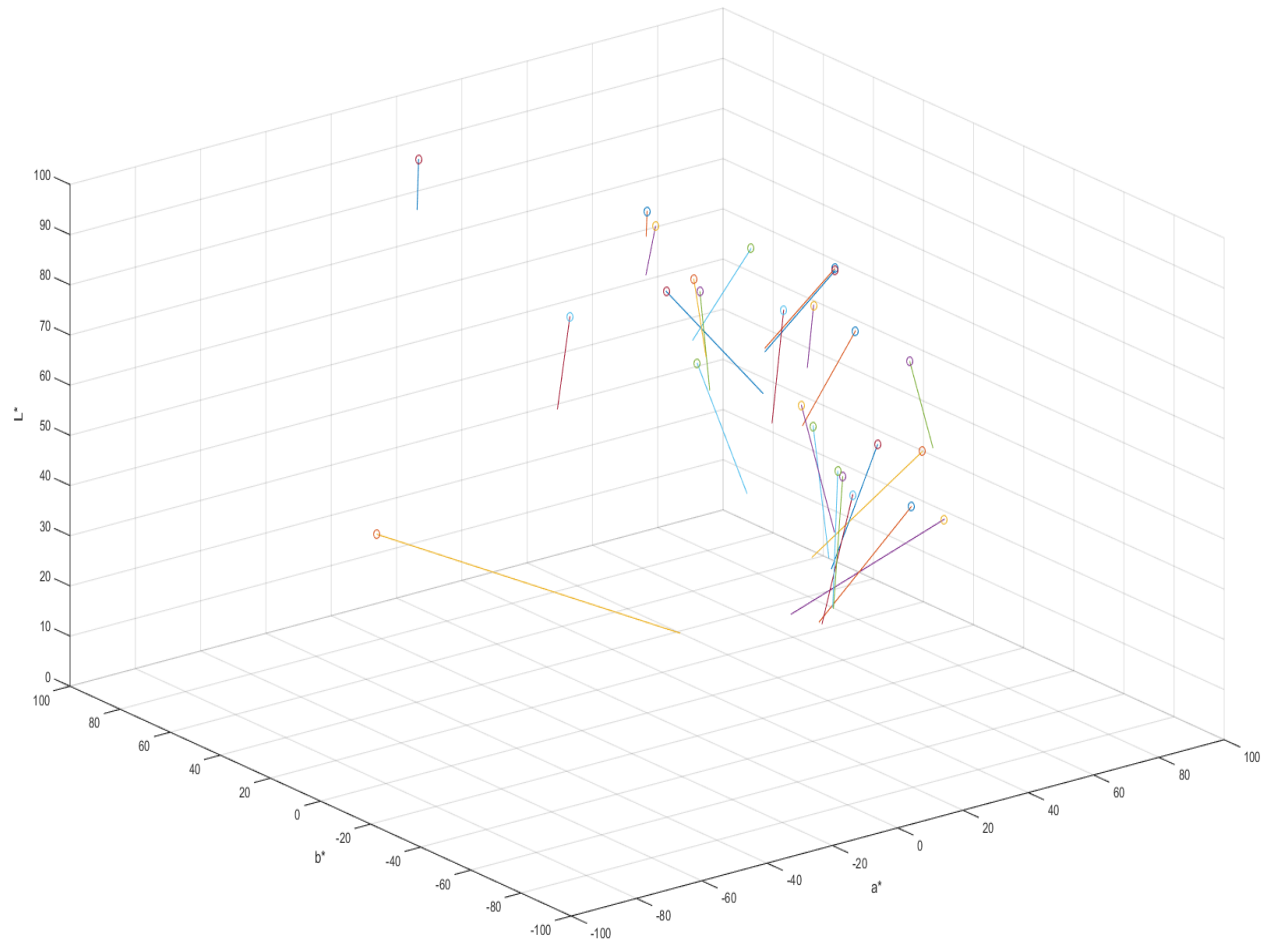
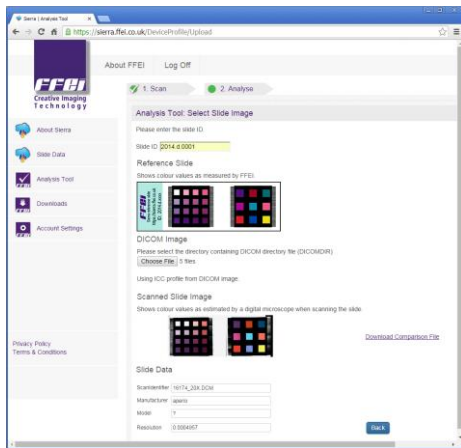
# Slide image colour accuracy (FFEI)



# Slide image colour accuracy (Leeds)



# Slide image colour accuracy (FDA)



# Explanation of the Aperio result

Older ScanScope models were not corrected to a specific viewing condition. The profile was ***designed to adjust the color according to customer feedback*** and the effect is very small relative to no correction at all. The profile intentionally makes no correction for the tonal response of the instrument.

The color profile for our most recent system (AT2) was developed with the goal of reproducing the viewing experience of a tungsten light microscope with daylight filter. The profile is then chromatically adapted to D50 connection space per ICC specifications.

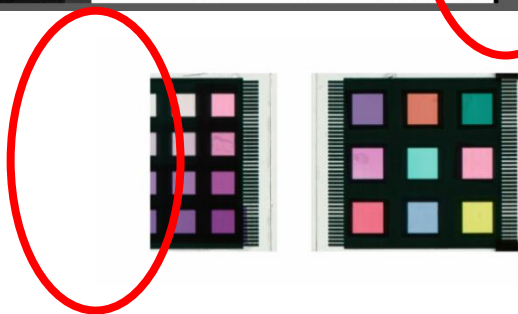
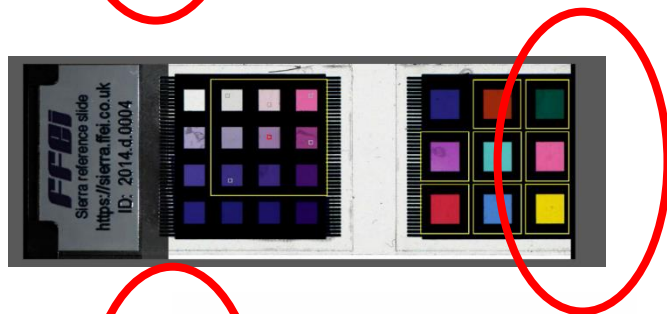
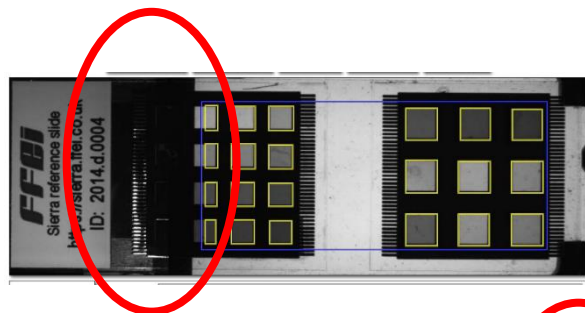
- *Note that if you look at the Leeds/FDA results, you will see that the major part of the difference is in L, as the lines point primarily in the vertical direction of the 3D graph. As I said, we intentionally left the tonal response alone, since this was the what customers preferred. The two systems have nearly the same result and are very consistent.*

*Allen Olson, Ph.D.*

*Senior Scientist, R&D Engineering and Analysis*

*Aperio ePathology*

# Problem with automatic scanning



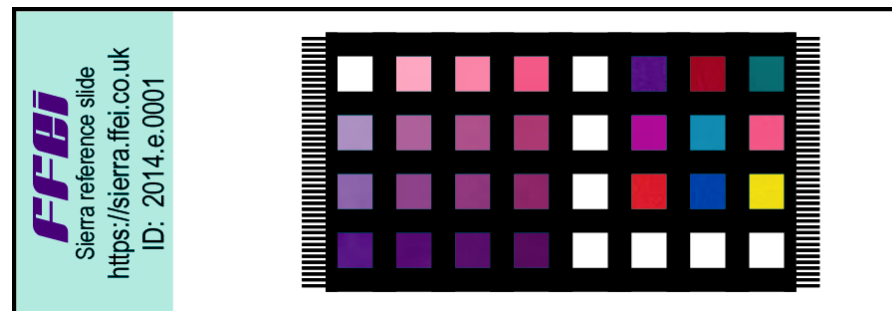
The current layout may limit the usefulness of the slide:

*We have a bit of an issue when we scan the slide in automatic mode: Some of the patches were not detected by the scanner. It could be because the luminance of the patch is low and we have a black background. Not the whole area of the patches is detected by the scanner. We found that the label on the calibration slide is a bit slimmer than the usual slide labels we have in here.*

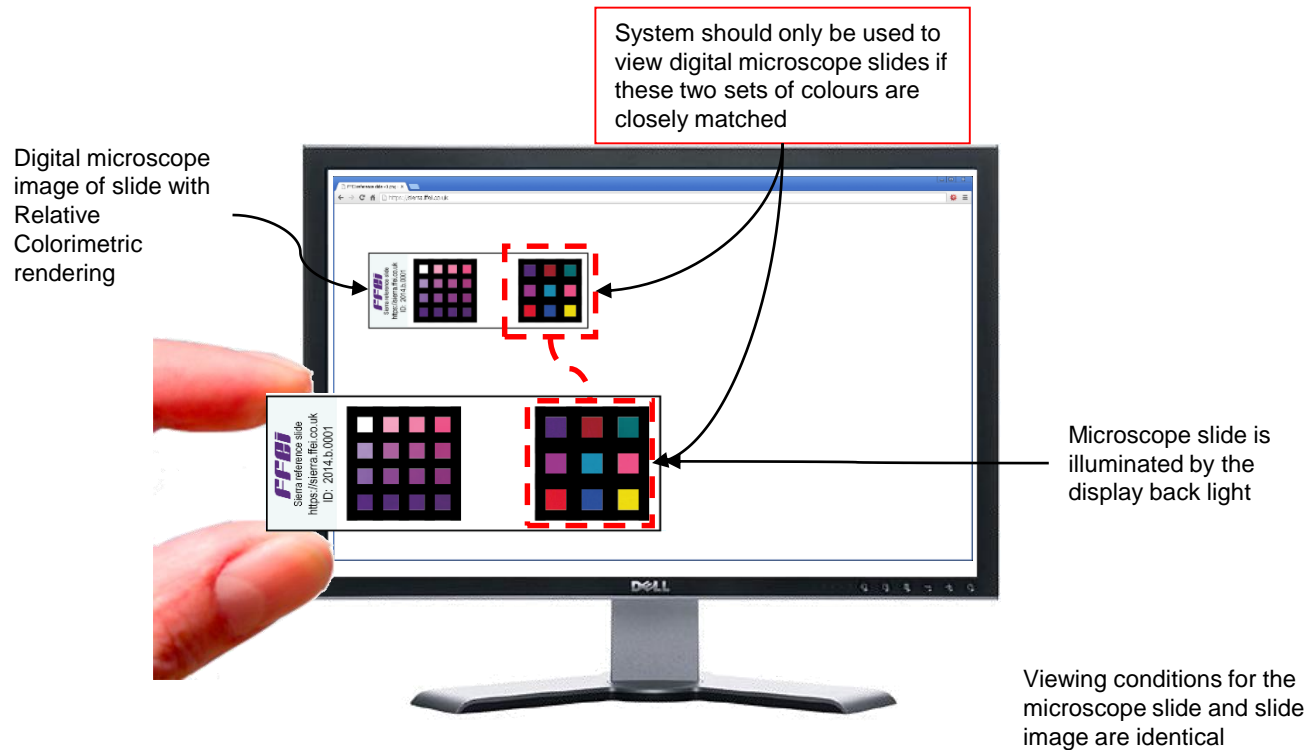
*Kindly refer to the attached images.*

*Thanks.  
-Pinky*

Revised layout of slide currently being tested

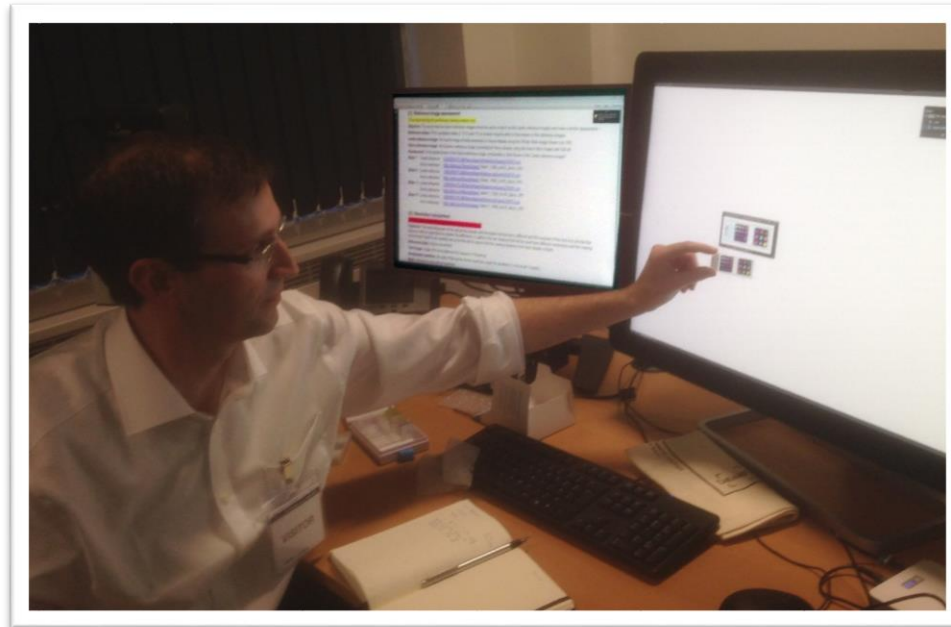


# Visual assessment



Based on a method developed and promoted by Yukako Yagi and Pinky Bautista

# Pathologist test as part of project Sierra



Initial feedback indicates that this method is easy to use and provides immediate feedback as to whether the system is calibrated to an acceptable level



# Questions?

Color Translator tool – possible use in calibration assessment      Marti Maria

- **No presentation as Marti described the Color Translator product that uses LittleCMS colour engine which could be useful for some kinds of colour assessment**  
— see <http://www.littlecms.com/professional/>

# Sierra accelerated fading test and extended slide

**1<sup>st</sup> November 2014**

# Accelerated fading test protocol

**Sierra Calibration Assessment Slides exposed to light for different time periods over the course of six weeks in three phases:**

- **Phase 1:** all slides scanned daily but otherwise kept out of any light sources in order to establish a baseline for one week
- **Phase 2:** individual slide protocol for low light light exposure observed for 2 weeks
- **Phase 3:** individual slide protocol for high level of light exposure observed for four weeks

## **Light source details**

- Low light: Diall Halogen Eco bulb (24W, ~205 lumen)
- High light: Diall Halogen Eco Bulb (37W, ~370 lumen)
- Source deployed 250 mm above the slides

# Accelerated fading test



## **Slide 2014.d.0007 Negative Scan Control Slide**

Not exposed to any light after the first week until the end of the experiment (total exposure of 8 measurement scans)



## **Slide 2014.d.0008 Positive Scan Control Slide**

exposed to light only during the scan process (daily measurement scans)



## **Slide 2014.d.0009 Half Exposure**

exposed to an additional light source for four hours per day (daily measurement scans)



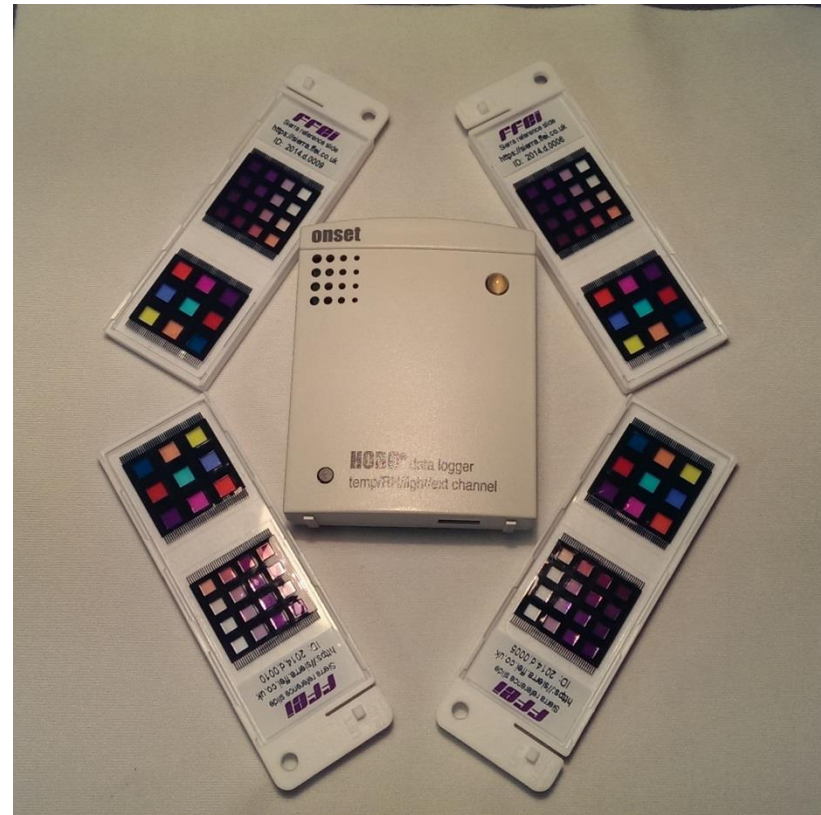
## **Slide 2014.d.0010 Full Exposure**

exposed to an additional light source at all times (daily measurement scans)

# Accelerated fading experimental setup



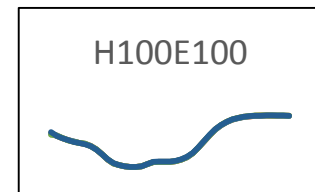
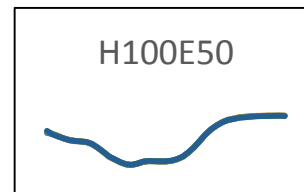
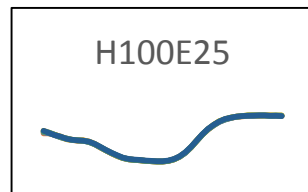
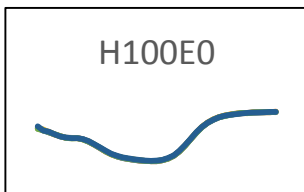
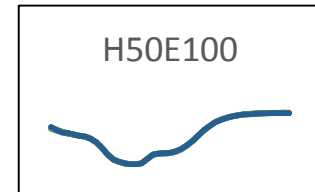
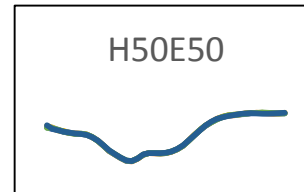
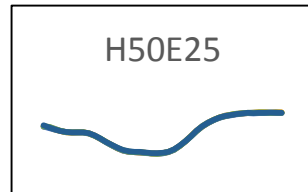
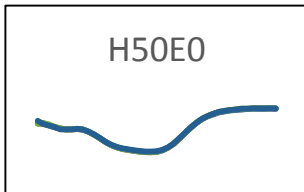
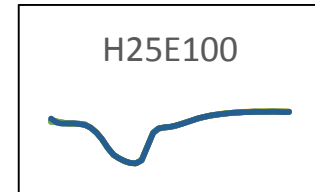
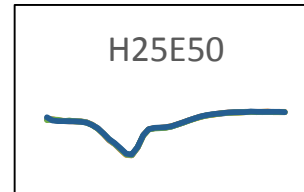
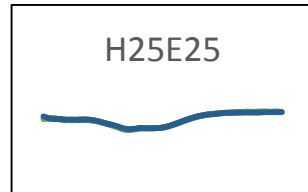
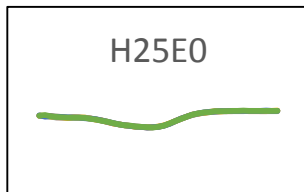
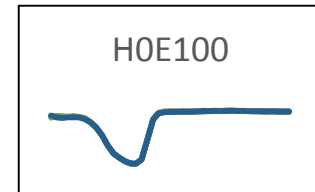
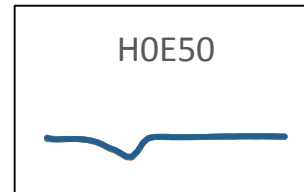
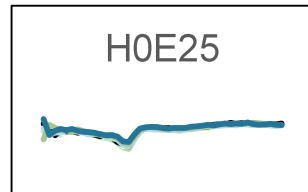
Distance from lamp to slide was approximately 250 mm



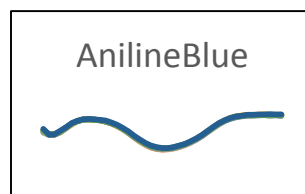
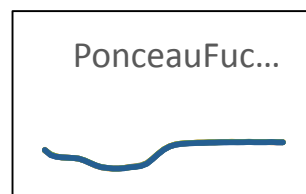
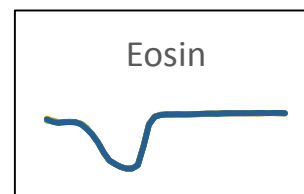
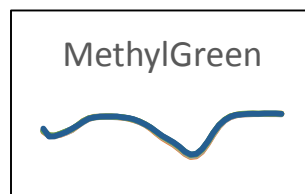
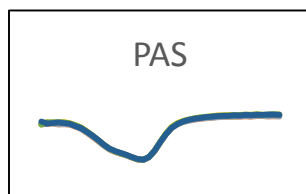
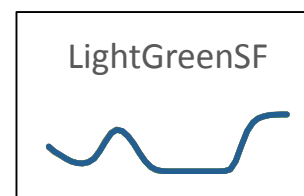
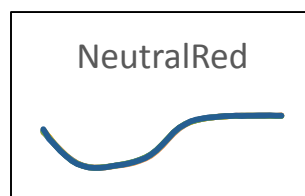
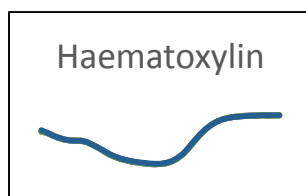
Data logger records light level, temperature and humidity

# Slide 2014.d.0007 Negative Scan Control Slide

**Initial  
analysis**



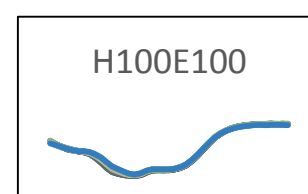
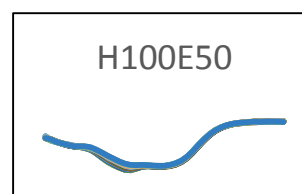
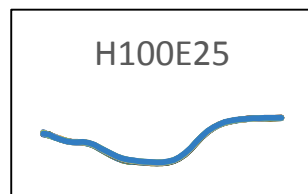
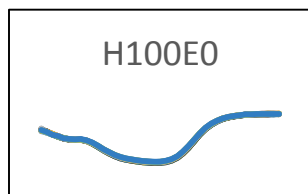
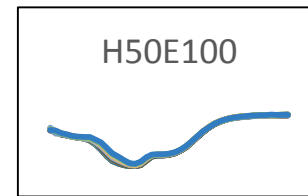
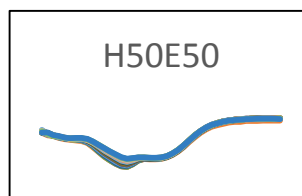
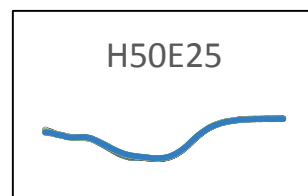
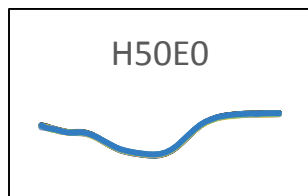
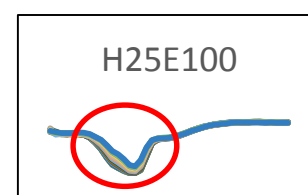
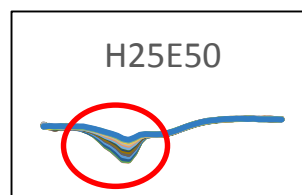
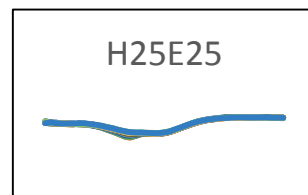
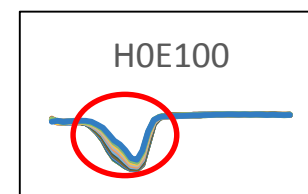
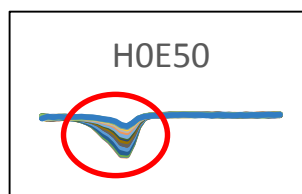
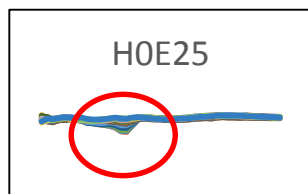
# Slide 2014.d.0007 Negative Scan Control Slide



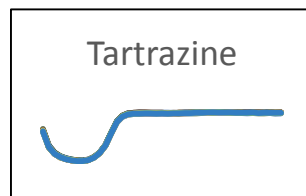
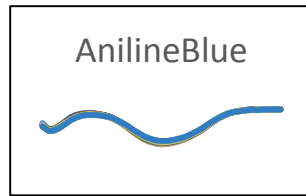
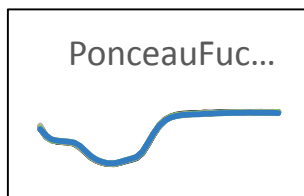
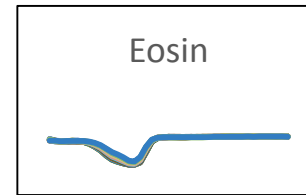
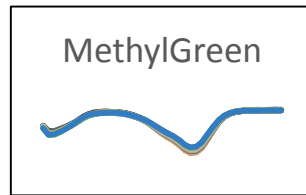
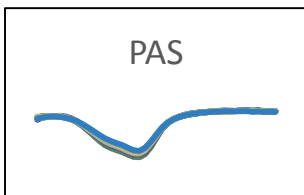
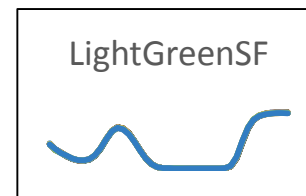
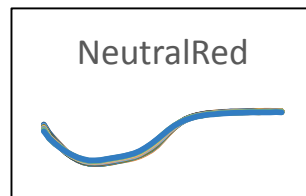
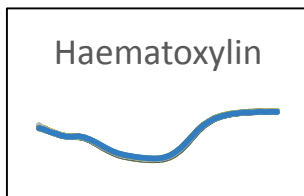


# Slide 2014.d.0010 Full Exposure

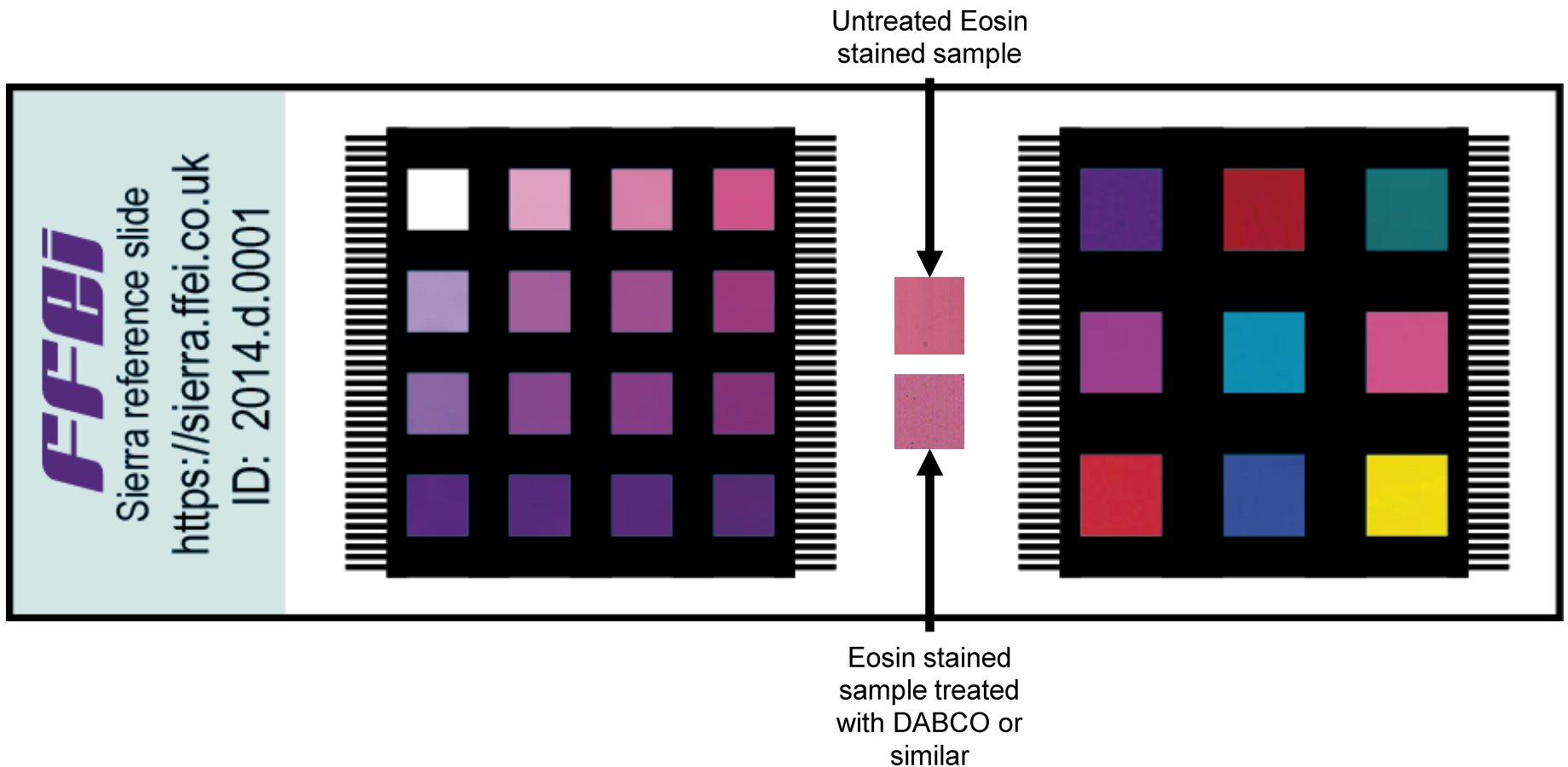
Eosin lightfastness can be improved by slight modification to our manufacturing process



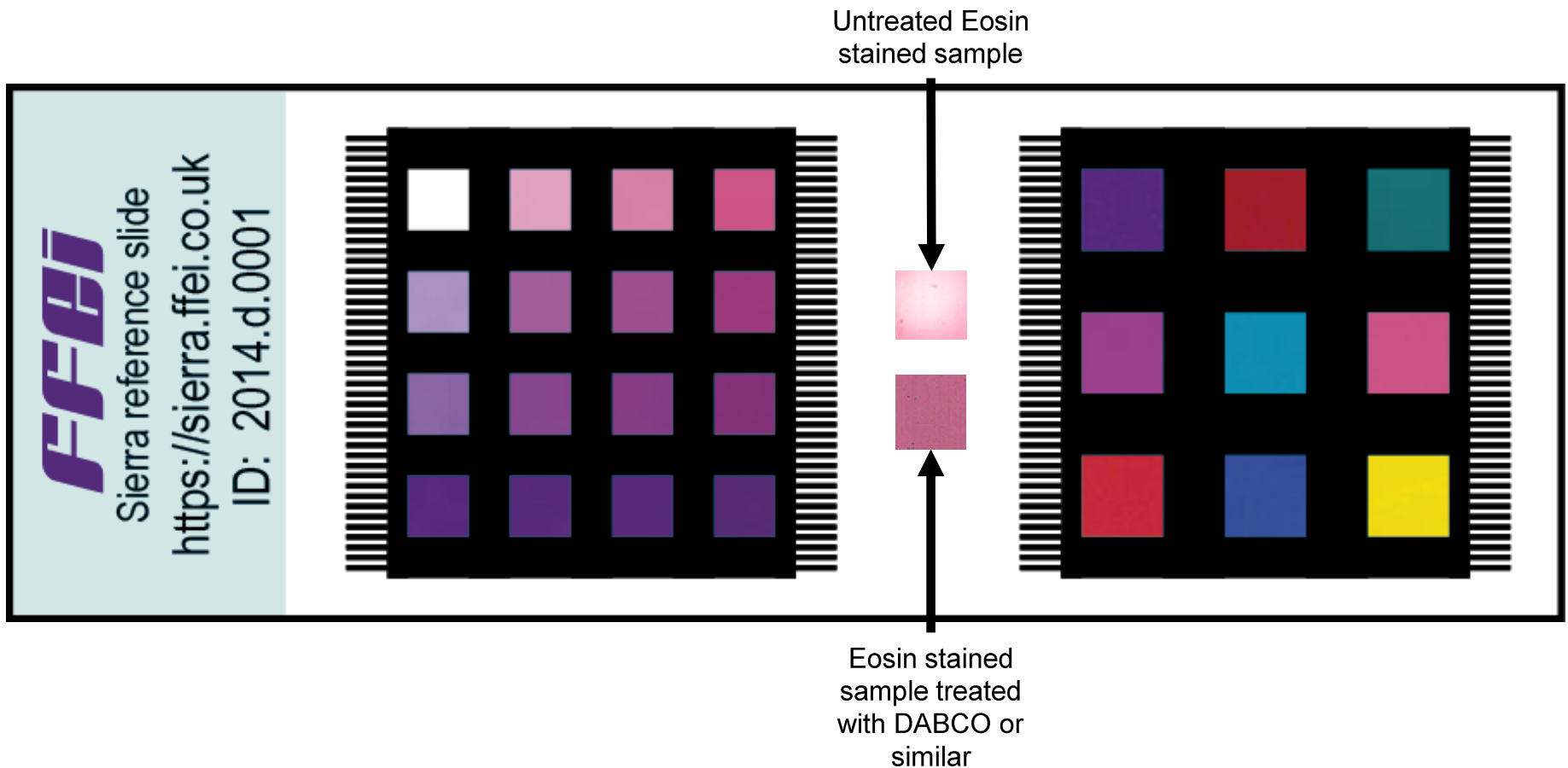
# Slide 2014.d.0010 Full Exposure



# Light exposure control patches



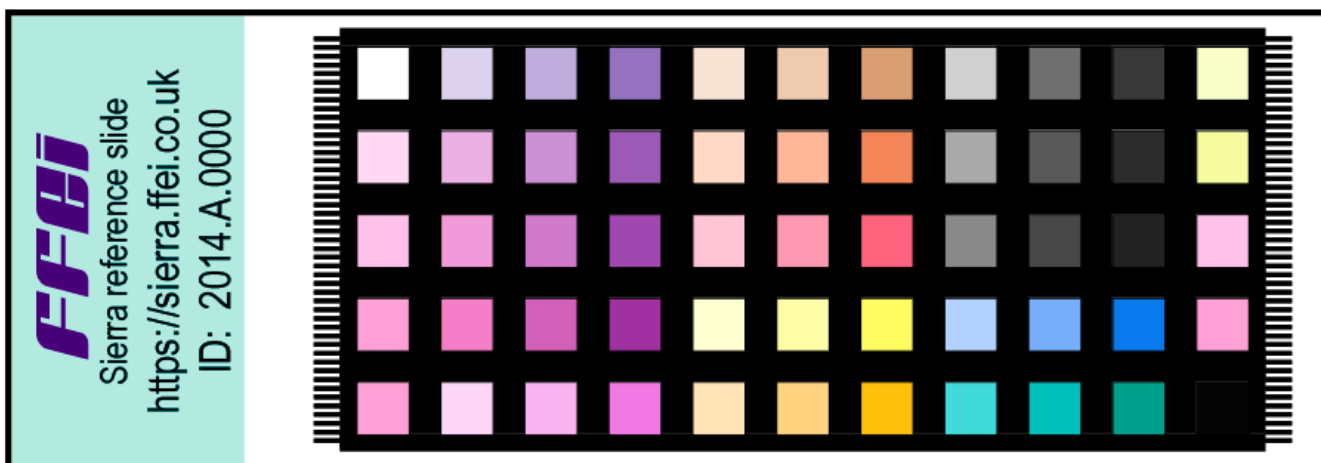
# Light exposure control patches



# Sierra Calibration Slide

- As part of the Sierra project FFEI plans to create a small number of prototypes of a slide that could be used to calibrate digital microscopes
- Slide will have a set of 55 patches based on the set of spectra we use to build our ICC Profiles for our system
- The uniformity of biopolymer staining has been improved
- The slide will incorporate neutral patches and a fading assessment patch
- ***We currently have no plans to manufacture this slide but are in the process of identifying possible partners***

# Sierra Calibration Slide



H0E0	H25E0	H50E0	H100E0	DAB_25	DAB_50	DAB	N1	N4	N7	PicricAcid_50
H0E25	H25E25	H50E25	H100E25	NeutralRed_25	NeutralRed_50	NeutralRed	N2	N5	N8	PicricAcid
H0E50	H25E50	H50E50	H100E50	CrystalScarlet_25	CrystalScarlet_50	CrystalScarlet	N3	N6	N9	Elastin_50
H0E100	H25E100	H50E100	H100E100	Tartrazine_25	Tartrazine_50	Tartrazine	AnilineBlue_25	AnilineBlue_50	AnilineBlue	Elastin
EosinControl	PAS_25	PAS_50	PAS	OrangeG_25	OrangeG_50	OrangeG	LightGreenSF_25	LightGreenSF_50	LightGreenSF	DarkPatch


Eosin 25
Eosin 50
Eosin 100

Haematoxylin 25
Haematoxylin 50
Haematoxylin 100

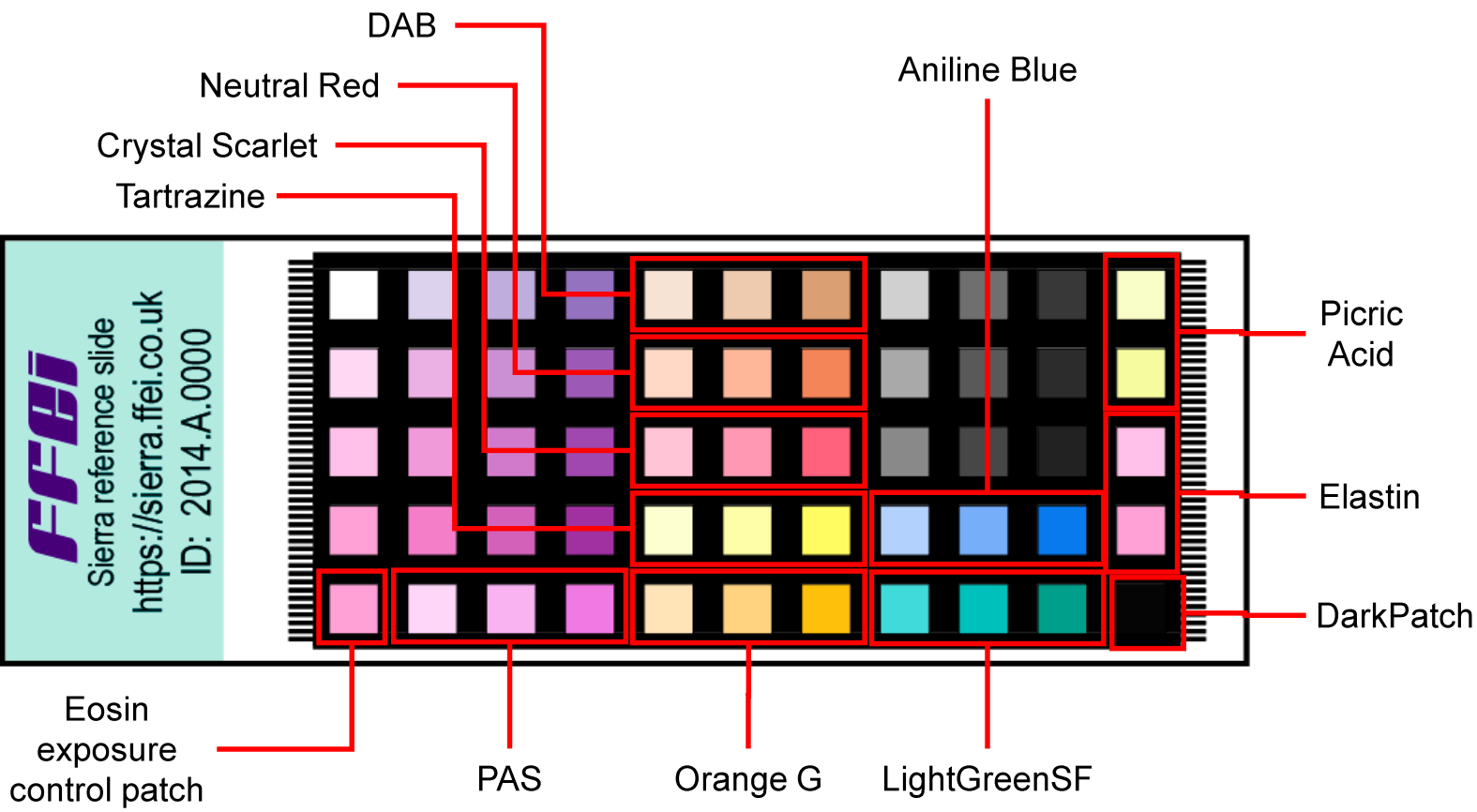
Clear
0.2
0.4

0.2	0.8	1.4
-----	-----	-----

**FFCI**  
Sierra reference slide  
<https://sierra.ffe.co.uk>  
ID: 2014.A.0000



The image shows a Sierra reference slide with a grid of color patches. Two red boxes highlight specific areas: one on the left side of the grid containing various shades of pink and purple, and another on the right side containing various shades of gray. Red lines connect these boxes to the corresponding color calibration charts shown above.







# Framework for Multispectral Imaging Application to digital pathology

Masahiro Yamaguchi, Tokyo Institute of Technology

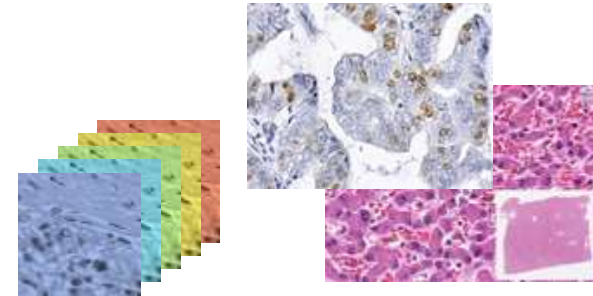
Bas Hulsken, Phillips

Max Derhak, Onyx Graphics Inc.

# Multispectral imaging in pathology

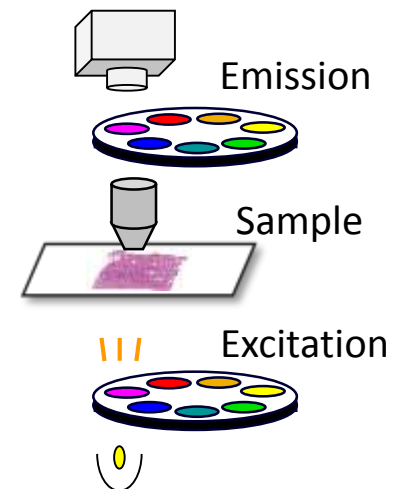
- Brightfield

- HE stain, Special stains, Immunohistochemistry(IHC) stains
- Object detection, segmentation
- Color unmixing – Stain amount image
- Digital adjustment of staining strength
- Digital staining



- Fluorescence

- Simultaneous imaging of multiple markers
- Cross-talk, auto-fluorescence removal
- Combined brightfield and fluorescent images



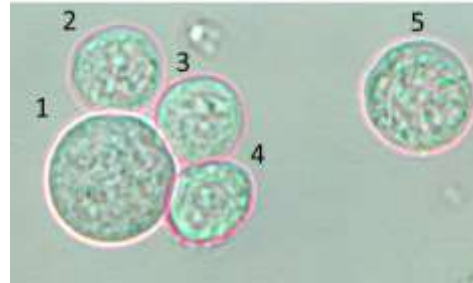
# Classification of B lymphocytes without staining

Spectral imaging:

Nonproducing cells (NP)



Ab producing cells A (PA)

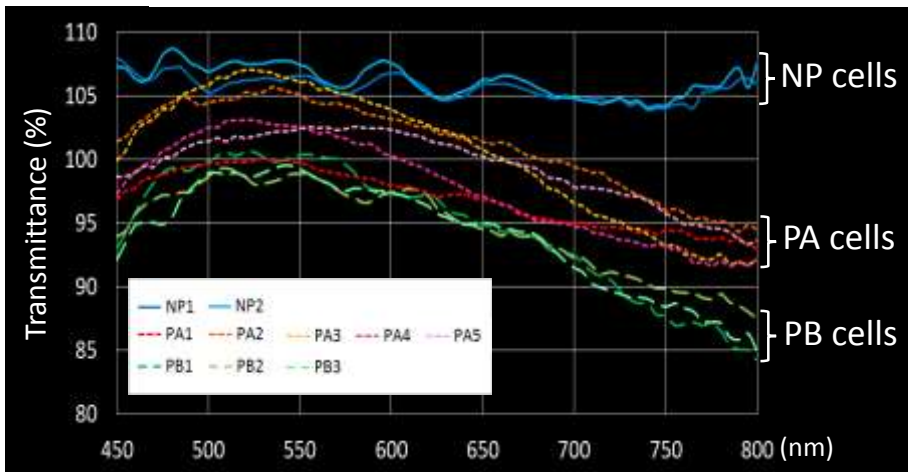


Ab producing cells B (PB)



\*(Ab: antibody)

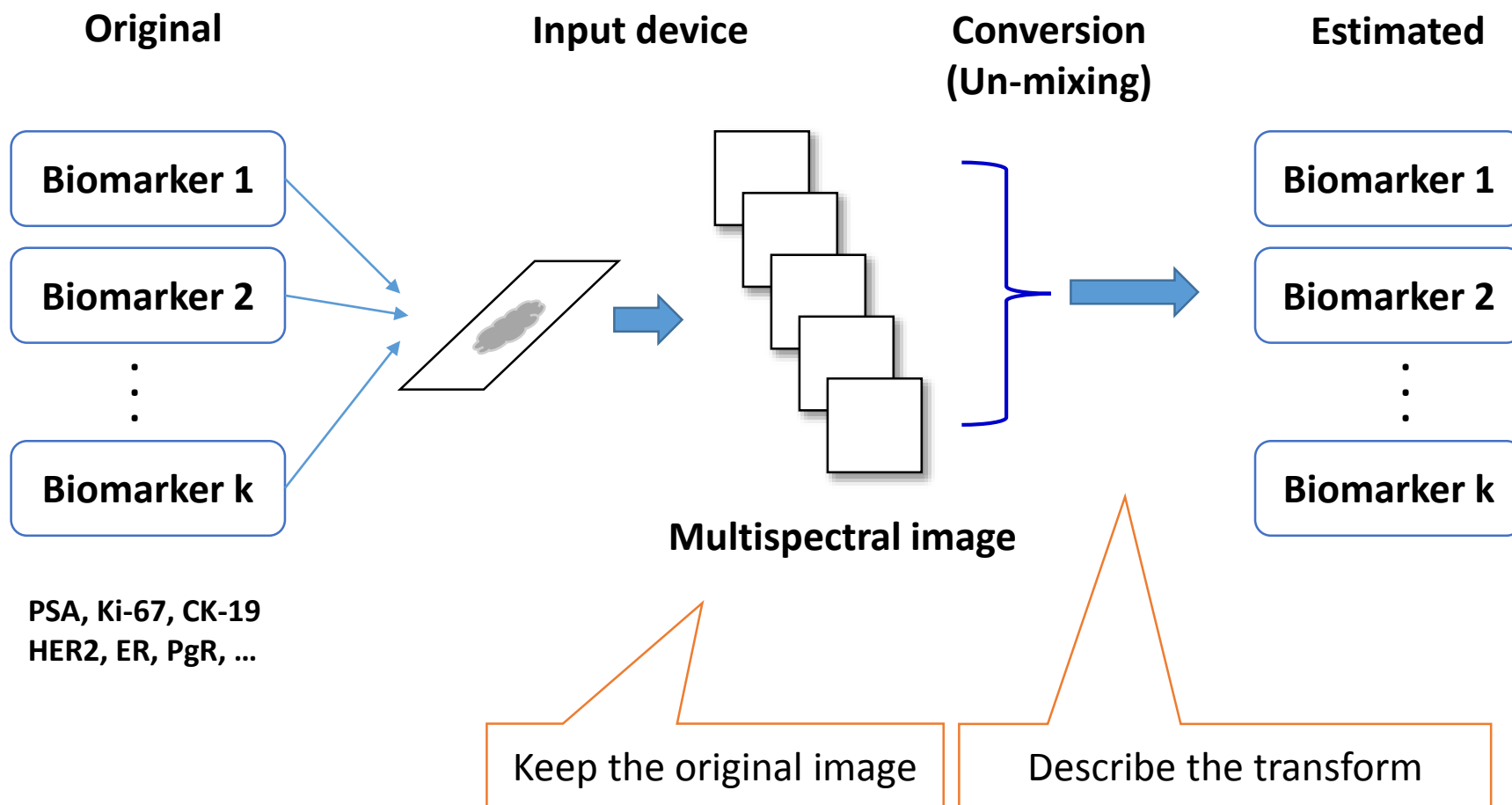
Spectral comparison



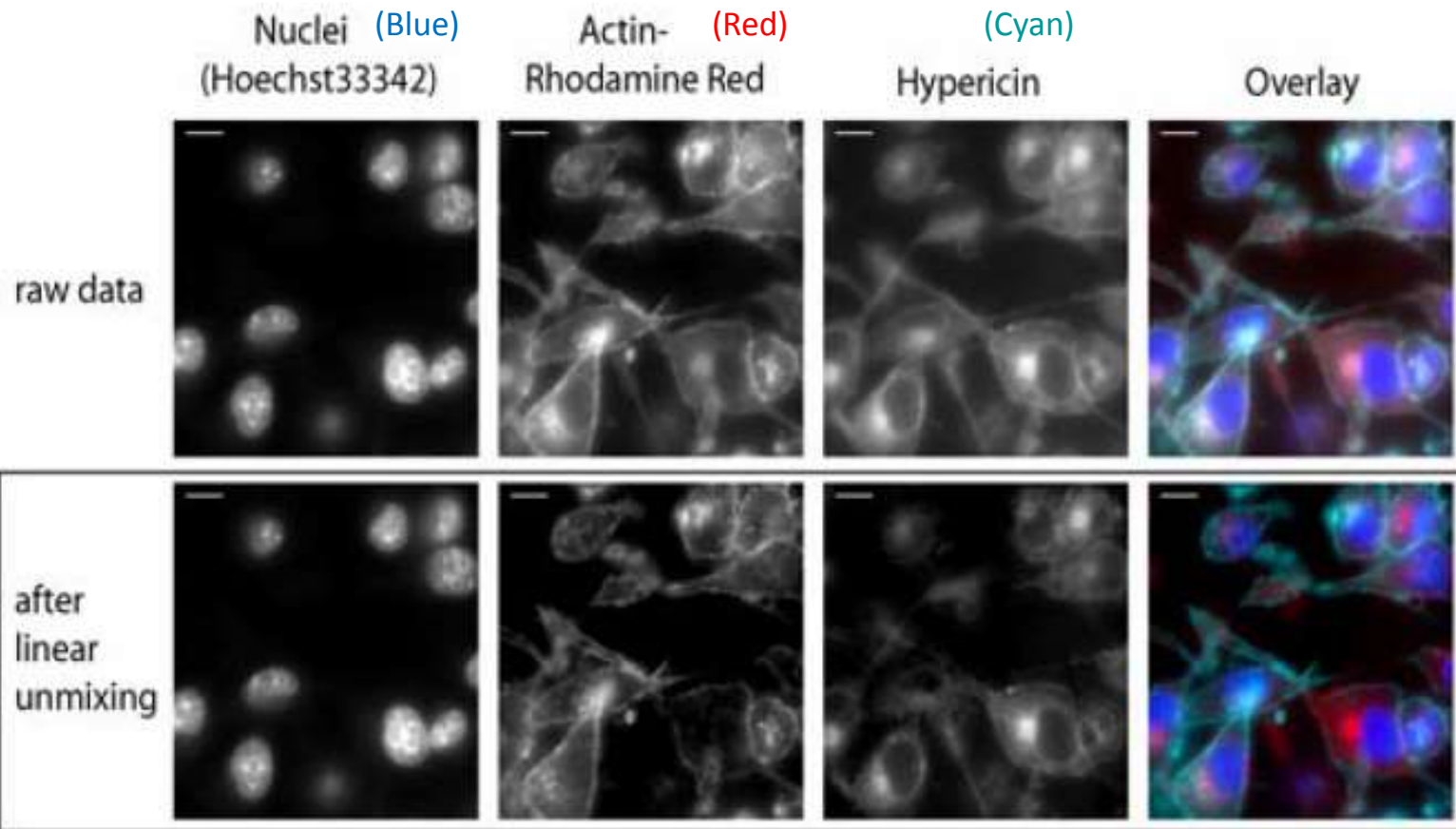
Live cell imaging and discrimination without any staining can be achieved using hyperspectral data.

Qualitative evaluation of live cells (eg. activation state of cells) is also possible.

# General model for multispectral un-mixing

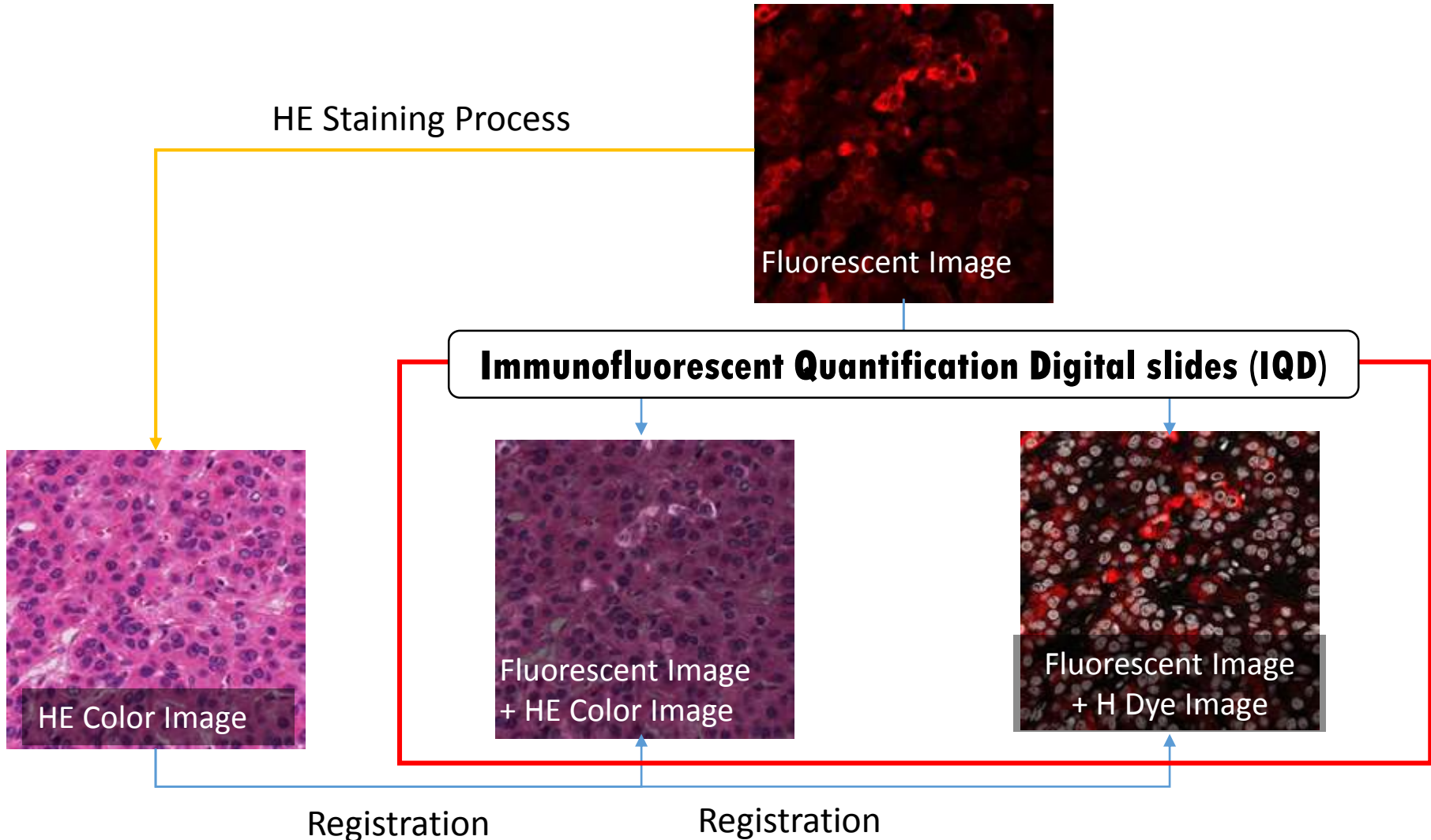


# Color unmixing for unwanted fluorescence removal



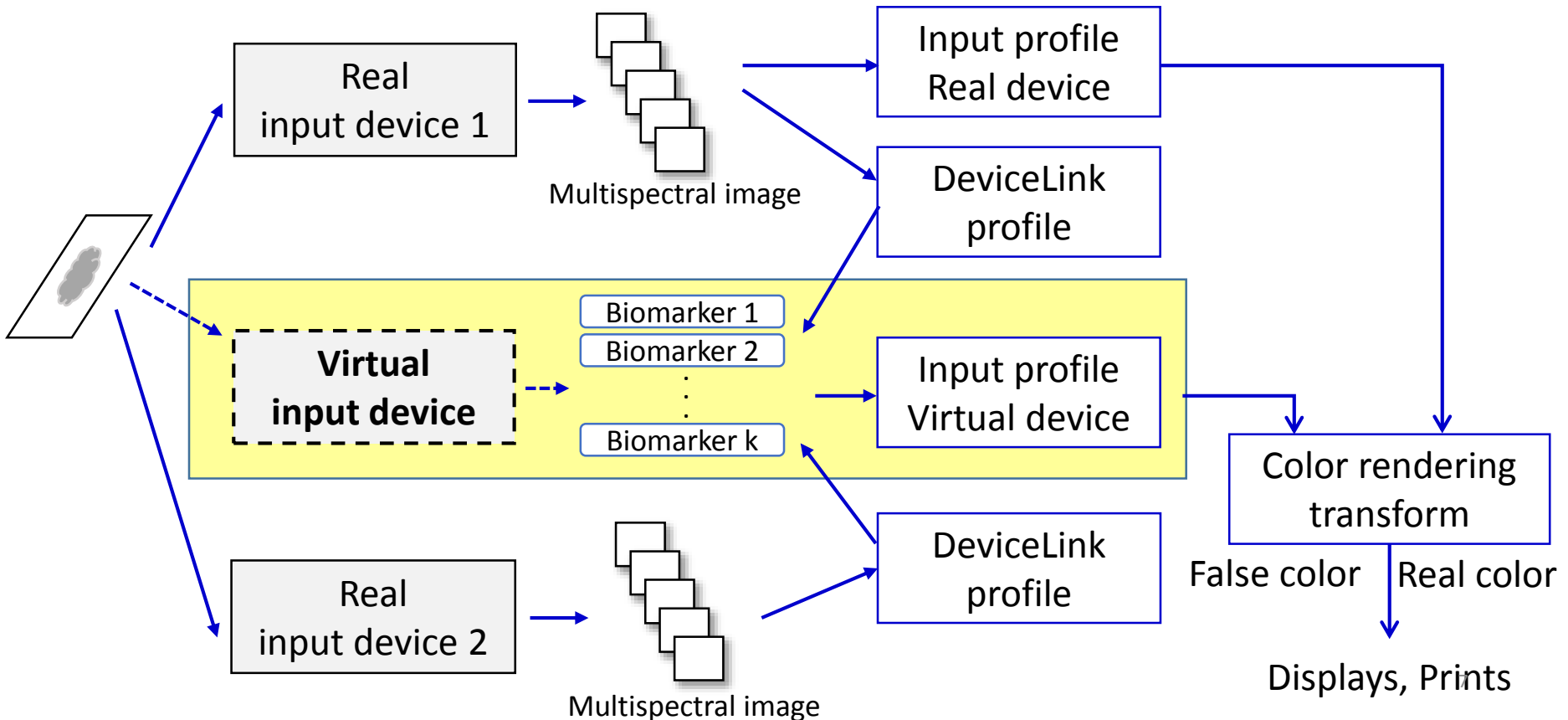
Modern Research and Educational Topics in Microscopy.  
A. Méndez-Vilas and J. Díaz (Eds.)

# Combination of Fluorescent and HE-stain



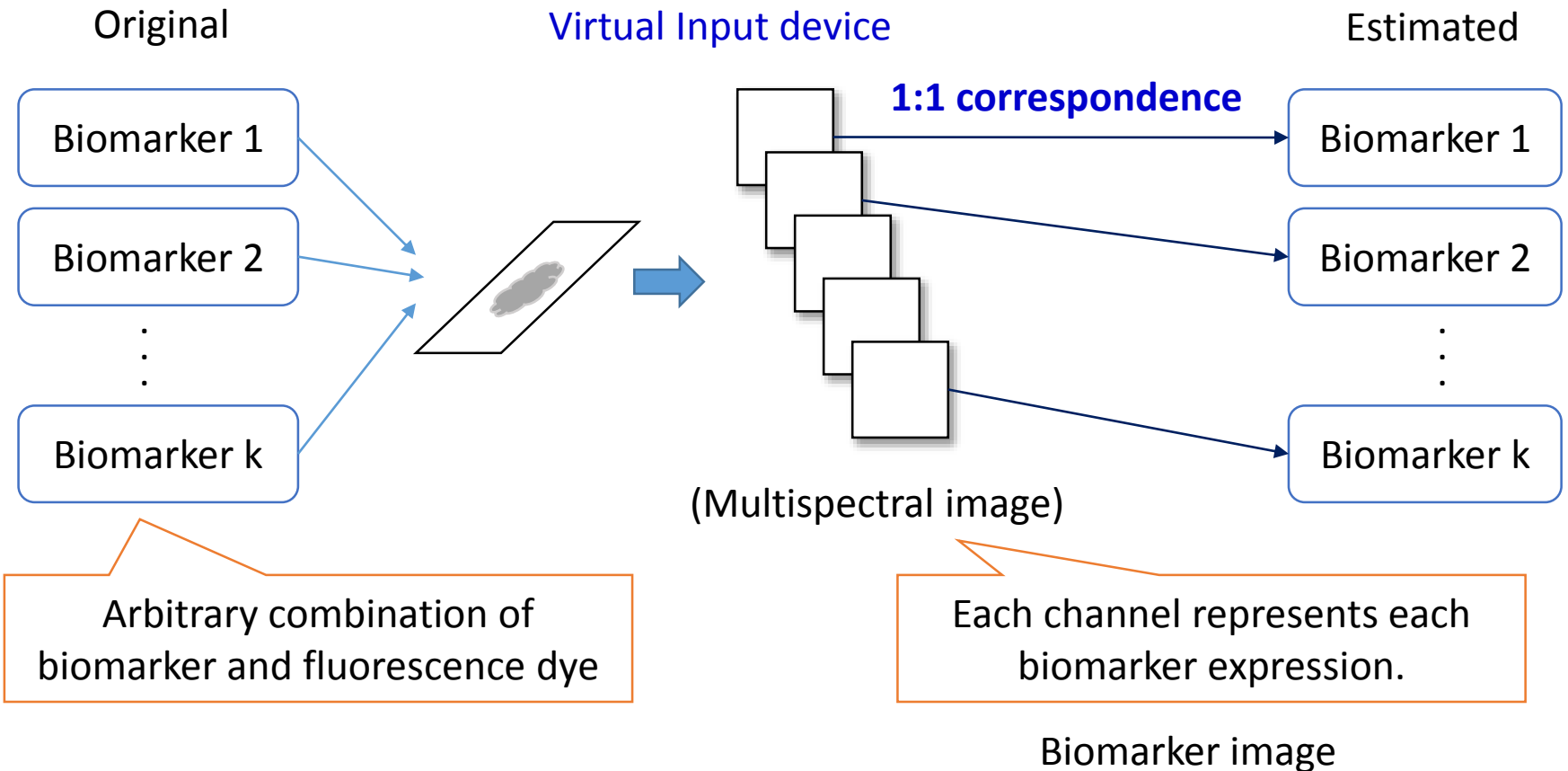
# Solution to color unmixing by ICC v4

- Consider a virtual input device that can directly capture un-mixed biomarker images
- Use DeviceLink profile



# Virtual 'ideal' input device

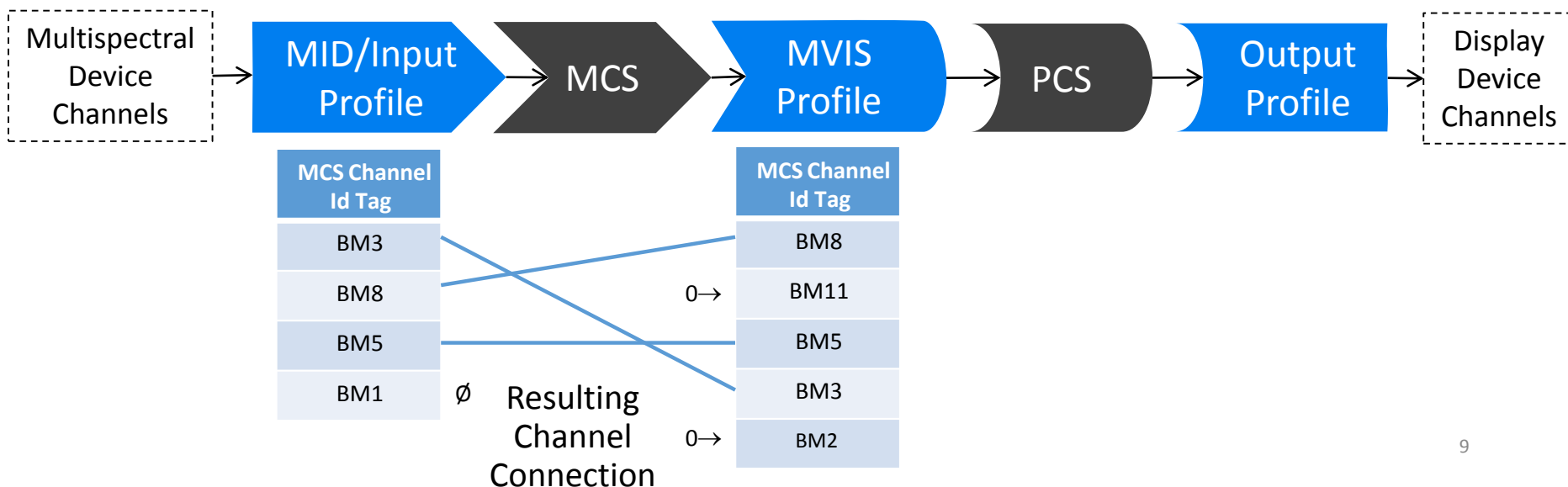
that can directly capture un-mixed biomarker images





# Solution to color unmixing by iccMAX

- Spectral profiles
- Multi-processing elements
- Application of “Material Connection Space” Profiles
  - MCS connection allowed between source biomarker Material Identification (MID) and destination biomarker Material Visualization (MVIS) profiles



# Current status

- Investigating the adoption of ICC v4 considering the upper compatibility in iccMax.
- Documentation for implementation to DICOM.
- Planning the test implementation of iccMax MCS approach for investigating its advantage and feasibility.
- \* We welcome contribution from anyone who can provide sample multispectral fluorescent images

# Dicomizing multi-spectral presentation states

**Bas Hulsken**

Digital Pathology Solutions

October 31, 2014

# Steps to take to get multi-spectral presentation states in DICOM

- 1) Discuss high-level DICOM implementation in WG26
- 2) WG26 to agree on high-level implementation
- 3) Send to WG6 for discussion
- 4) Write the full DICOM implementation

# Required DICOM extensions

## Simple removal of limitations:

- 1) Allow multiple ICC profiles in one DICOM object (**1 now**)
- 2) Allow devicelink ICC profiles (**only input profile now**)
- 3) Allow ICC profiles for  $n$  channels (**3 channels now**)

## More complex additional functionality

- 4) Add module describing multi ICC profile render pipeline
- 5) Define way to store multi-channel image data
- 6) Add module describing characteristics of channels (spectral, biomarker, etc.)

## 4) module describing multi ICC profile render pipeline

- Add module to describe chaining of ICC profiles in render pipeline
  - Can use softcopy presentation state either for inspiration, or by extending
  - Need to combine multiple images and/or channels
    - Option A: enhance presentation states to blend more than 2 images
      - Currently the advanced blending and display pipeline is both too complex and too limited. Too complex because ICC profiles contain all required functionality, too limited because maximum 3 data frames are supported(C7.6.23-1 of Part3).
    - Option B: add new module describing chaining, use (chained) ICC profiles for rendering pipeline

# 5) Define way to store multi-channel image data

- Option **A**, pack all channels in existing DICOM image IOD elements & modules
  - For  $n$  channel data: specify samples per pixel (0028,2000) to  $n$   
(**currently allowed, but meaning undefined**)
  - Define Photometric Interpretation (0028,0004) to *multi-spectral*  
(**currently allowed, but meaning undefined**)

Advantages: easy, no DICOM enhancements required

Disadvantages: legacy DICOM tools can not do anything with these images, no easy way to define subsampling, different bit depths (only via Photometric Interp.)

- Option **B**, channels in separate DICOM IOD's and combine with presentation states
  - For  $n$  channel data: use  $n$  monochrome images, or RGB + monochrome images  
(**currently allowed**)
  - Enhance or make new softcopy presentation state
    - Allow  $n$  referenced images

Advantages: legacy DICOM tools can handle the separate images (channels)

Disadvantages: requires new/enhanced presentation states. Cannot define correct rendering in Image IOD itself.

- Option **C**, channels in raw data, not image data, define all from scratch
  - Advantages: full flexibility
  - Disadvantages: lose all existing image handling tools/functionality

## 6) Add module describing characteristics of channels (spectral, biomarker, etc.)

- Add module that describes for the image the channel characteristics
  - Can use multi-spectral MR for inspiration
  - Multiple modes:
    - 1) Spectral characterization per channel (Illumination spectrum, detection spectrum, excitation spectrum)
    - 2) Biomarker concentration representation
- Module should describe derived channels?
  - ICC profile pipeline has intermediate results, which can have meaning (biomarker concentration)





# Material Connection Space Profiles in iccMAX

# Problem Statements

- **Provide ability to define how to process input channels for the purpose of deriving quantitative representations of individual material values**
- **Provide means of visualizing these quantitative representations**
- **Need to answer the question “What is it?” rather than “What does it look like?”**

# Possible Solution with Problems

- **One proposed approach to solving these problems would be to successively apply device link profiles**

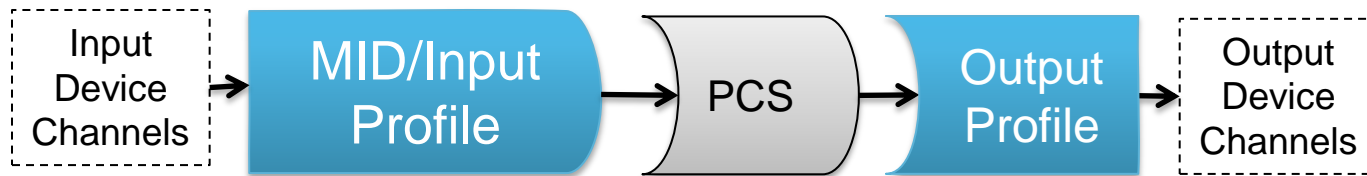


- **Intermediate device channels must exactly match**
- **This can quickly become unwieldy!**
  - Requires standardization of intermediate device channels for linking purposes
  - Explicit channel order is required
  - Cannot easily compensate for cases when only a subset of channels is needed/provided by one of the profiles

# Material Connection Spaces

- **An entirely new method of connecting profiles based on the concept of a Material Connection Space (MCS)**
  - Can conceptually be thought of as a material “device like” space with flexibility in channel routing
- **An MCS is completely separate from a PCS and therefore has no computational relationship to a PCS**
  - Relationships are completely provided by transforms in profile
    - AtoM0Tag, MToA0Tag, MToB0Tag, MToS0Tag - All based on multiProcessElementType
  - CMM performs no direct conversions between an MCS and a PCS
- **Like device channels, encoding of MCS channel data values is not explicitly defined**
  - Encoding of MCS channel values allowed to be defined by domain specific use cases
- **Rules for connecting profiles and routing channel data are well defined to be clearly implemented by a CMM**
- **MCS usage defined by new profile classes or extension to Input Class**

# MCS Connection Workflows



# Material Channel Connection Rules

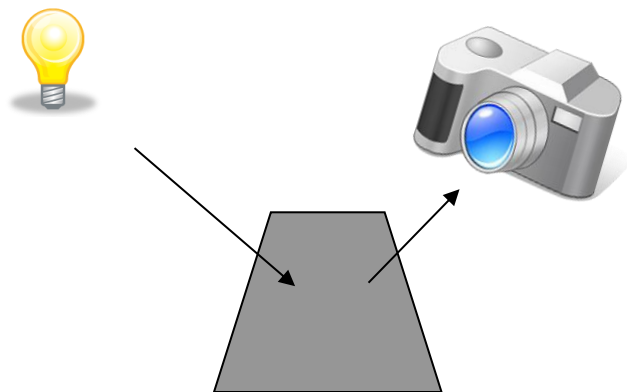
- **MCS connection allowed between source profiles with AToM0Tag [Input profiles / Material Identification profiles] and destination Material Visualization (MVIS) or Material Link (MLNK) profiles**
- **Apply MCS subset requirements**
  - If profile has MCS subset flag set then it's MCS channels need to be a proper subset of MCS channels in the connected profile
  - This ensures interoperability where channel requirements are needed
- **CMM simply passes channel data directly between source profile to destination profile for channels with same material channel identifications**
- **Ignore AToM0Tag MCS channels not present in MVIS/MLNK profile**
- **Use materialDefaultValuesTag values as input for MVIS/MLNK MCS channels not present in source MID profile**
  - This assumes independence of material channels
  - Use zero if materialDefaultValuesTag not present

# Example



# Hypothetical Scanner

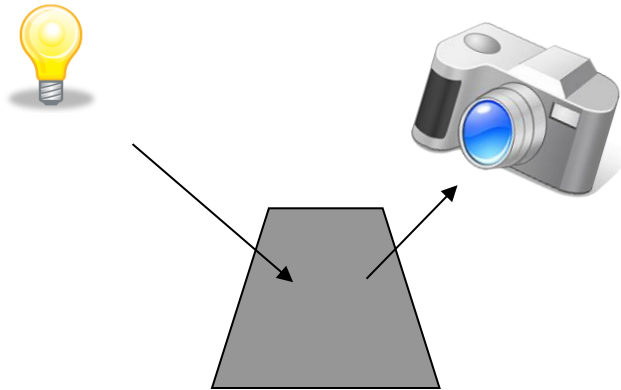
## Measurement #1



Results in Channels  
 $c_0, c_1, c_2$

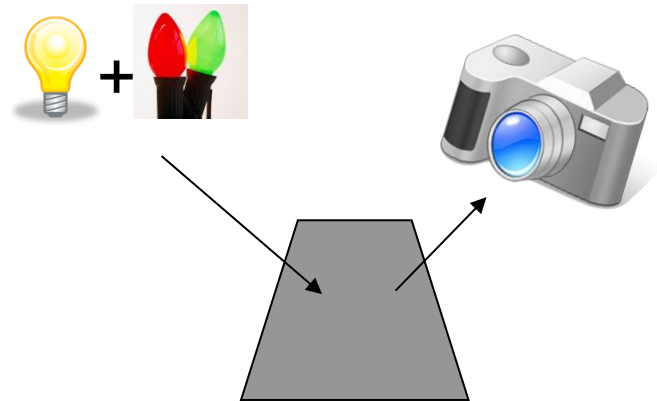
# Hypothetical Scanner

## Measurement #1



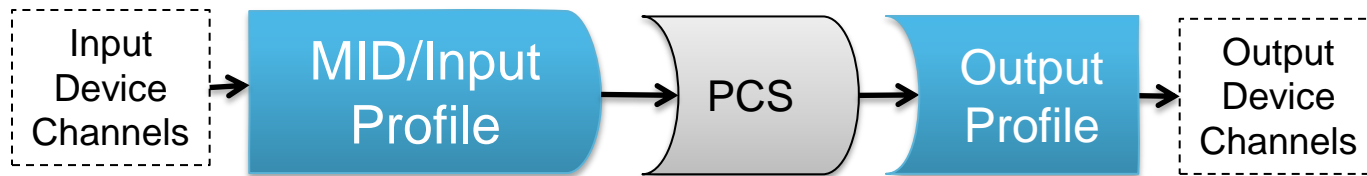
Results in Channels  
c0, c1, c2

## Measurement #2



Results in Channels  
c3, c4, c5

# MCS Connection Workflows



# Example MCS Input Profile

## Profile Header

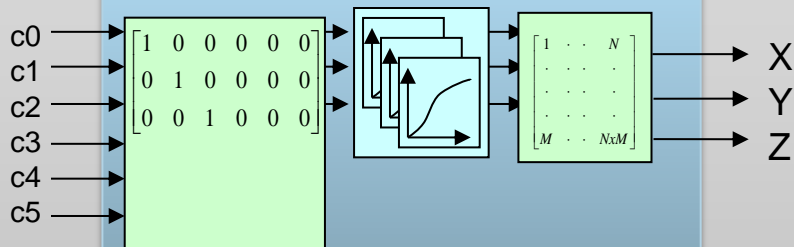
Profile Class: 'scnr'
Flags : MCS Sub-Set Requirements=false
Device: "nc0006"
PCS: XYZ
MCS: "mc0004"
Profile SubClass: TBD

## Material Type Array Tag

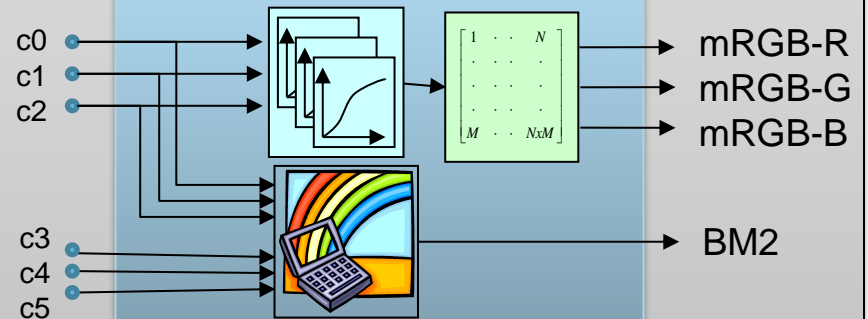
Channel 0: "mRGB-R"
Channel 1: "mRGB-G"
Channel 2: "mRGB-B"
Channel 3: "BM2"

Other  
Metadata  
Tags)

## AtoB1Tag (MPE)



## AtoM0Tag (MPE Calc)



# Example MCS Identification Profile

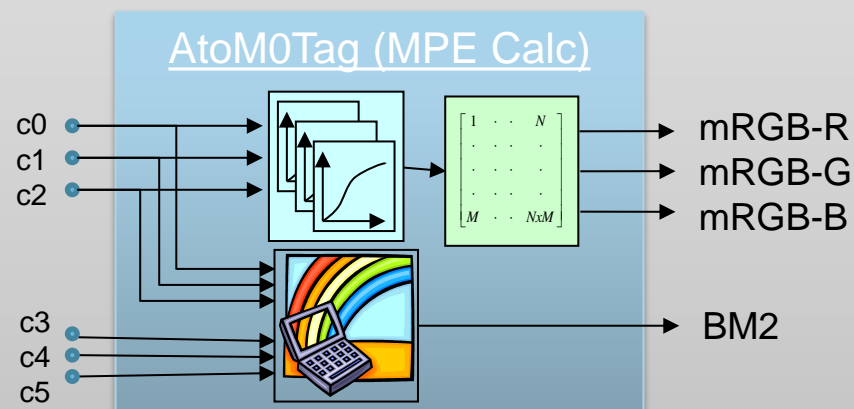
## Profile Header

Profile Class: 'mid '  
 Flags : MCS Sub-Set Requirements=false  
 Device: "nc0006"  
 MCS: "mc0004"  
 Profile SubClass: TBD

## Material Type Array Tag

Channel 0: "mRGB-R"  
 Channel 1: "mRGB-G"  
 Channel 2: "mRGB-B"  
 Channel 3: "BM2"

Other  
Metadata  
Tags)



# Example MCS Visualization Profile

## Profile Header

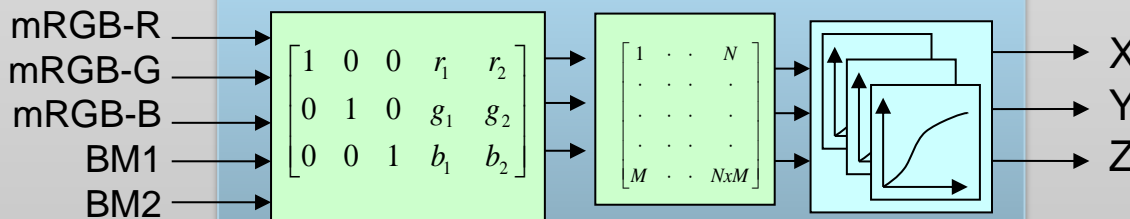
Profile Class: 'mvis'  
 Flags : MCS Sub-Set Requirements=false  
 MCS: "mc0005"  
 PCS: XYZ  
 Profile SubClass: TBD

Other  
Metadata  
Tags)

## Material Type Array Tag

Channel 0: "mRGB-R"  
 Channel 1: "mRGB-G"  
 Channel 2: "mRGB-B"  
 Channel 3: "BM1"  
 Channel 4: "BM2"

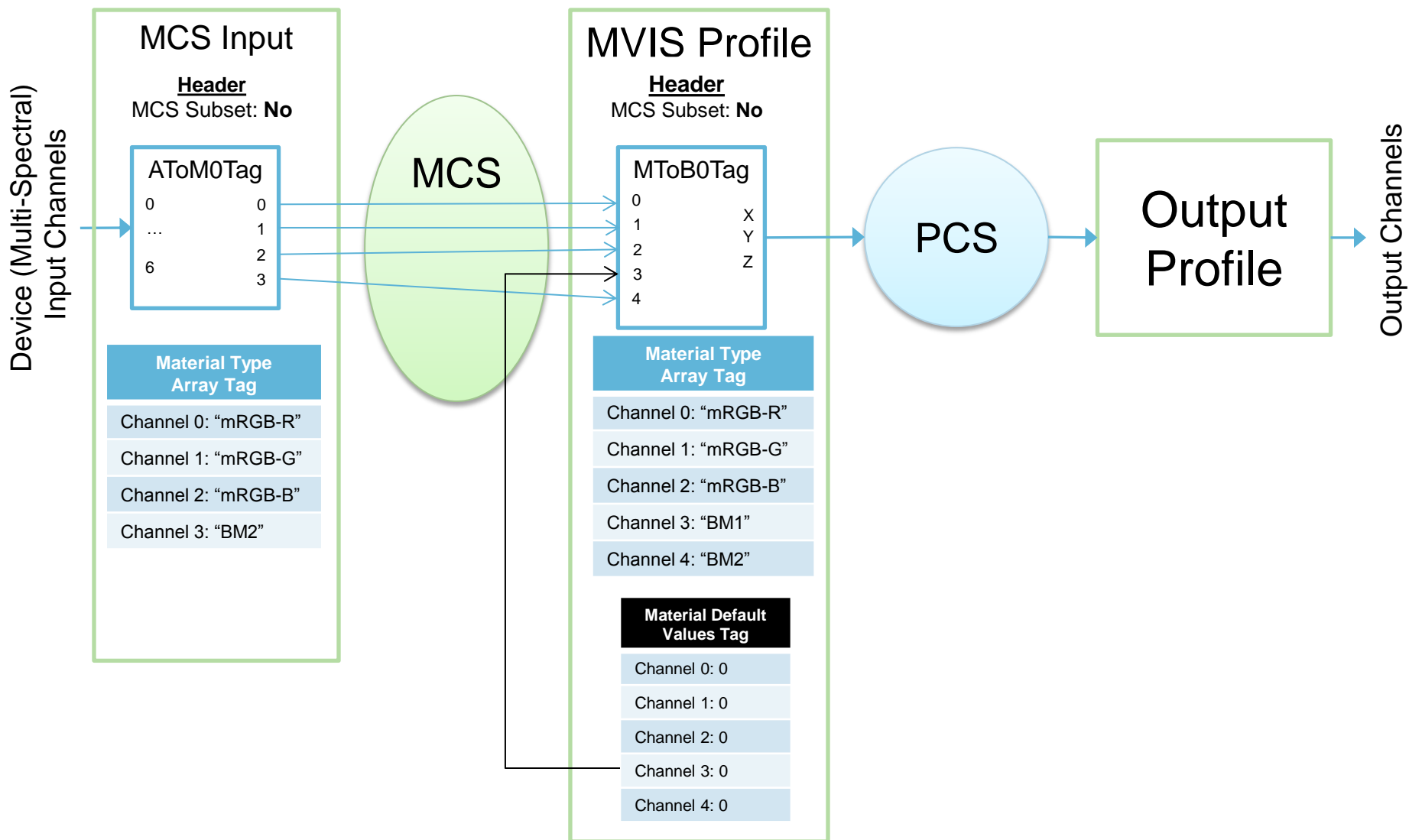
## AtoB1Tag (MPE)



## Material Default Values Tag

Channel 0: 0  
 Channel 1: 0  
 Channel 2: 0  
 Channel 3: 0  
 Channel 4: 0

# MCS Connection Example

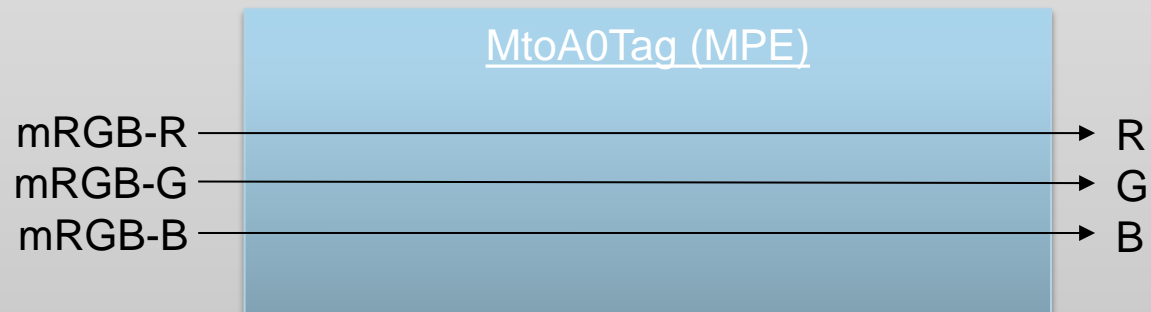


# Example MCS Link Profile

Profile Header
Profile Class: 'mInk'
Flags : MCS Sub-Set Requirements=true
Device: RGB
MCS: "mc0003"

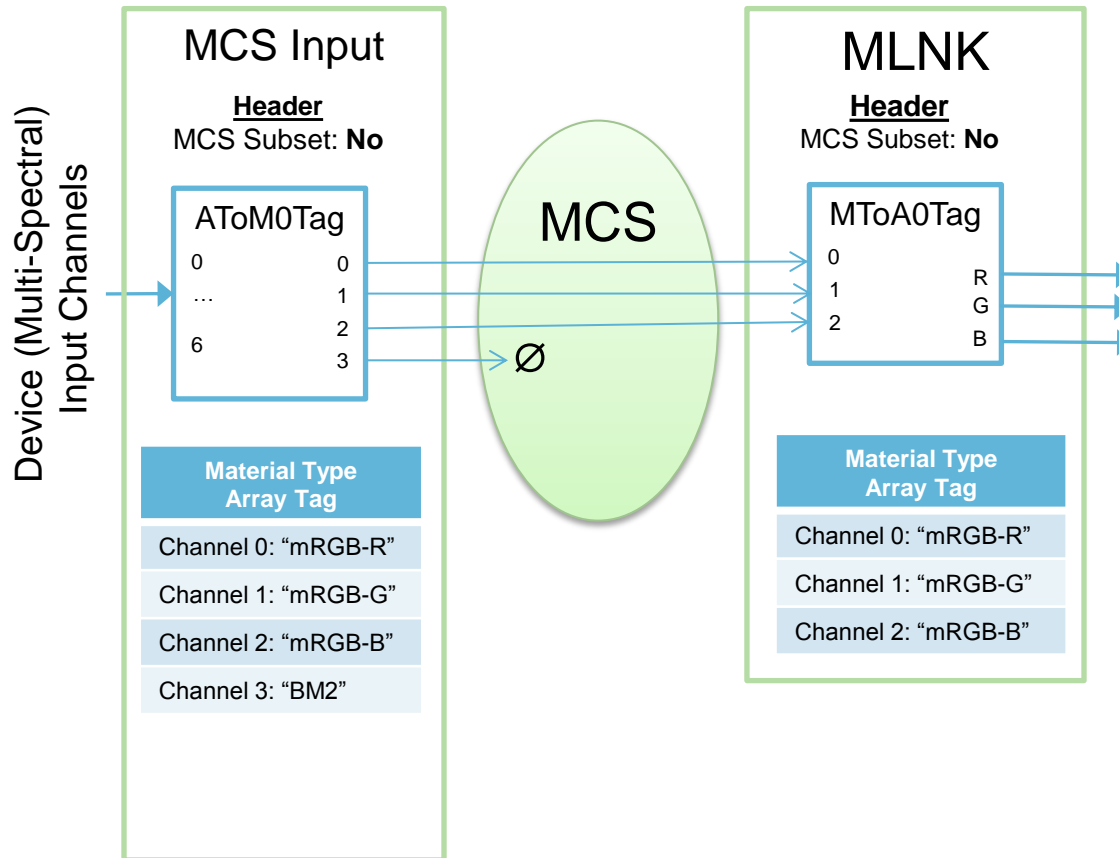
Other  
Metadata  
Tags)

Material Type Array Tag
Channel 0: "mRGB-R"
Channel 1: "mRGB-G"
Channel 2: "mRGB-B"





# MCS Connection Example



**Thank You!**

**Questions**



# **Spectral Measurement of Human Skin Colour**

**Kaida Xiao**

**University of Liverpool**

**ICC MIWG Meeting, Boston**



## Objectives

- ❑ To establish a skin spectral database for different ethnic groups, aging and body areas;**
- ❑ To develop a method to predict skin spectral using a digital camera;**
- ❑ To develop a skin image database covering true information of colour, spectral, texture, gloss, and shape.**



## Why Spectral Reflectance?

- More informative**
- Independent of illumination**
- True colour reproduction**
- Direct connects with skin chromophores**
  - melanin, haemoglobin

## Procedures

- Lighting measurement
- Image capture for colour chart
- Consent and information**
- Spectrophotometer Measurement
- Spectroradiometer Measurement
- Skin image capture by a 2D camera
- Facial image capture by a 3D camera



## Lighting in the booth

- Diffuse light
- D65 simulator

## Lighting Measurement

- TSR (White Diffuser)
- Digital Camera (white board)

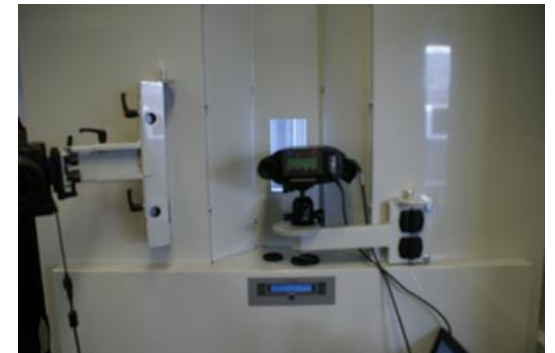
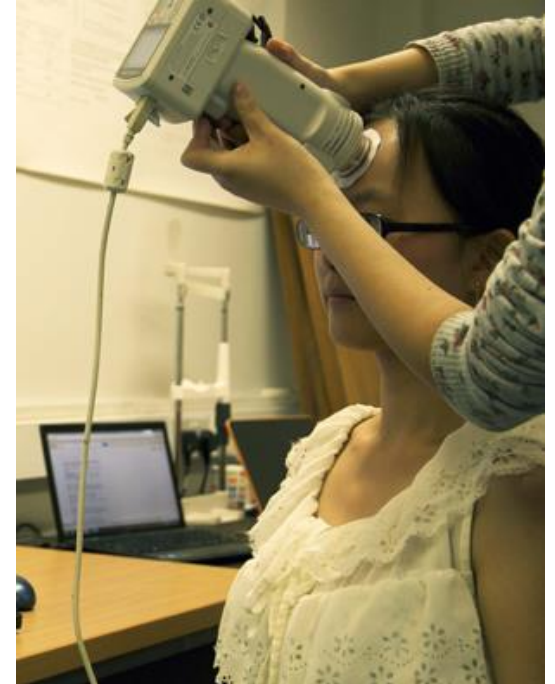


## Spectrophotometer

- 10 body positions
- 2 measurement sizes
- Low measurement pressure
- 3 repetitions

## Spectroradiometer

- 5 body positions
- Fixed distance
- Fixed measurement angle
- 3 repetitions





## Digital SLR Camera



- Fixed capture distance
- Fixed capture angle
  
- Fixed lens focus
- Fixed exposure setting
- Fixed ISO setting
- Fixed white balance
  
- Save in raw image
- 3 repetitions



## 3D photogrammetry system

- Built-in flash lighting
- Room Lighting
  
- Fixed capture distance
- Fixed capture angle
  
- Fixed lens focus
- Fixed exposure setting
  
- Save in 3D image





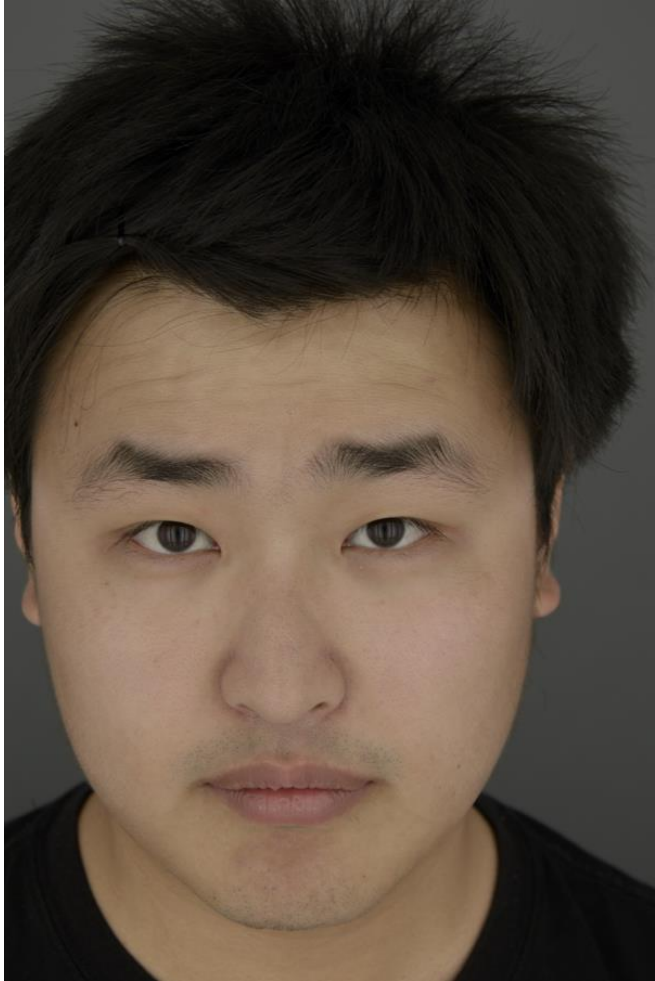
# Skin Image Capture



3D facial image



# Skin Image Capture



2D camera image



3D camera image



## Model Development

- ❑ Selection of colour chart
- ❑ Selection of skin colour database
- ❑ Applied Mathematical models
  - camera colour characterisation
  - camera sensitivity function prediction
  - skin reflectance re-construction

## Model Evaluation

Proposed skin spectral data



## Skin spectral data

Caucasian	Oriental	Sub Asian	African
32	61	4	5

## Skin colour chart

Silicone skin colour chart

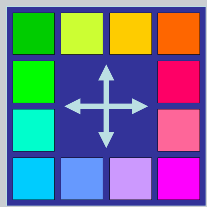
## Skin colour reflectance prediction model

New reflectance reconstruction model



# Thanks

**[k.xiao@liverpool.ac.uk](mailto:k.xiao@liverpool.ac.uk)**



ICC Medical Imaging Working Group  
Boston Meeting: Nov 11, 2014



dRGB

AAPM TG196 Progress

Michael Flynn  
Radiology Research  
Henry Ford Health System  
Detroit, MI





- sRGB is a standard RGB color space created cooperatively by HP and Microsoft in 1996 for use on monitors, printers and the Internet.
- the sRGB gamma cannot be expressed as a single numerical value. The overall gamma is approximately 2.2, consisting of a linear (gamma 1.0) section near black, and a non-linear section elsewhere
- IEC 61966-2-1:1999 is the official specification of sRGB. It provides viewing environment, encoding, and colorimetric details.

<http://en.wikipedia.org/wiki/sRGB>

## IEC 61966-2-1

### Colour Measurement and Management in Multimedia Systems and Equipment Part 2-1: Default RGB Colour Space – sRGB

1. GENERAL
  1. Introduction
  2. Scope
  3. Normative References
  4. Definitions
2. REFERENCE CONDITIONS
  1. Reference Display Conditions
  2. Reference Viewing Conditions
  3. Reference Observer Conditions
3. ENCODING CHARACTERISTICS
  1. Introduction
  2. Transformation from RGB values to 1931 CIE XYZ values
  3. Transformation from 1931 CIE XYZ values to RGB values

ANNEX A: Ambiguity in the Definition of the Term "Gamma"

ANNEX B: sRGB and ITU-R BT.709-2 Compatibility

ANNEX C: Usage Guidelines

ANNEX D: Typical Viewing Conditions

ANNEX E: Recommended Treatment for Viewing Conditions

ANNEX F: Bibliography



- The Adobe RGB color space is an RGB color space developed by Adobe Systems in 1998.
- It was designed to encompass most of the colors achievable on CMYK color printers, but by using RGB primary colors on a computer display.
- A gamma of 2.2 is assumed.
- The color space encompasses roughly 50% of the visible colors specified by the Lab color space, improving upon the gamut of the sRGB color space primarily in cyan-greens.

**Adobe RGB (1998)**

**Color Image Encoding**

*Version 2005-05, May 2005*

Introduction

1. Scope
2. References
3. Terms
4. Requirements
  1. General
  2. Reference Viewing Environment
  3. Adobe RGB (1998) Color Image Encoding
5. Indicating the use of Adobe RGB (1998) ..

Annex A: The Adobe RGB (1998) ICC profile

Annex B: Practical tolerances for display devices

Annex C: Implementation notes

[http://http://en.wikipedia.org/wiki/Adobe\\_RGB\\_color\\_space](http://http://en.wikipedia.org/wiki/Adobe_RGB_color_space)  
<http://www.adobe.com/digitalimag/pdfs/AdobeRGB1998.pdf>



## Reference Document:

### ACR-AAPM-SIIM standard

- The ACR-AAPM-SIIM technical guideline for electronic imaging was recently revised with participation by three professional Radiology organizations:
  - American College of Radiology
  - American Assoc. of Physicists in Medicine
  - Society for Imaging Informatics in Medicine
- The recently published guidelines contain specific recommendations for viewing conditions and display characteristics.
  - DICOM Grayscale with defined  $L_{\max}$  and  $L_{\min}$
  - D65 white point.
  - Undefined color gamut.

## ACR–AAPM–SIIM Technical Standard for Electronic Practice of Medical Imaging

JT Norweck, JA Seibert, KP Andriole,  
DA Clunie, BH Curran, MJ Flynn,  
E Krupinski, RP Lieto, DJ Peck, TAMian

---

### **Display**

1. Workstation Characteristics
  - f. Ergonomic factors
  2. Viewing Conditions
2. Display characteristics
  - a. Luminance response
    1. Ambient Luminance,  $L_{\text{amb}}$
    2. Minimum Luminance,  $L_{\text{min}}$
    3. Maximum Luminance,  $L_{\text{max}}$
    4. Luminance Ratio, LR
    5.  $L_{\text{max}}$  for Diagnostic & other
    6. Luminance vs Gray Level
    7. Calibration
    8. Quality Control
    9. White Point.
  - b. Pixel Pitch and Display Size

...  
J Digit Imaging (2013) 26:38–52

### AAPM on-line report No. 03 (2003)

- *Assessment of Display Performance for Medical Imaging Systems*
- “The intent of this report is to provide standard guidelines to practicing medical physicists, engineers, researchers, and radiologists for the performance evaluation of electronic display devices intended for medical use.”

### IEC 62563-1 (2009)

- *Medical Electrical Equipment – Medical Image Display Systems*  
*Part 1: Evaluation Methods.*
- “This International Standard provides evaluation methods for testing image display systems used in medical electrical equipment and medical electrical systems for diagnostic imaging.”

### CIE TC 1-93 (formed 2013)

- *Calculation of self-luminous neutral scale*
- Charge: To recommend a formula or computational method for an achromatic, neutral or gray scale for self-luminous (i.e. non-reflective) surfaces. (This computation complements CIE Lightness,  $L^*$ , which serves a similar purpose for reflective surfaces.)



AAPM Task Group No. 196

Requirements and methods for color displays in medicine.

Aldo Badano, PhD \*

Paul Boynton

Wei-Chung Cheng

Danny Deroo

Michael Flynn

Mikio Hasegawa

Patrick Le Callet

Takashi Matsui

Balazs Nagy

John Penczek

Craig Revie

Ehsan Samei \*

Peter Steven

Stan Swiderski

Gert Van Hoey

\* co-chair



---

A medical RGB color space (dRGB)  
for color managed emissive displays

---

Report of AAPM Task Group 196

First reading -> Dec 2014

<http://www.aapm.org/pubs/reports/>



# Color spaces compared

## (1) IEC 62563 terminology

Specification (1)	sRGB	aRGB	ACR	dRGB
Luminance Response	~2.2 power function	2.199 power function	DICOM GSDF	DICOM GSDF
Color Gamut	HDTV based ITU-R BT.709-5	'Wide' (extended G)	-nd-	[*] (referenced)
$L_{max}$ , cd/m <sup>2</sup>	80	160 (125-200)	350/420/250	350 (250-450)
$L_{min}$ , cd/m <sup>2</sup>	-nd-	0.56	$L_{max} / LR$	$L_{max} / LR$
Luminance Ratio (LR)	-nd-	287.9 (230-400)	350 (> 250)	350 (300-400)
White Point	D65	D65	D65	D65
Gray tracking	-nd-	-nd-	-nd-	IEC MT51
Surround	20% refl. lx	Gray (D65, 2°) 20% $L_{max}$	-nd-	Gray (D65, >2°) 20% $L_{max}$
Ambient Illumination, lx	64 (D50)	32 (D65) (16-64)	20-40	-nd-
Veiling Glare	1.0%	accounted	-nd-	-nd-
$L_{amb}$ , cd/m <sup>2</sup>	-nd-	-nd-	$L_{amb} < \frac{1}{4} L_{min}$	$L_{amb} < [\frac{1}{4}, \frac{2}{3}] L_{min}$



## dRGB - as a color space framework.

- Variable neutral luminance response:
  - DICOM GSDF (Lmax & Lmin in profile description)
  - Lmax = 250-450.
  - LR = 300-400 (LR = Lmax/Lmin)
  - May have Lamb dependence.
- Variable color gamut:
  - sRGB default
  - Other spaces indicated by profile description
- Surround is specified as the near field effecting adaption.
- Lamb uses the 1/4<sup>th</sup> or 2/3<sup>rd</sup> criteria from AAPM TG-18.



- Monitor Calibration: The dRGB color space will provide a full specification for the calibration of a medical monitor including white point, color space, luminance response, luminance ratio, and viewing conditions.
- Monitor Specifications: The dRGB color space will provide specifications for the performance of a monitor for which a manufacturer can provide firmware correction through OSD selection for the target performance.
- Display Profile: For monitors conforming with the dRGB color space, an ICC profile can be used by a color management system to transform image values from profile connection space to display values.
- Source Profile: For images in the dRGB color space, an ICC profile can be used by a color management system to transform these image values to profile connection space.

*AAPM approved charge, 'AAPM-IPC-IISC\_TG-mRGB\_Charge-v3'*



### Primary (a.k.a. diagnostic)

- Primary display systems are those used for the interpretation of medical images.
- They are typically used in radiology and in certain medical specialties such as orthopedics.

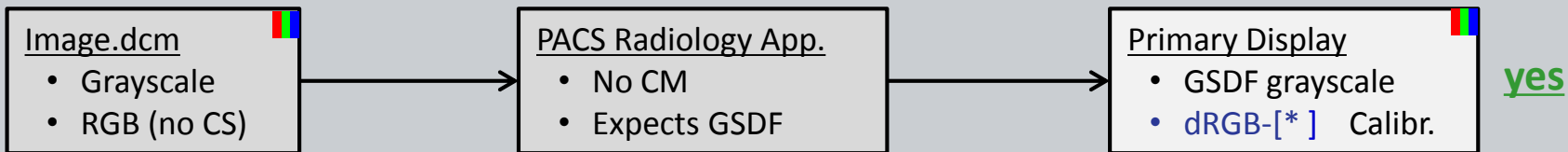
### Secondary (a.k.a. review, enterprise, other)

- Secondary systems are those used for viewing medical images for purposes other than for providing a medical interpretation.
- They are usually used for viewing images by general medical staff and medical specialists other than radiologists and utilized after an interpretive report is provided for the images.

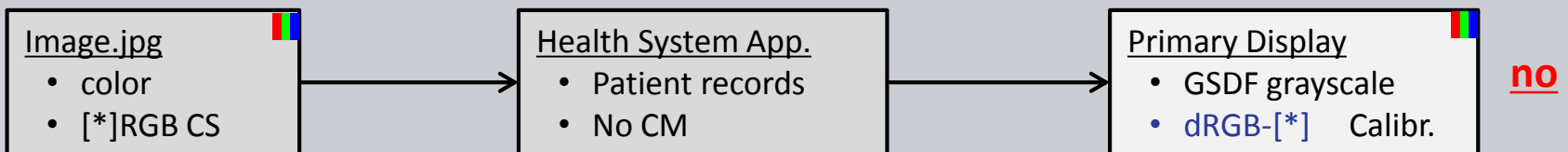
*From Section 2.5 of AAPM On-Line Report No. 3*

1. Medical image presentation on a workstation with DICOM calibrated primary monitors used for medical interpretations

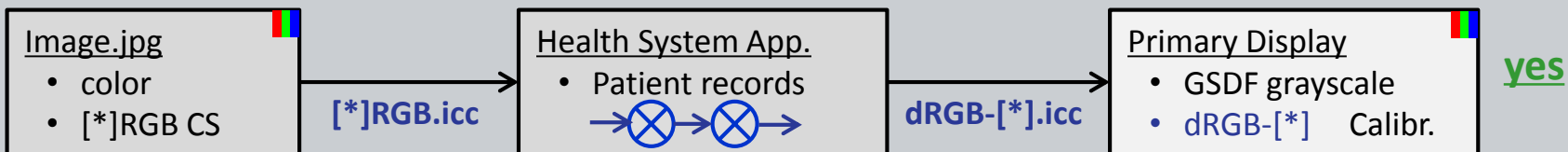
1-A



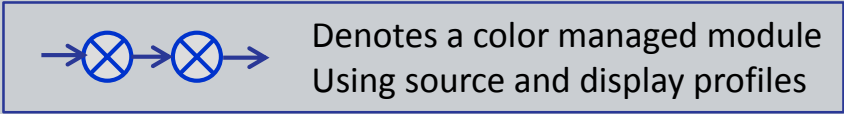
1-B



1-C

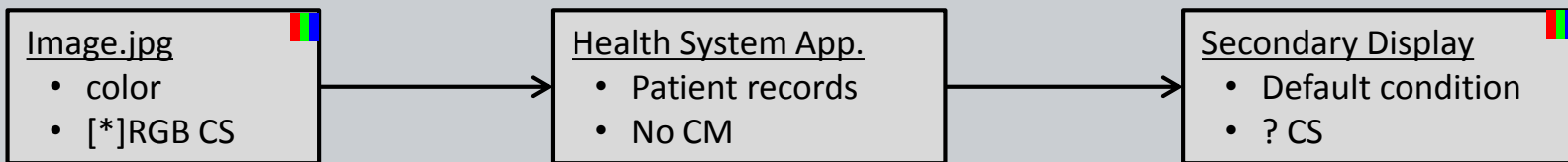


- Case A: Correct    Grayscale & pseudo-color DICOM images with GSDF neutral tones.
- Case B: Incorrect    Color photograph is presented with GSDF neutral tones.
- Case C: Correct    Color photograph is presented with the intended color space.



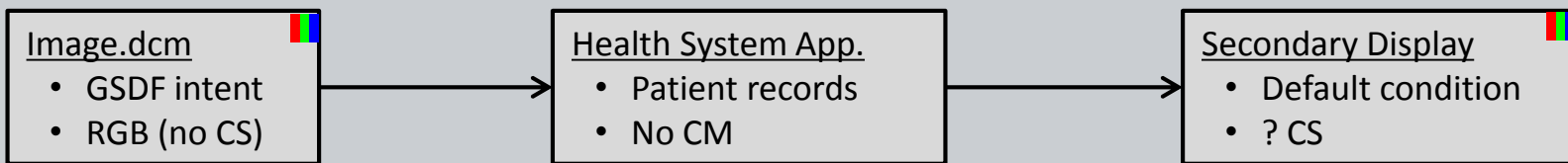
2. Medical image presentation on a workstation with secondary monitors used for reviewing patient information.

2-A



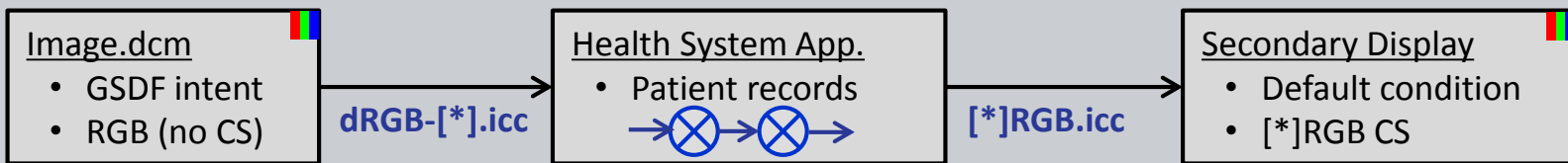
?

2-B



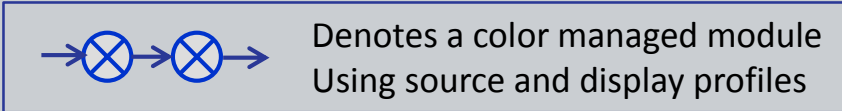
no

2-C



yes

- Case A: Uncertain Color photograph is presented with the default configuration .
- Case B: Incorrect Grayscale & pseudo-color DICOM images not presented with GSDF.
- Case C: Correct Grayscale & pseudo-color DICOM images are mapped to GSDF



## Dependence of GSDF on Lmax, Lmin, and Lamb

- Display Profiles

The approach is to have a profile for each luminance states (Lmin & Lmax) and state these conditions in the profile description. This is usually shown when profile lists are displayed, thus supporting both manual and automated selection (*see MIWG 6-Aug-2014 minutes*) .

- Source Profiles

The approach is to require that the color management engine register the Lmax, Lmin and Lamb conditions of the color space associated with image creation. (*Note: DICOM does not currently define these values but simply denotes the GSDF space as p values. Thus default values for the luminance state are appropriate.*)

- dRGB by reference

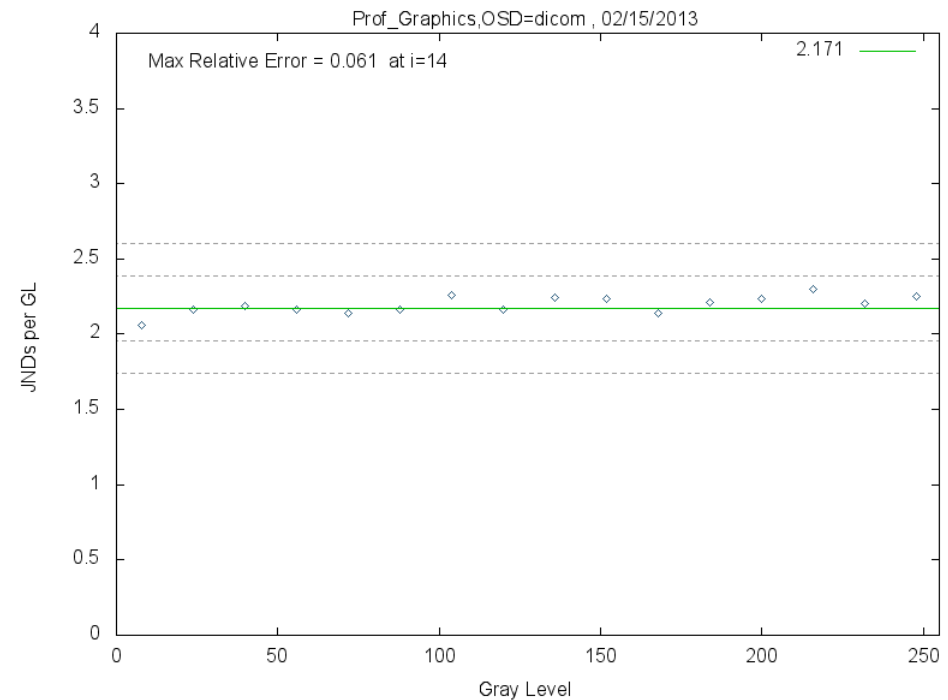
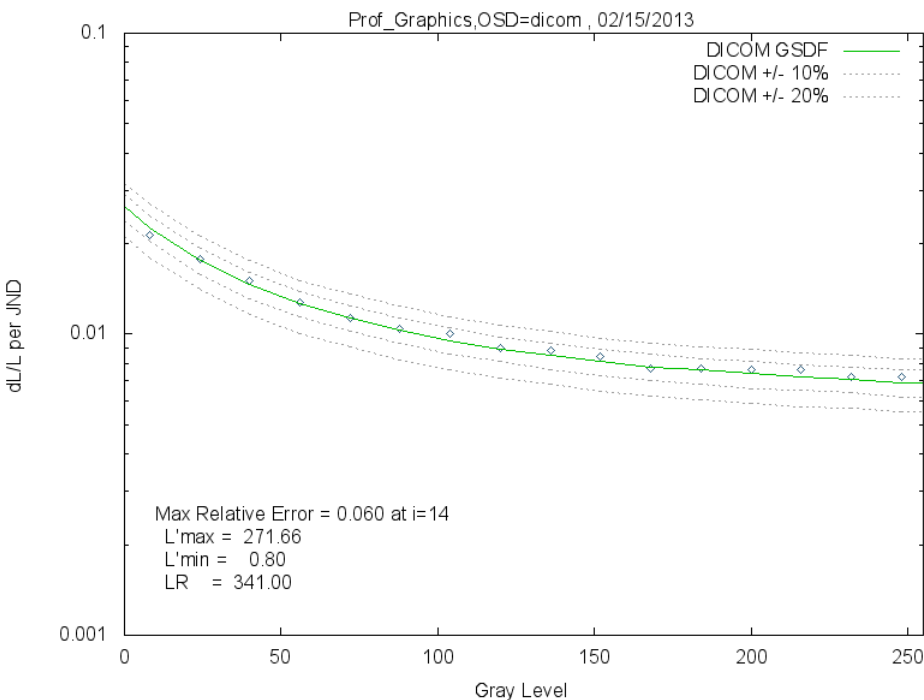
A color management engine may identify the color space as dRGB by reference and register the luminance state in the CMS application. This permits use of the DICOM GSDF polynomial expressions.



## Neutral scale allowed error

The dRGB neutral scale, DICOM GSDF, is to be evaluated in terms of the slope (i.e. contrast) in relation to display value.

- dL/L vs DDL (AAPM TG18)
- JNDs vs DDL (DICOM)

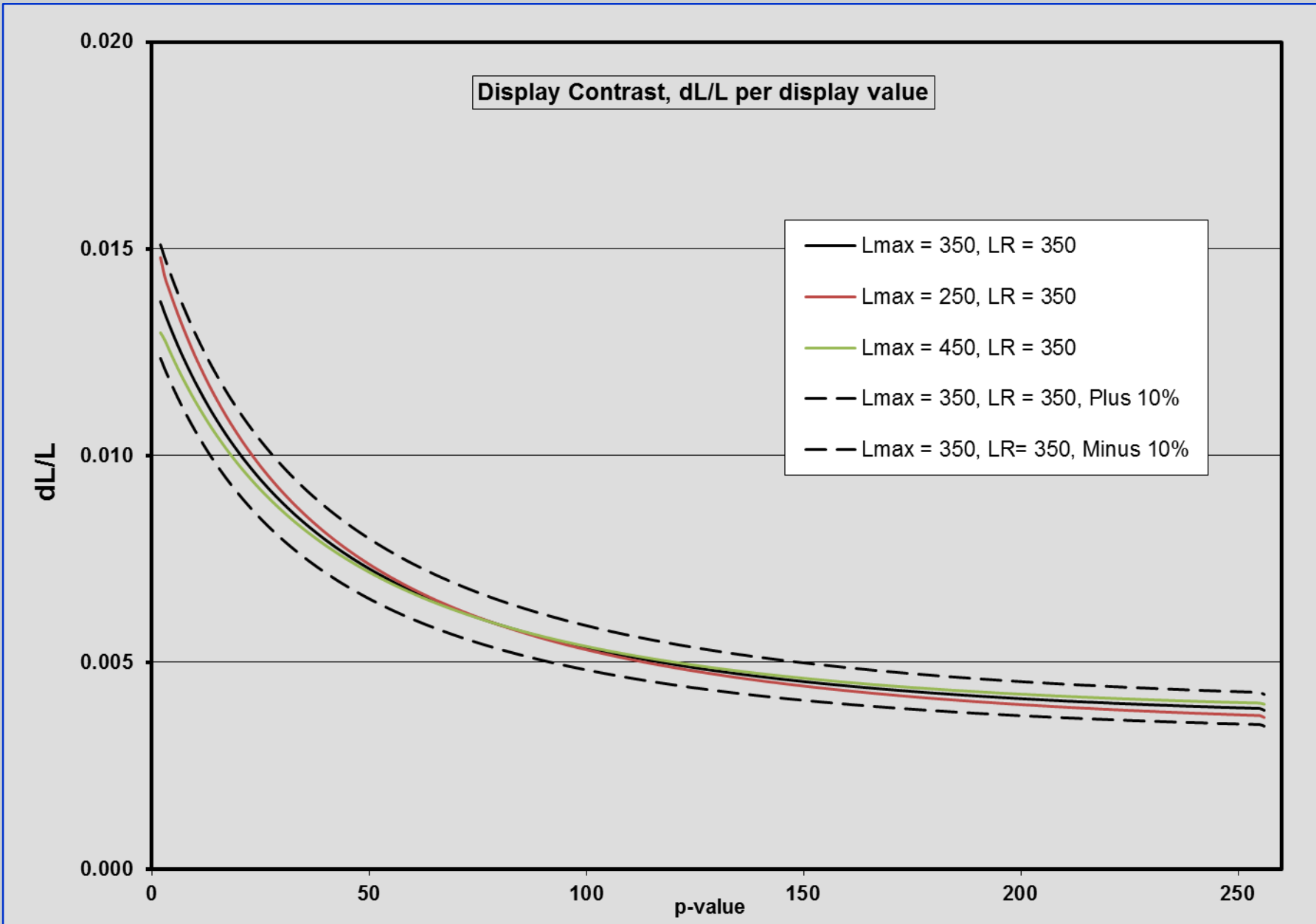


For coarse measurements made at 17 gray levels (AAPM TG18), conformance is well established (ACR Guidelines).

- 10% for primary displays
- 20% for secondary displays

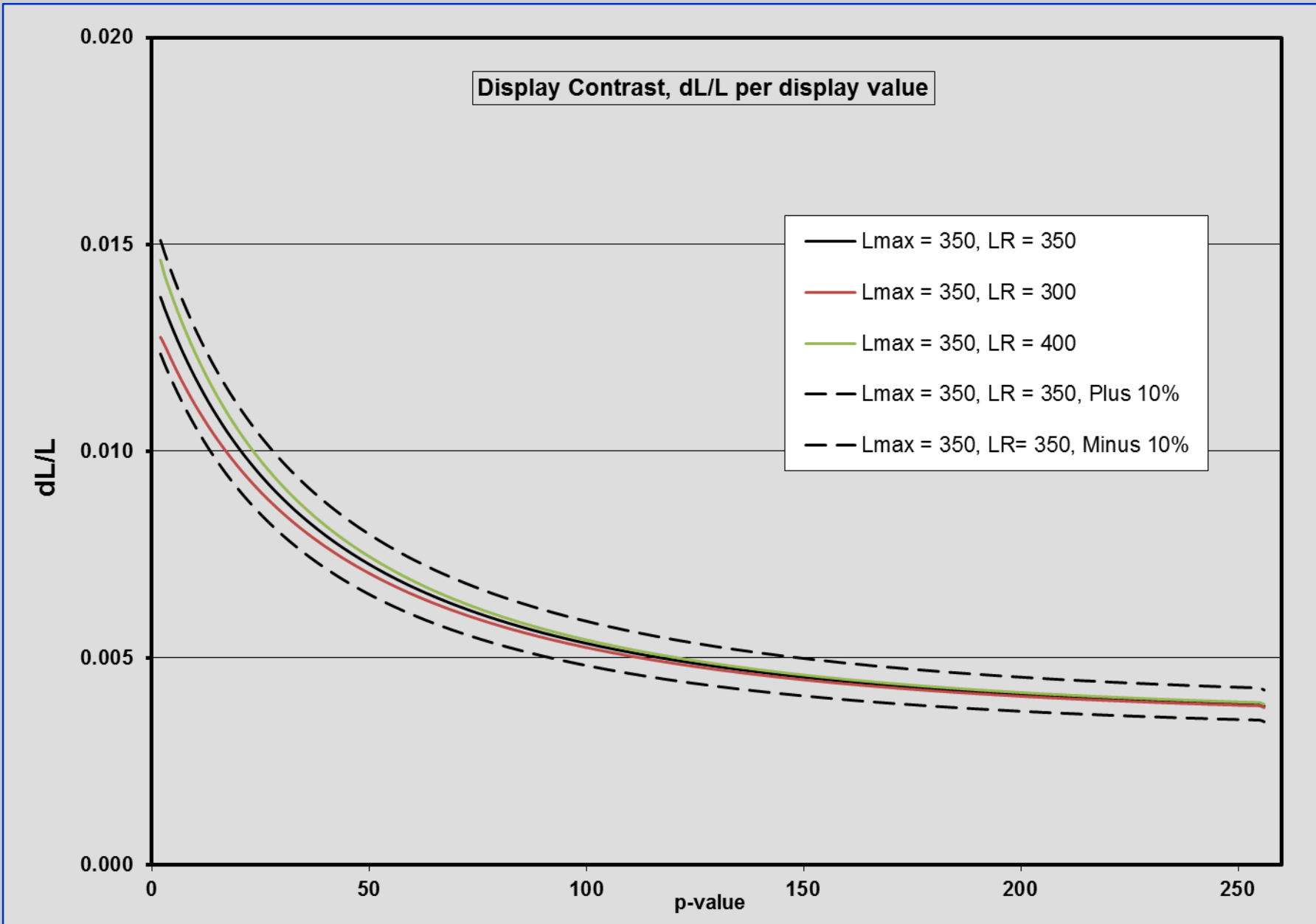


Lmax variation of 350 +/- 100 (LR=350)





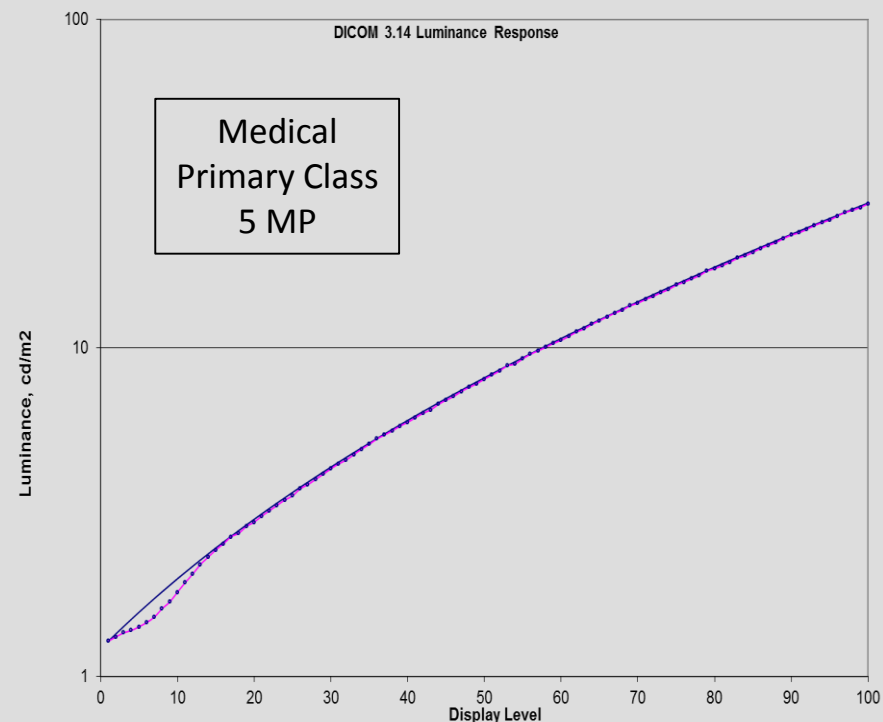
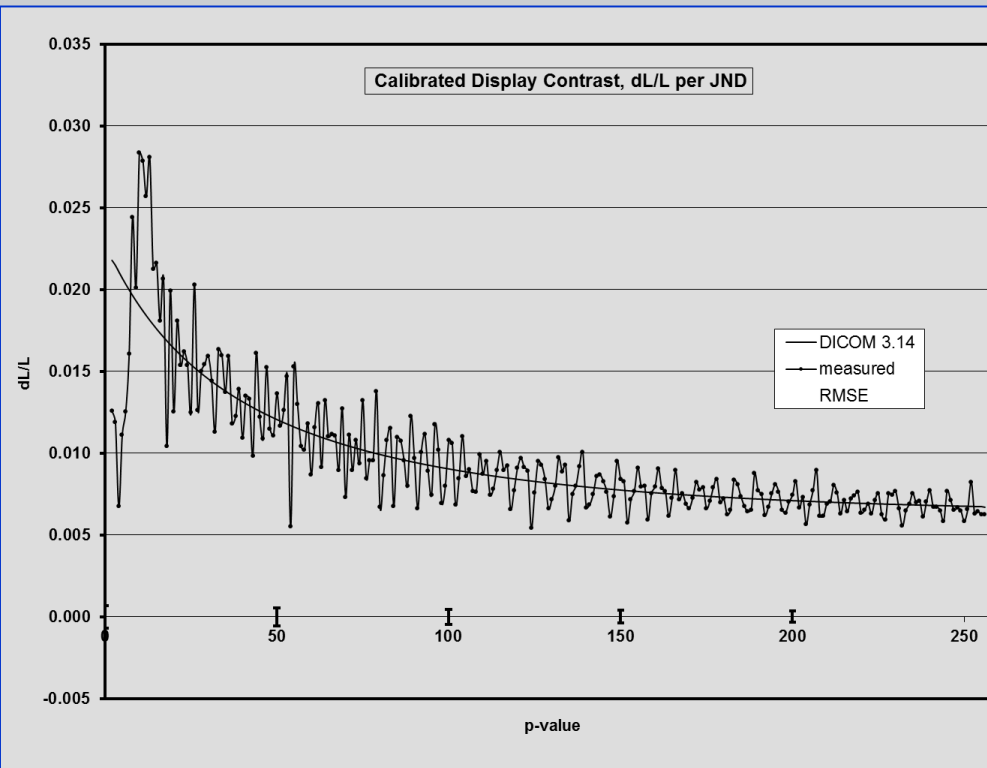
LR variation of 350 +/- 50 (Lmax=350)





## Neutral scale allowed error

- GSDF contrast performance for incremental display values is also required for dRGB conformance.
- This result shows low luminance contrast deviations that were not identified on the coarse measurement (17 levels)

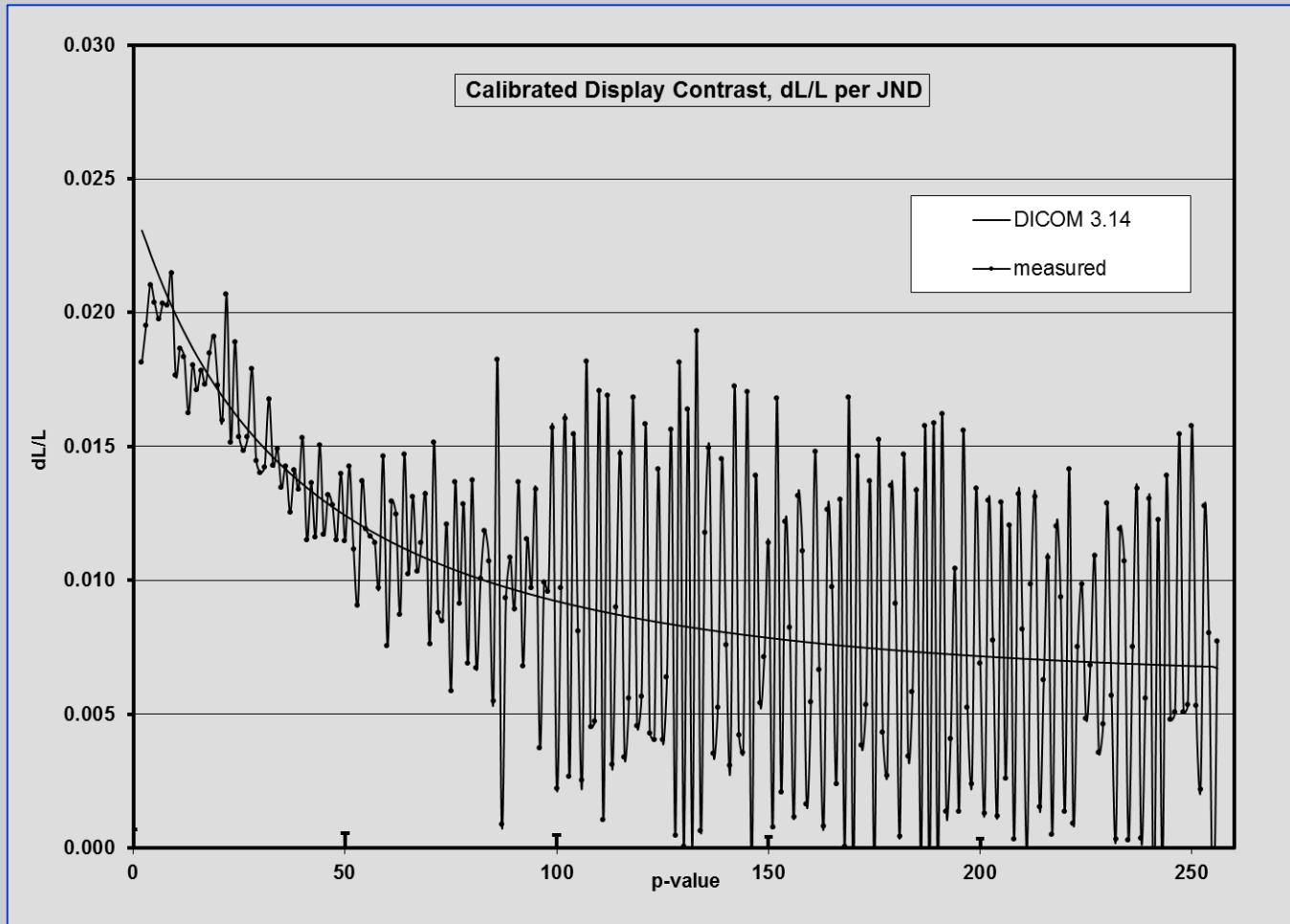


Appropriate conformance requirements for 256 level measures are currently under discussion. It is generally recognized that these measurements will require precise photometers and may have more deviation than coarse measures.



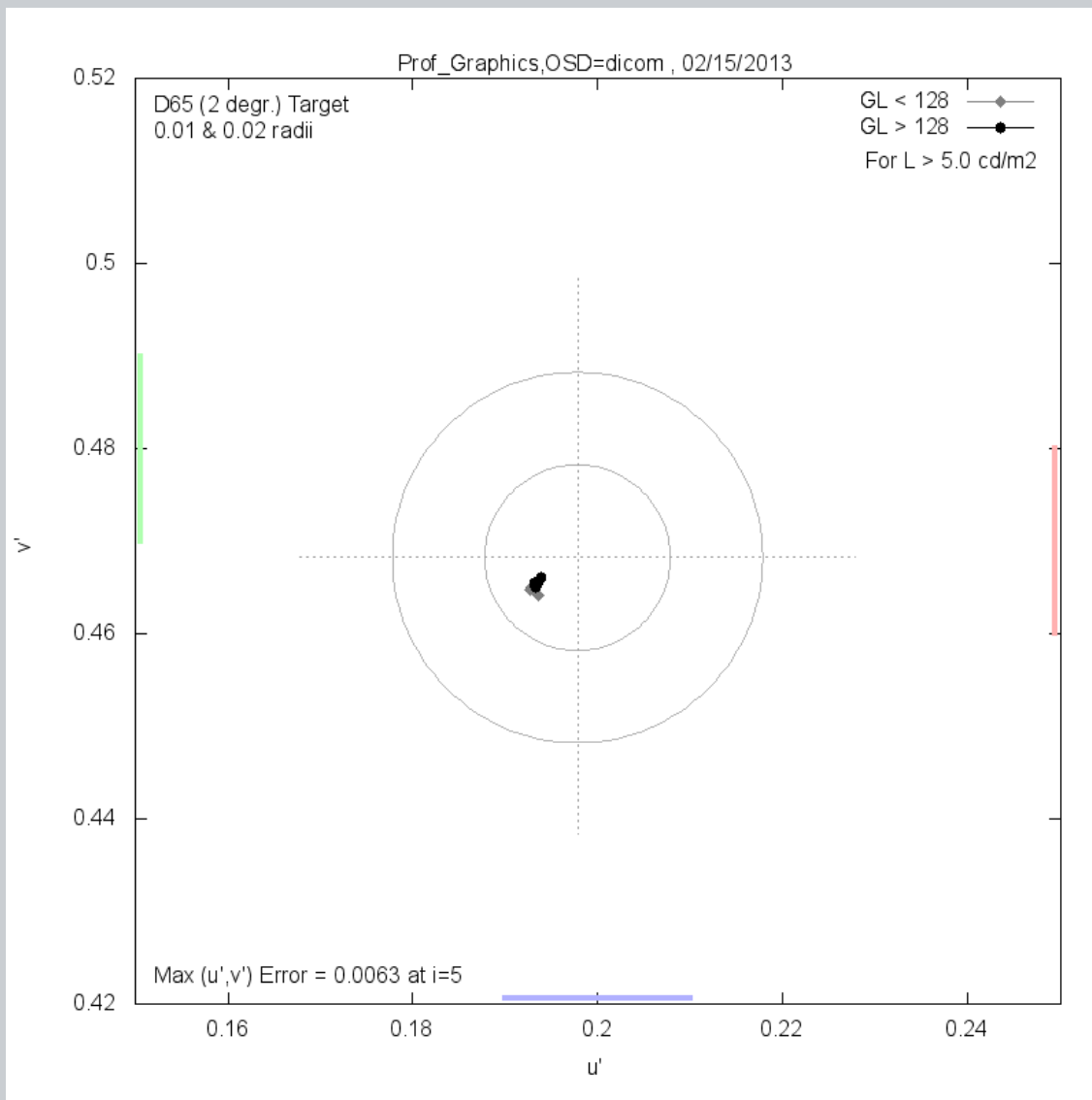


This result shows cyclic contrast deviations at high luminance indicative of a luminance response with staircase characteristics.





The default white point for dRGB is d65 (2 degr).



- AAPM TG196 has recently reported on medical gray tracking measures.
- dRGB presently requires conformnace to within an 0.01 radius for coarse measurements.
- Values below 5 cd/m<sup>2</sup> are not considered.



The default color space for dRGB is sRGB.

- Other color spaces are permitted with documentation in the profile description.
- **Appropriate conformance error for the color primaries are presently being discussed.**

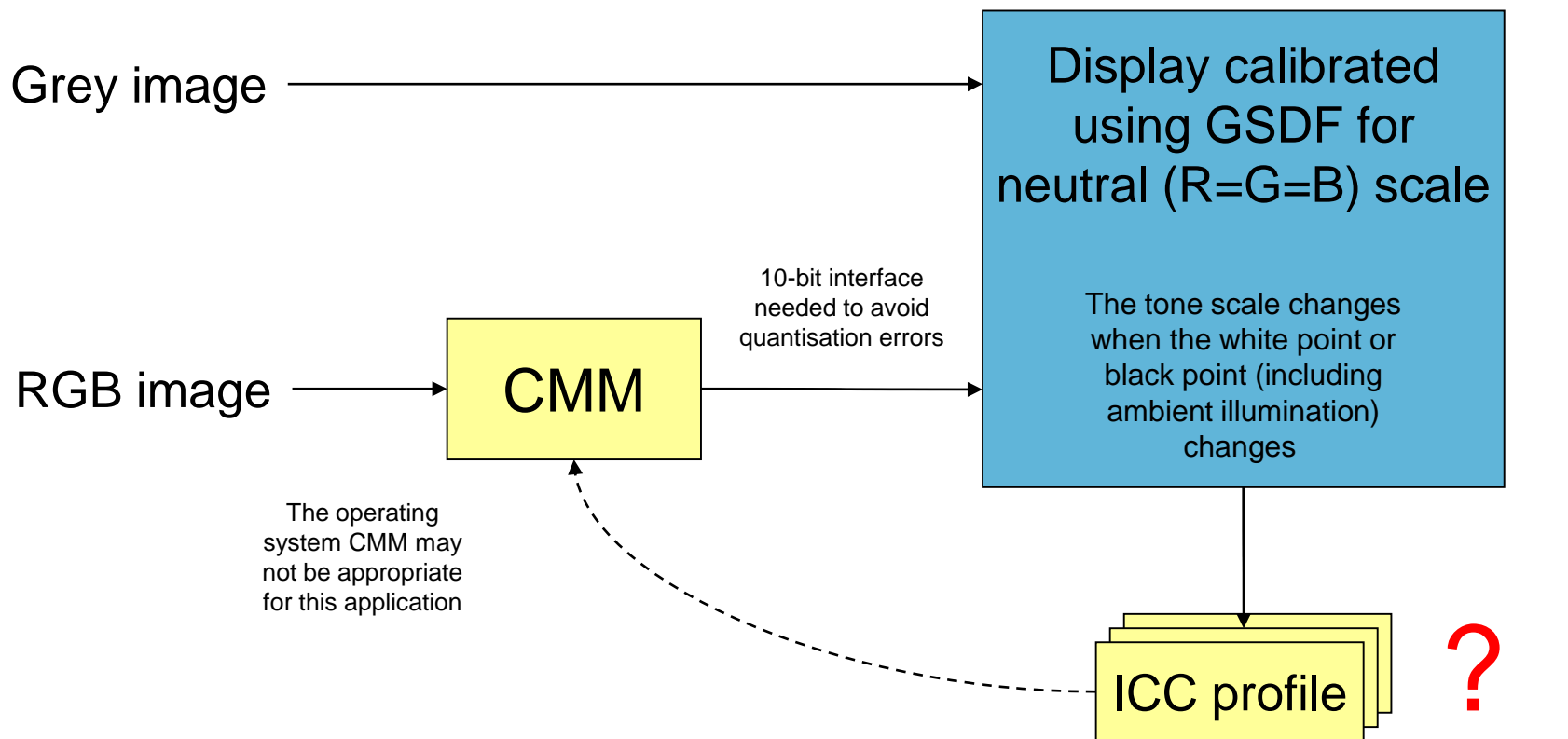
Note: dRGB is a color space framework primarily directed at the incorporation of the DICOM GSDF neutral scale. As such wide latitude is provided for the color gamut and white point.

# Display profiling requirements for mRGB / dRGB

**Craig Revie, FFEI Limited**

**1<sup>st</sup> November 2014**

# Proposed DICOM RGB colour space



A set of example/default profiles could be developed for different white/black range and could be posted on the ICC web site

## Requirements

- **Matrix / TRC profiles are not suitable for this application**
- **Few ICC Profiling applications support LUT-based tables**
- **Commercially-available profiling software that can build LUT-based profiles modifies the graphics card LUT**
  - This produces poor results for self-calibrating displays

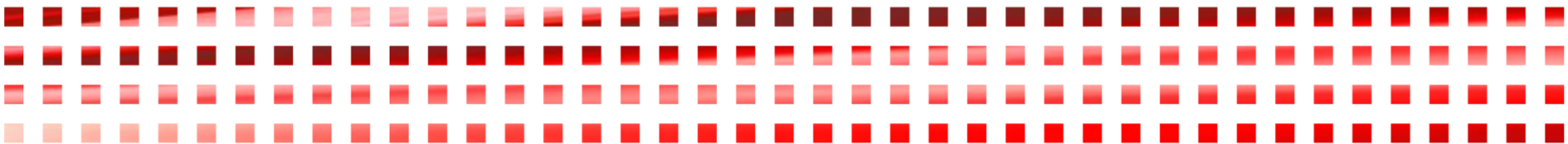
***Proposal: the ICC MIWG should specify requirements for display profiling software and work with manufacturers to test and deliver suitable profile creation software***



# MIWG – Displays

Update reference images for display testing

November 1st 2014



## Call for participation

- On Oct 3rd 2014 a call for participation was sent out to the MIWG – Displays.
- We are looking for medical color images that can be used as reference/test images by the MIWG.
- We are also looking for people that are willing to characterize their (relevant) color display and share the measurement data with the group
- See: [ftp://tok\\_icc\\_miwg\\_displays:xbochi0@ftp.barco.com](ftp://tok_icc_miwg_displays:xbochi0@ftp.barco.com)



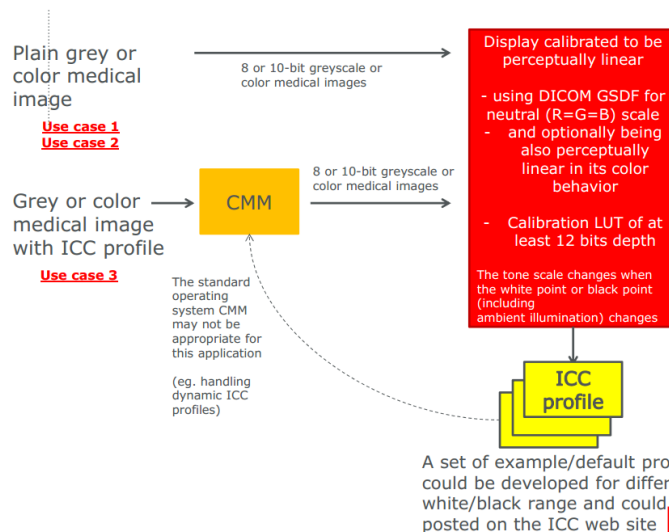
# Status

- Several people reacted to the request and indicated they are willing to contribute.
- Some radiology and pathology images already have been contributed to the FTP server
- David provided a list of publicly available images
- Other people expressed interest but raised specific questions about the goals and the method.
- The next few slides will try to answer these questions

# Background of this exercise

## ■ Background

- The MIWG Displays defined an architecture of which we believe it is suitable for color medical imaging. (see eg. [http://www.color.org/groups/medical/Minutes\\_Aug2014\\_SpecialTopics.pdf](http://www.color.org/groups/medical/Minutes_Aug2014_SpecialTopics.pdf))
- This architecture however has not been validated yet, so we don't know whether it is possible to meet the requirements and tolerances that are needed for accurate visualization of medical (color) images



## Goals of this exercise

- Goal: validating by means of bench testing and software simulations whether the proposed architecture is fit for purpose
  - Assembling a representative set of medical color images, together with corresponding requirements /performance needs for visualization
  - Selecting a number of display systems that will be used for the bench testing & simulations
  - Measuring/characterizing these display systems such that we can run software simulations of different configurations instead of having to “build” all configurations (eg. display bit depth, size of LUTs, ...)
  - Performing the bench testing and simulation work

## Relevance of gathering “unrelated measurement data”

- It is true that performing accurate, consistent measurements to characterize color behavior of color displays is a challenge (variability of measurement equipment, measurement environment, stability of displays, ...)
- This was discussed in the MIWG group and it was decided that nevertheless we feel it is useful to gather measurement data (details of what to measure have been provided) as long as the data is accompanied with a clear description of display ID & setting, measurement conditions and measurement equipment description.

## Relevance of gathering “medical color images”

- We are not aiming to collect any “color medical image”
- The goal really is to look for a limited set of representative/relevant color medical images, with corresponding specification of what minimum performance the visualization needs to conform to.
  - Eg. a greyscale PACS image, requirement (guidance): deviate no more than 10% from GSDF
  - Eg. a dermatology image, requirement: absolute colorimetric correct visualization with no more than 2  $\Delta E_{2000}$  maximum error
  - Eg. a quantitative imaging doppler ultrasound image, requirement: perceptually linear color behavior with no more than 15% variability in  $\Delta E_{2000}$  contrast/step along the color scale

## Suggested planning

- Until Nov 28th
  - Continue collection of representative images and display measurement data
  
- December (email & conference call)
  - Selection of the exact display systems to be used
  - Selection of the exact images to be used
  
- Early Q1 2015: bench testing & simulation
  
- March 2015: in depth discussion of the results during ICC MIWG face-to-face meeting

Questions?

# Recommended Image Capture Workflow for Medicine Photography

John Penczek

Oct. 13, 2014

## Introduction:

This general procedure outlines a recommended digital camera image capture workflow that can be used to improve image color accuracy.

## Required equipment:

- Digital color camera capable of exporting raw format image files. An ability to perform an in-camera white balance is also desirable.
- Reference color test chart (e.g. ColorChecker or Spyderchecker)
- Light source that can provide uniform illumination on the image plane at a 45 degree inclination angle. The light source should produce spectrally smooth broadband white light, approximating daylight.
- Software that is able to import RAW image files from camera, perform a white balance, and export the image in a 16-bit TIF format.
- Color correction software that can recognize each color in the reference color chart, create an HSL Preset file or ICC profile, and use this file to color correct the images of interest.

## Procedure:

### Image capture

1. Setup up the illumination and background for photographing the object of interest. The background should be a neutral gray color, ideally a 20% gray. The light source should produce light that is nominally at a 45 degree inclination angle to the image plane, and uniformly illuminating the entire region for the relevant image area.
2. Place a uniform gray target in the image plane. It should ideally be 20% gray. Skip this step if the camera does not have in-camera white balance capability.
3. Position the camera in front of the gray target and align the camera so that its optical axis is centered on the chart and perpendicular to it. It is best to use a tripod, or similar mechanism, to hold the camera stationary for the remainder of the photographs. Adjust the camera field of view so that it does not extend beyond the gray background. This field of view should be fixed for all photographs.
4. Use the in-camera white balance function to determine the proper white balance for this lighting condition. Use this white balance setting for all subsequent photographs. Skip this step if the camera does not have in-camera white balance capability.



5. Place the reference color test chart in the image plane. The camera field of view should capture all of the colors in the chart. The optical axis of the camera should be centered on the chart and perpendicular to it.
6. Set the exposure time to such that the brightest objects in the image are approximately 90% of the maximum brightness. The brightest object should be the whitest color patch. Use the intensity histogram (if available) to ensure that the image is not overexposed.
7. Capture the image of the reference color test chart and export the image in RAW file format. In some cameras, it is possible to use a “neutral” mode RAW format which minimizes perceptual enhancements.
8. Replace the reference color test chart with the objects to be photographed. Determine the proper exposure times for each object, and export the images in the same RAW file format.

#### Color correction

1. Import the RAW file of the reference color test chart into a program that is capable of performing a white balance on the image. Use the program to set the whitest color patch to an exposure of 90%, or RGB= 230, 230, 230. Then set the darkest patch to an exposure of 4%, or RGB= 10, 10, 10. If the black patch is below this level, then use the current setting or reshoot the photograph with brighter illumination. Export the white-balanced image as a 16-bit color TIF file.
2. Repeat this process for all of the other images taken under these image capture conditions.
3. Import the 16-bit TIF file of the reference color test chart image into a program that can recognize the reference color chart and create a HSL Preset or ICC profile based on that image. The color correction software should automatically find the centers of each color patch. Use colorimetric mode to create the HSL Preset or ICC profile.
4. Import the TIF files of the other photographed objects into the image editing program that is capable of using HSL Presets or ICC profiles. Apply the HSL Preset or ICC profile to each image and save the new color-corrected image in the desired format.

# PETRI PLATES IMAGE ACQUISITION :

# A COLOR CALIBRATION METHOD

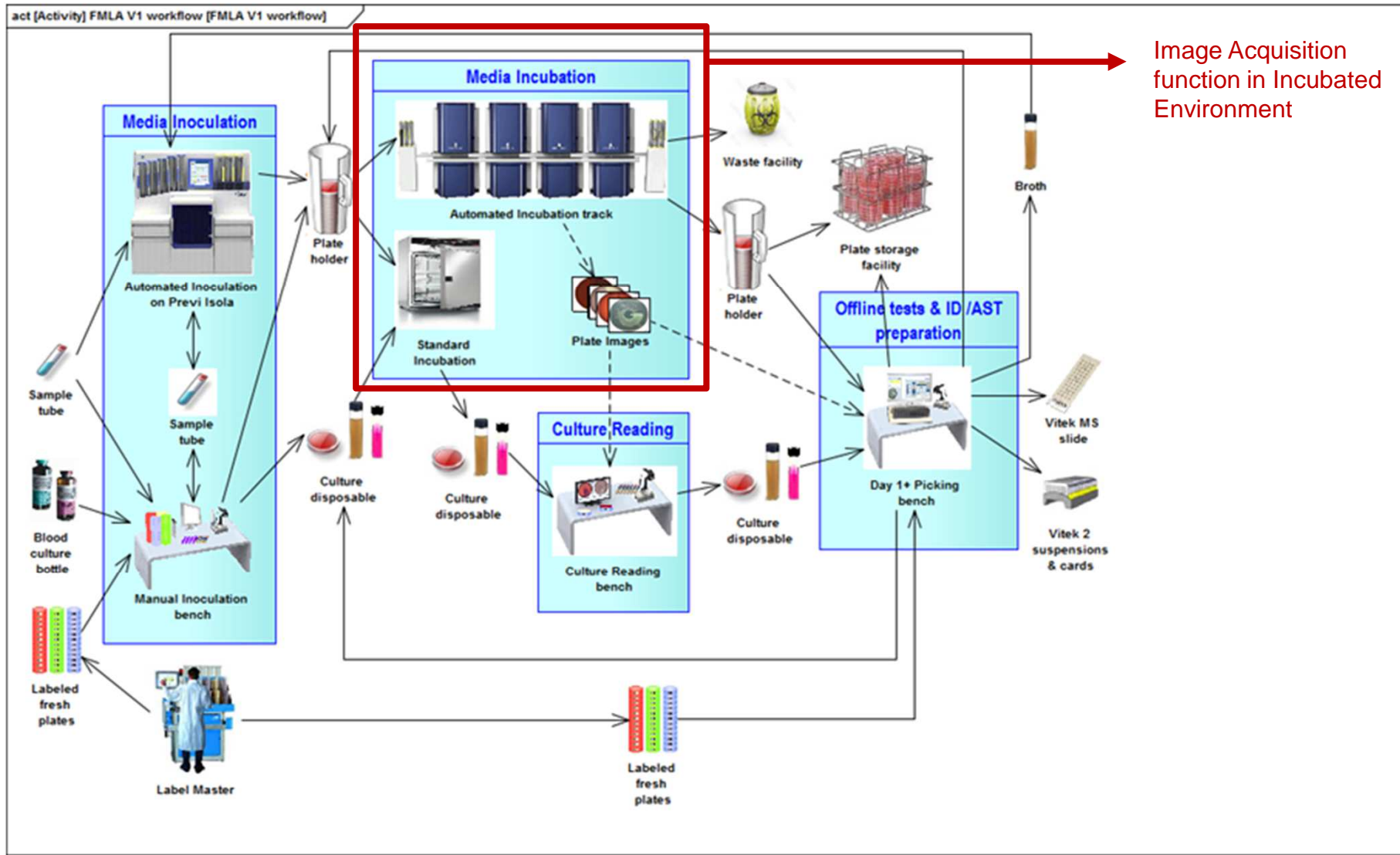
PIONEERING DIAGNOSTICS

Jeremie Pescatore, System Architect, bioMérieux

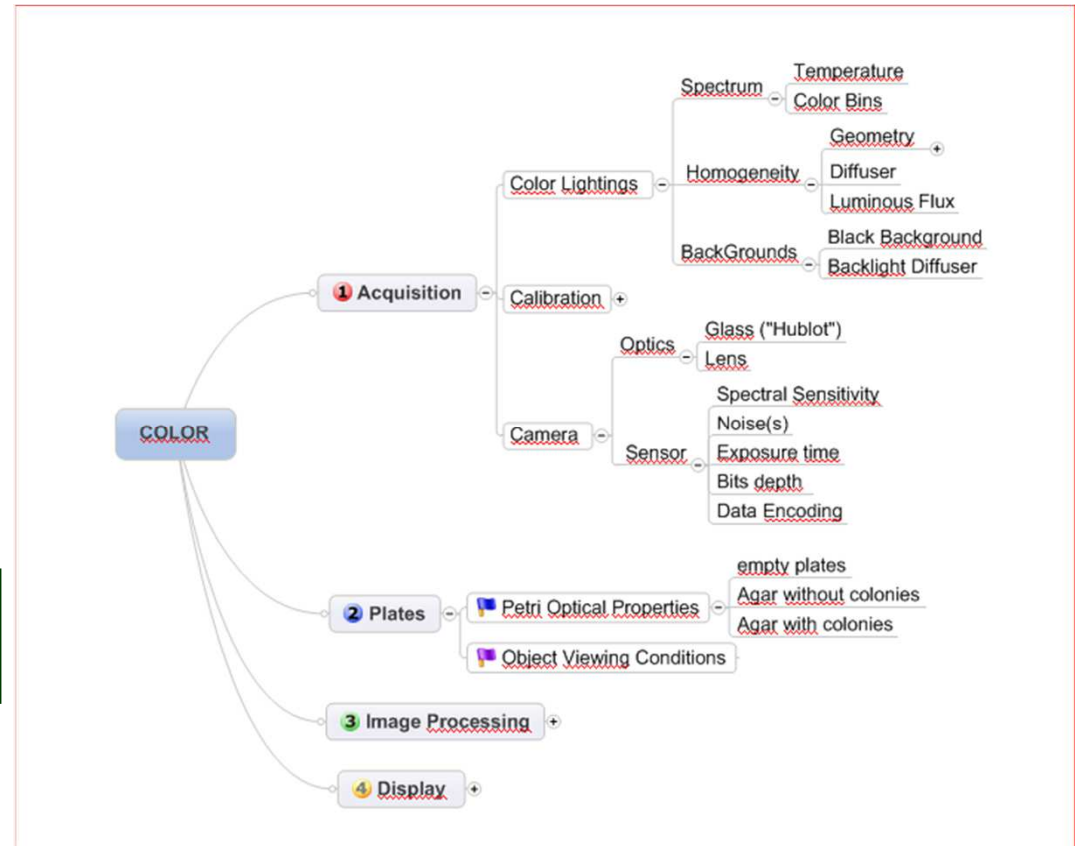
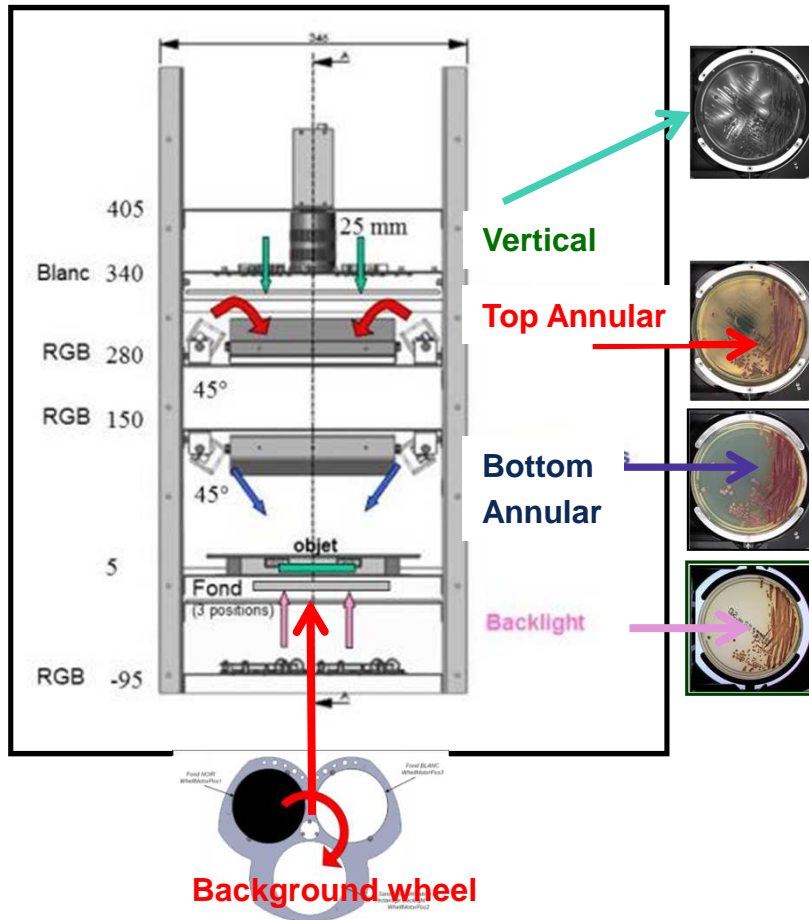
Revision 0



# Lab Automation Workflow : Where is Petri dish image acquisition ?

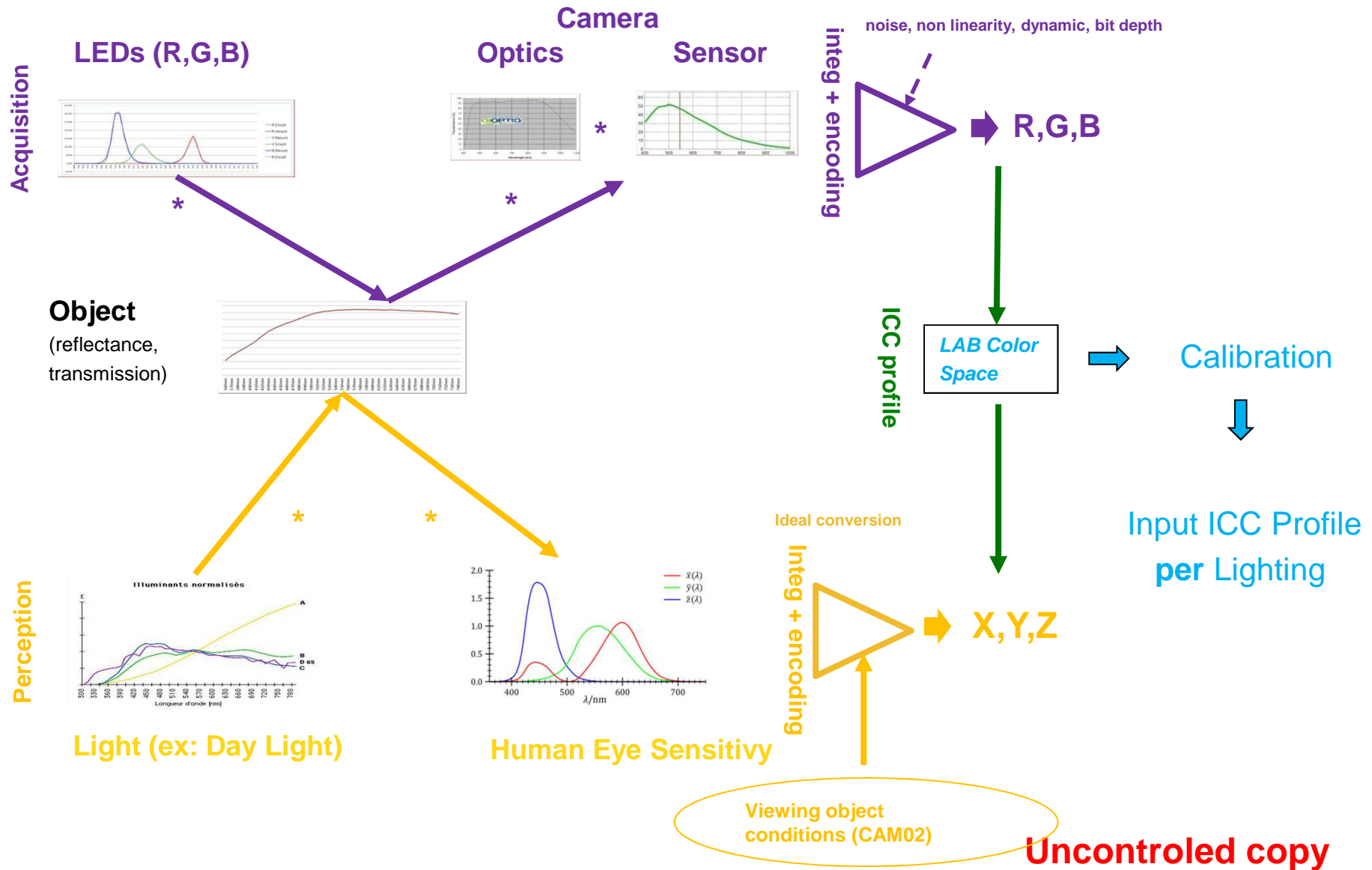


# Imager : Physical description

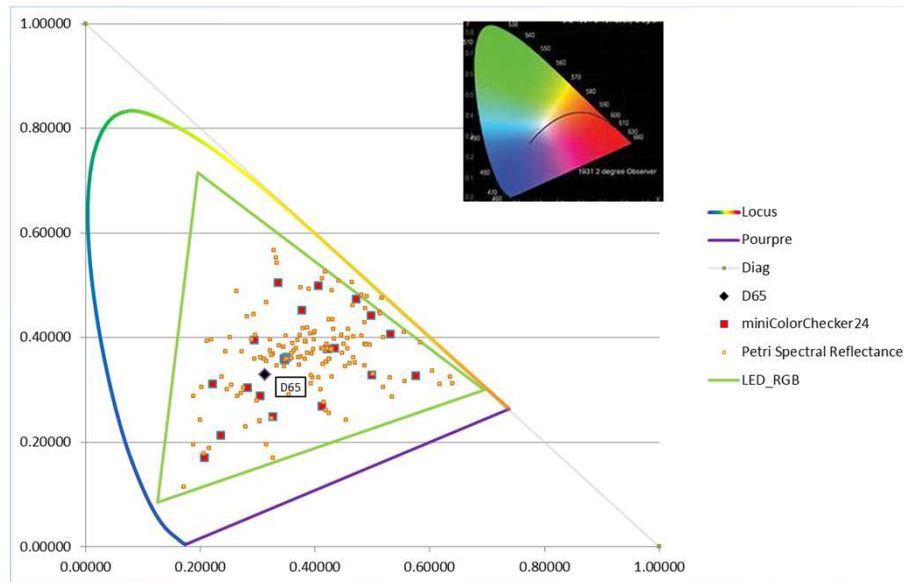
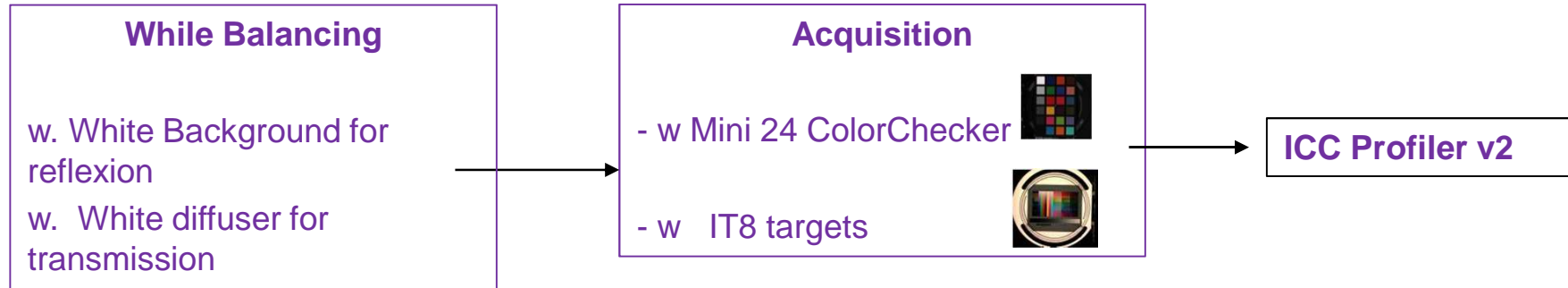


Imager Resolution = 1800 x 1800  
 Pixel resolution = 47-52  $\mu\text{m}$  w.r.t. focal length calibration  
 Field of view = 85,71 - 94,73 mm

# Perceived Colorimetric Model : Input ICC Profile

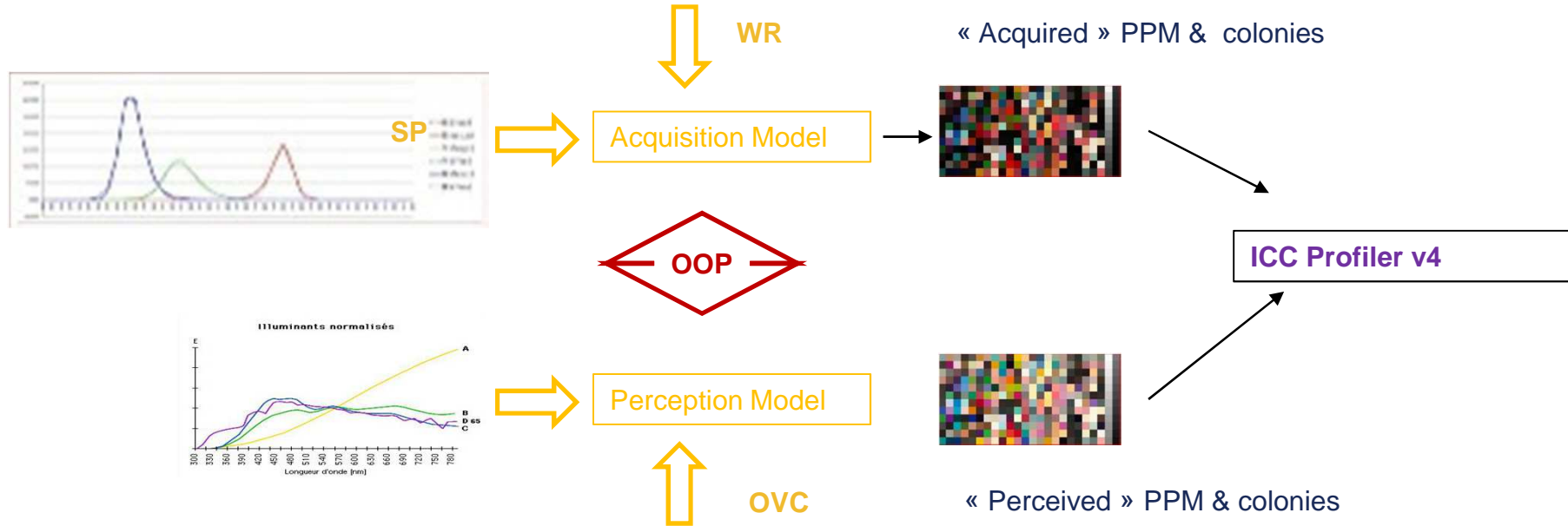


# Petri Imager : current color calibration & goal



**Goal :** minimize the perceived variability (metamerism errors)

# Proposal : ICC Model Base Color Calibration



## Legend :

SP : Spectral Light Properties

OVC : object viewing conditions (CAM02 model)

OOP = Object Optical Properties (Spectral Reflectance, Spectral Transmittance)

WR : white reference

How to **standardise** measure for defining spectral reflectance & transmittance ?

**Uncontrolled copy**

## PPM Plates Imaging : Spectral reflectance measurement

### Spectrophotometer



#### Konica Minolta 2600d

Measurement apertures ( $\varnothing$ ): 8 or 3 mm  
Size of integrating sphere ( $\varnothing$ ) : 52 mm  
Wavelength range : 360 nm to 740 nm  
Wavelength pitch : 10 nm  
Light : 3 pulsed xenon lamps  
Specular included (SCI) & excluded (SCE) measures

### Measurement Setup

#### Spectro photometer

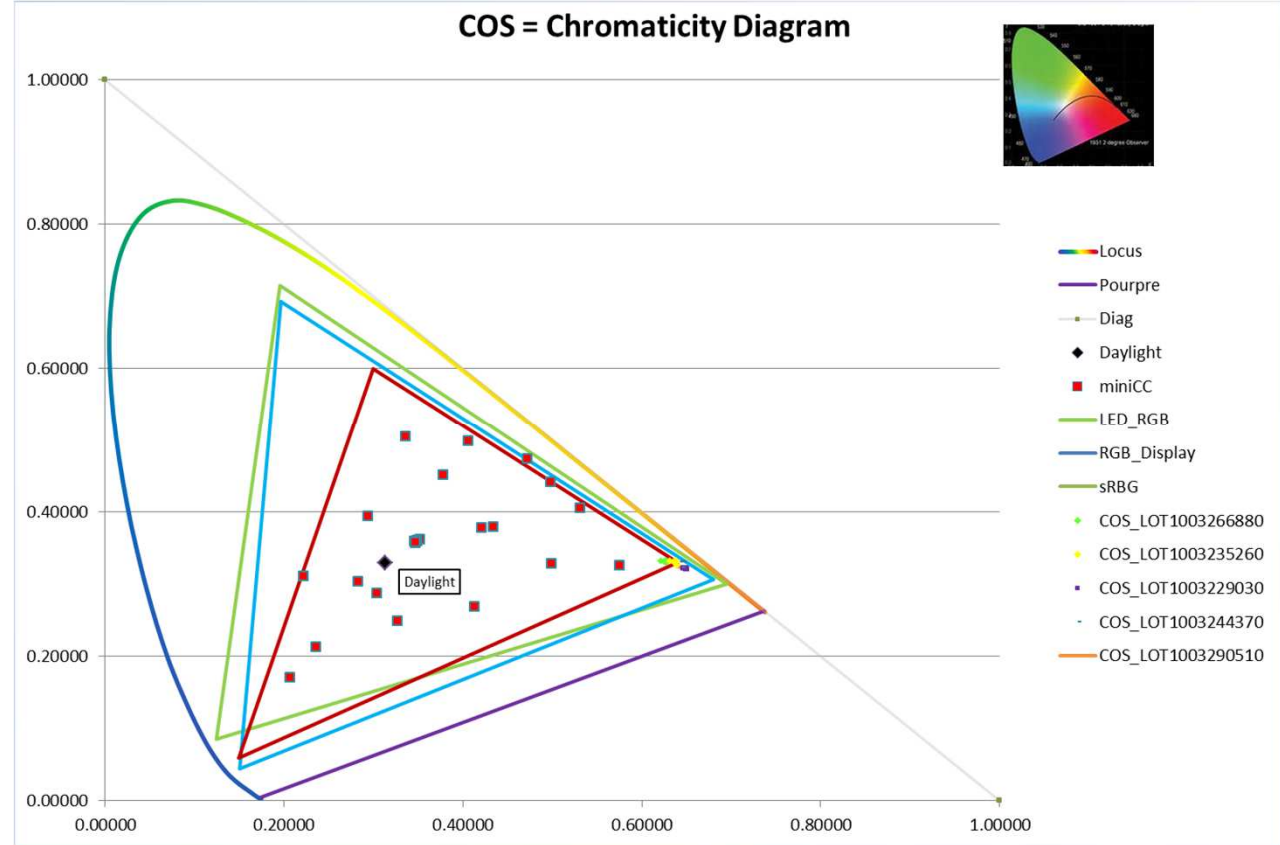
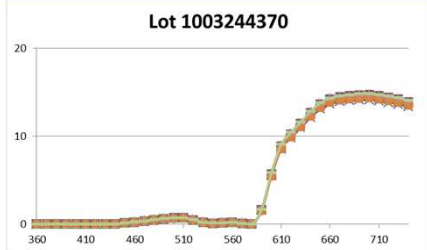
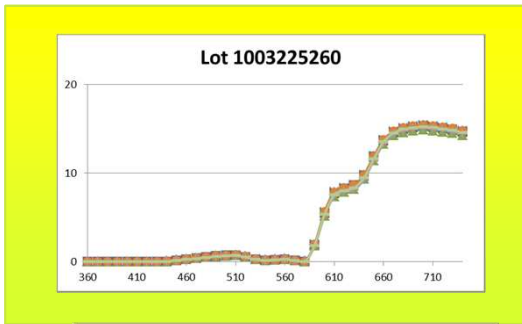
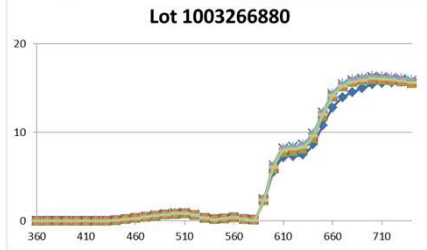


#### Petri Dish

#### Background (White or Black)



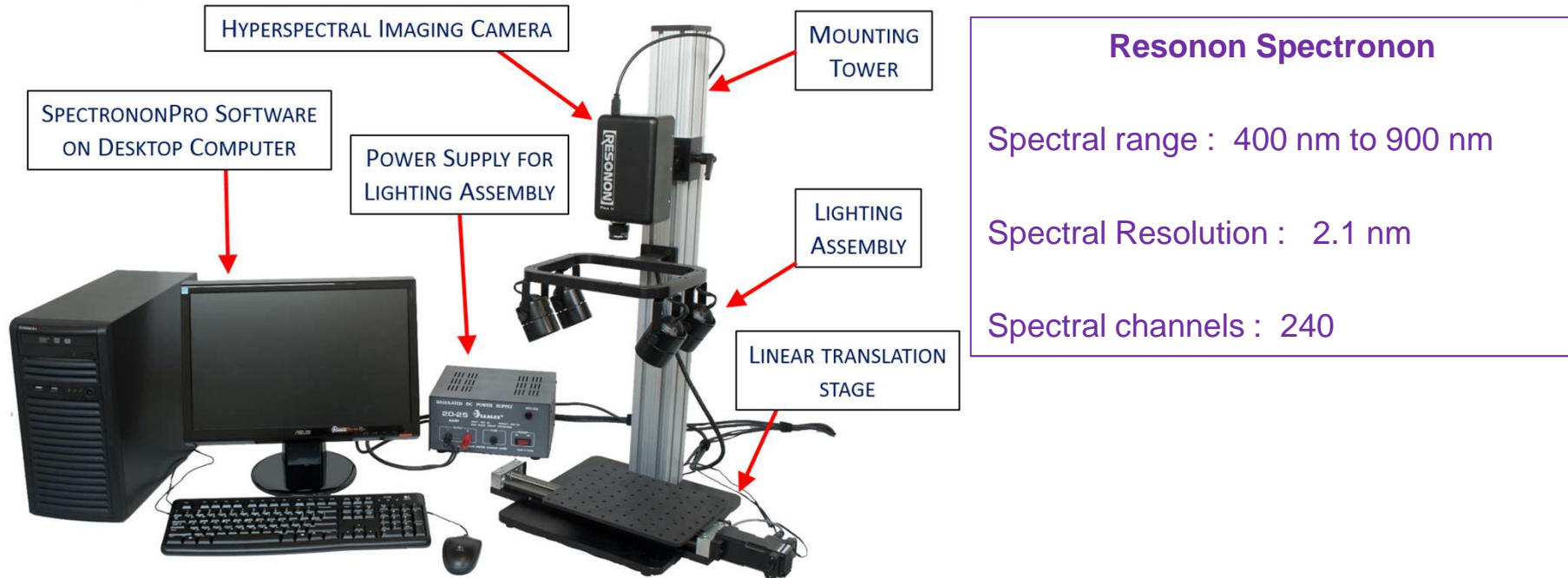
# An example : COS Spectral reflectance to XYZ (D50)



## Current Limitations

- ➔ The minimal size is too small (3 mm) : difficult to measure isolated colonies ( 1 to 2 mm) optical properties
- ➔ No spectral transmittance measure possible with this device
- ➔ Variability of the distance of the device to the petri plates may have an impact (ie : specular component)

# HSI : InVivo Spectral Measurement System



## Can make sense in a biological characterisation context:

- Fast** analysis (<5 seconds) → large spectral signatures database
- High** resolution measurements (< 1 mm) → single colony signature
- Reflectance & Transmittance **simultaneous** measures → no time effect
- Contact less** system → no cleaning & contamination issue

### ICC model based calibration

- Can we build a **normalised** setup to measure spectral signature of petri images (w and wo specular component) ?
- Can we build a **equivalence** measurement system a the golden standard being the spectrophotomer (at least for reflectance) ?



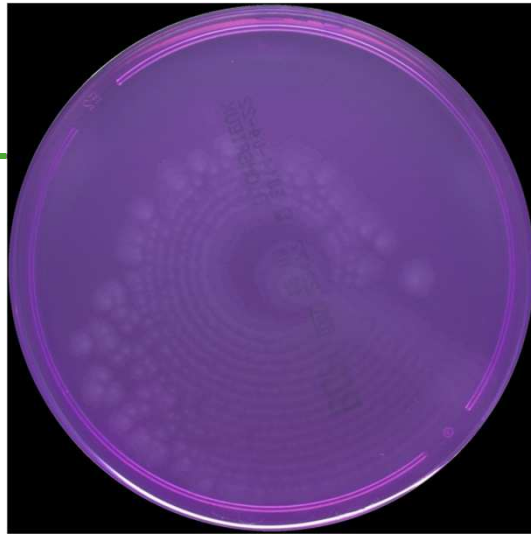
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## **A few images of Petri Plates ....**

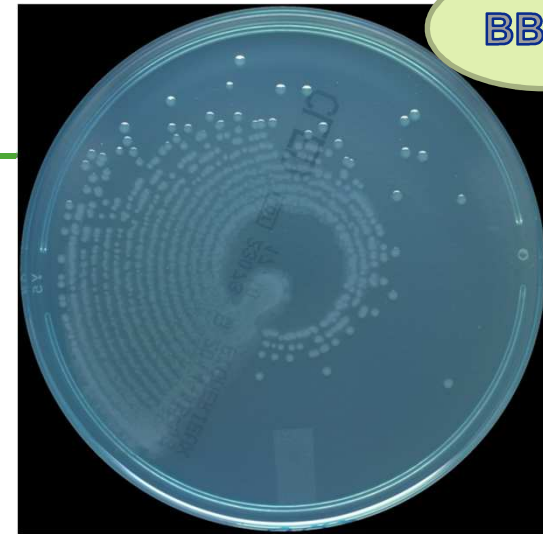




CAN2 / UCA

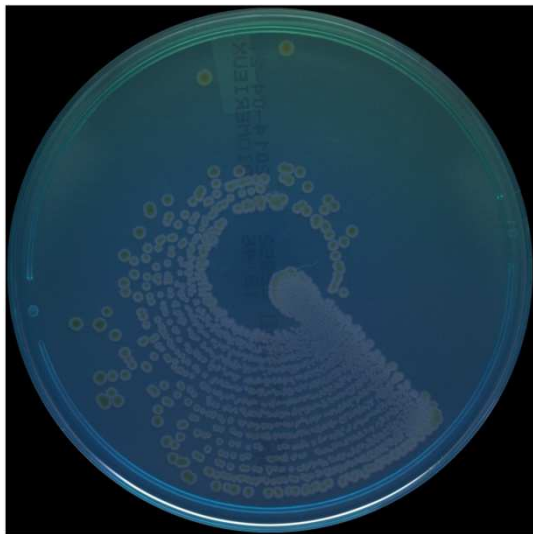


BCP / EQB

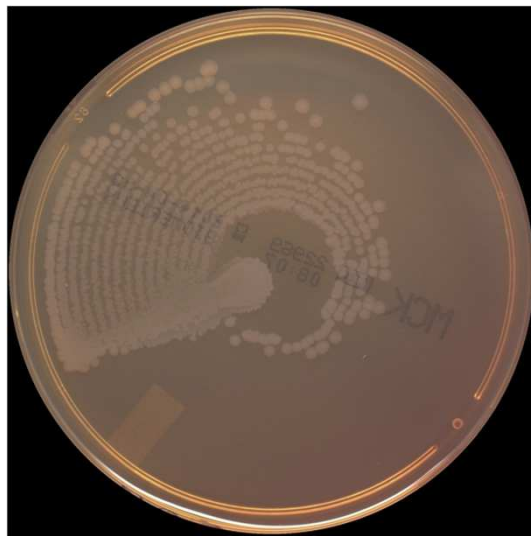


BBAB

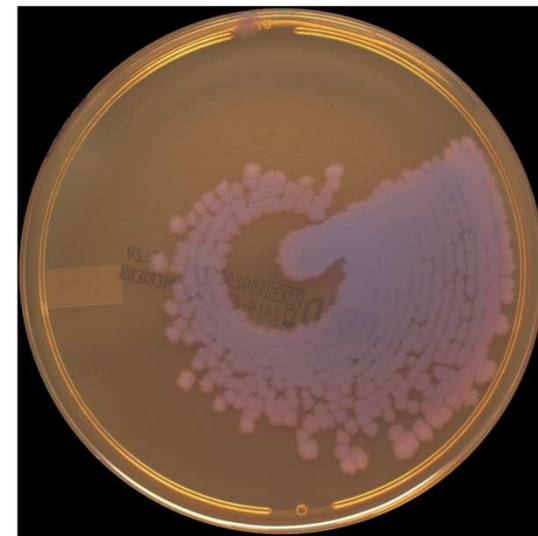
CLED / EQB



DRIG / EQB

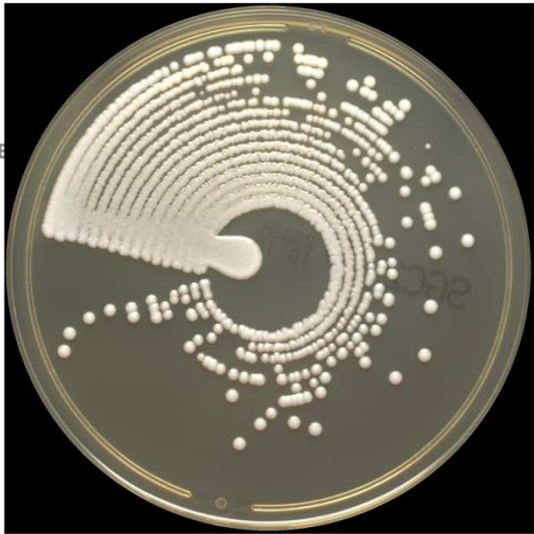


MCK / EQB

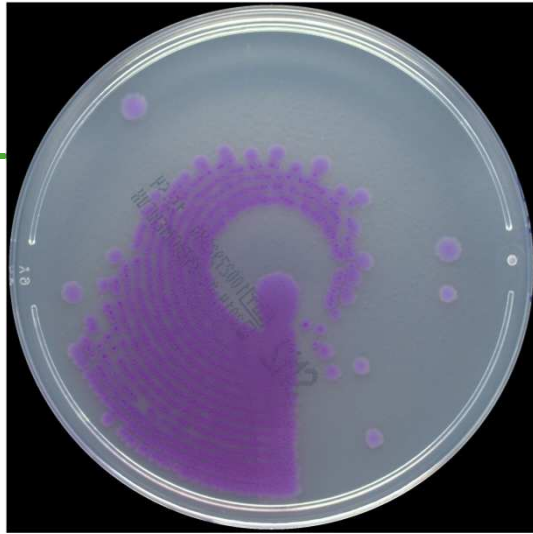


PAID / PPA

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SGC2 / UCA



SM2 / EOM

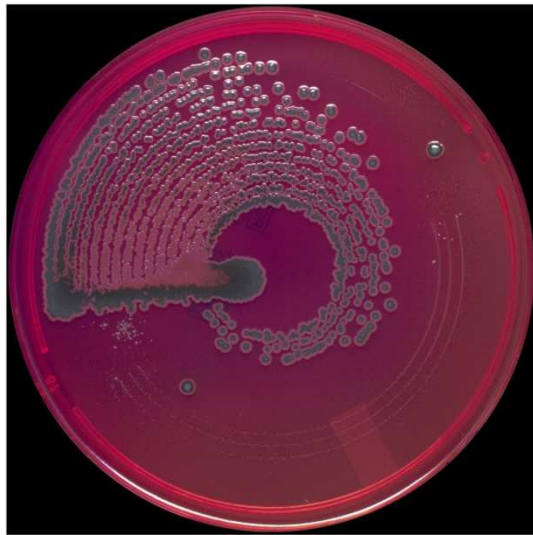


STRPB / SAG

BBAB



URI4 / KPN



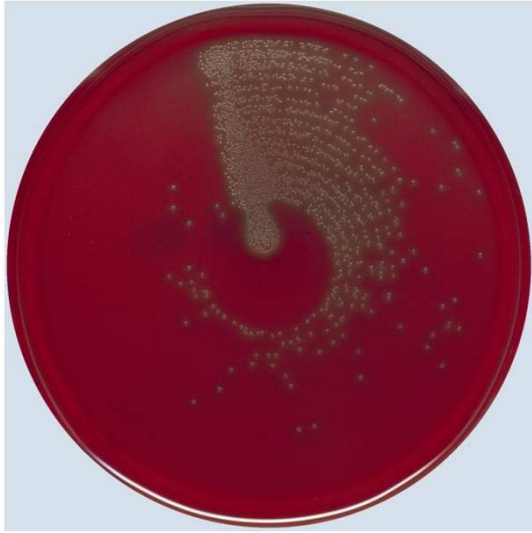
XLD / EOM



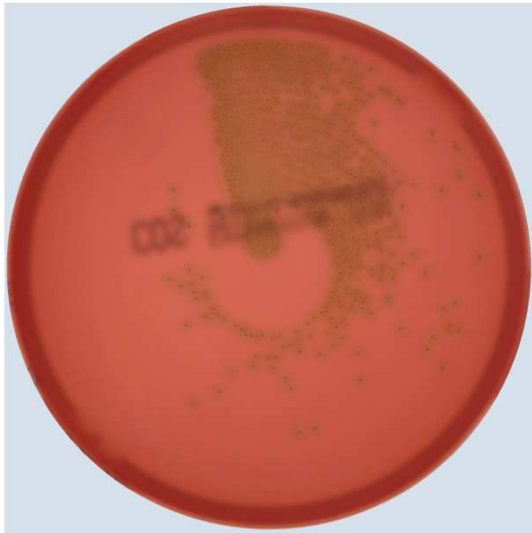
XLD / ESF

Uncontrolled copy

COS / SPN 0510039



BBAB



BL



Uncontrolled copy

**THANK YOU FOR YOUR ATTENTION**

[jeremie.pescatore@biomerieux.com](mailto:jeremie.pescatore@biomerieux.com)



# PETRI PLATES READING & VIEWING :

→ A NEED FOR STANDARDISATION

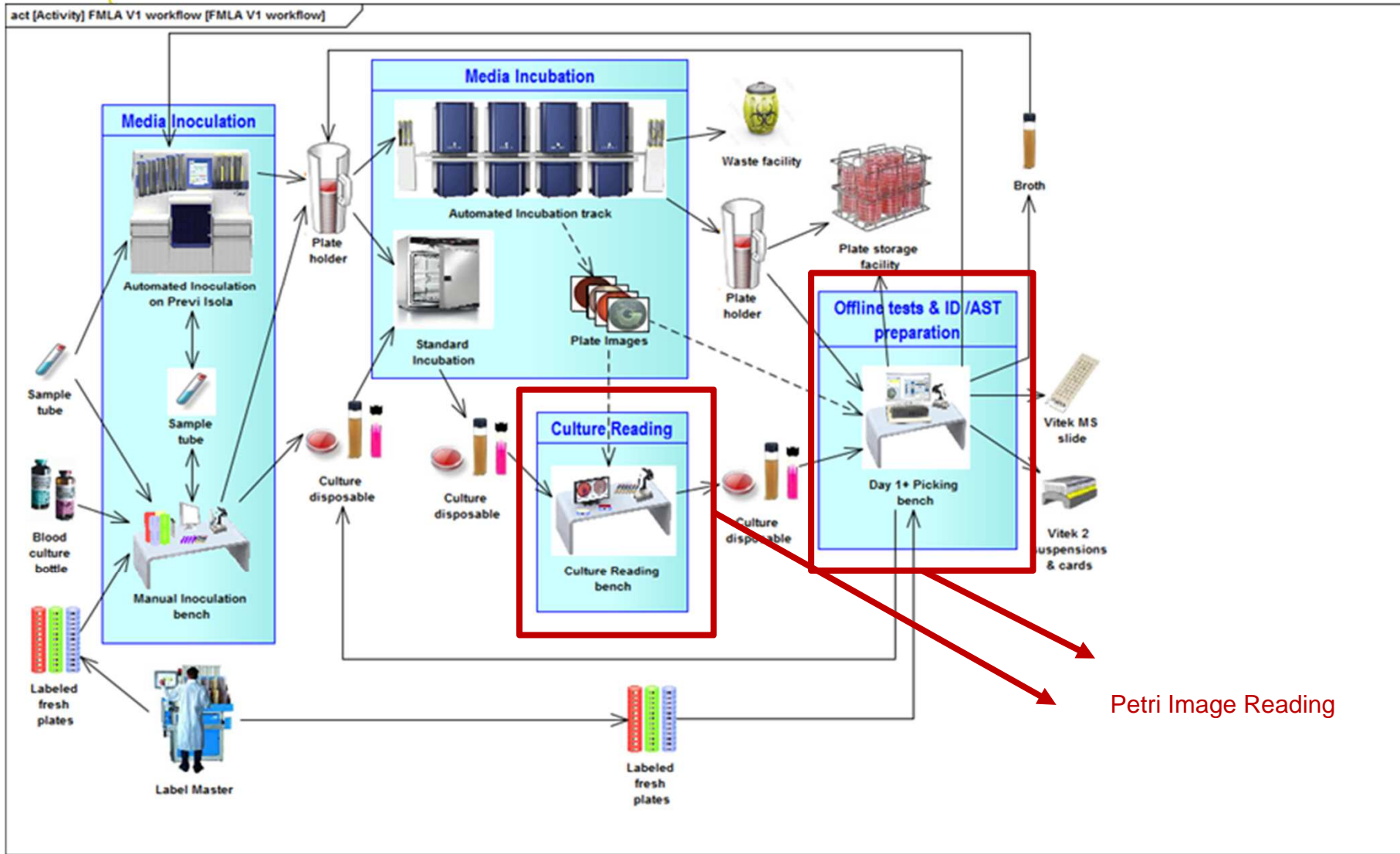
PIONEERING DIAGNOSTICS

Jeremie Pescatore, System Architect, bioMérieux

Revision 0



# Clinical Imaging & Aided Diagnosis (CIAD)



## PPM Plates : Reading Environments

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**Culture Reading =** Reading of petri plates to define « isolates »

**Clean Environment** where **clinical diagnostic** is performed.

→ ambient room conditions may (to some extent) be controlled

**ID / AST Reading =** Reading of petri plates to pick « isolates »

**Dirty Environment :** where **clinical diagnostic** is not necessarily performed

→ ambient room conditions is difficult to control

**Mobile Culture Reading =** Reading of petri plates out of the Laboratory

**Environment :** where **clinical diagnostic may be** performed

→ ambient room conditions is difficult to control

Can be build standardized common viewing conditions = Display ICC Output profile ?

# Culture Reading : Clinical Diagnostics Screen (I)

**Culture Reading**

Accession ID:  Culture ID:  OK

Lab ID: 123456789 Prescription ID: A1518  
Specimen Group / Type: Blood / -  
Collected Date: 7/1/13 3:03 PM

7/1/13 - Blood Culture BacT/ALERT FN Positive g+  
7/1/13 - Gram Stain

7/1/13 - Urine Salmonella spp  
7/1/13 - Blood Specimen Result2  
7/1/13 - Urine Salmonella spp  
7/1/13 - Blood Specimen Result2

Patient ID: 123456789 Age: 1 Years  
Patient Name: SCHNEIDER Thomas Gender: F  
Other specimen

Specimen Information Isolate Requests

Age of Culture: 53 Hours DAY 2

Predefined Comment:  Unselect All 0/100 Item(s) selected

No predefined comment found

Specimen Comment

Isolate	Culture Media	Test Order	Test Order Detail	Status / Result
1	MCK O2 72h	Detail Text		To prepare

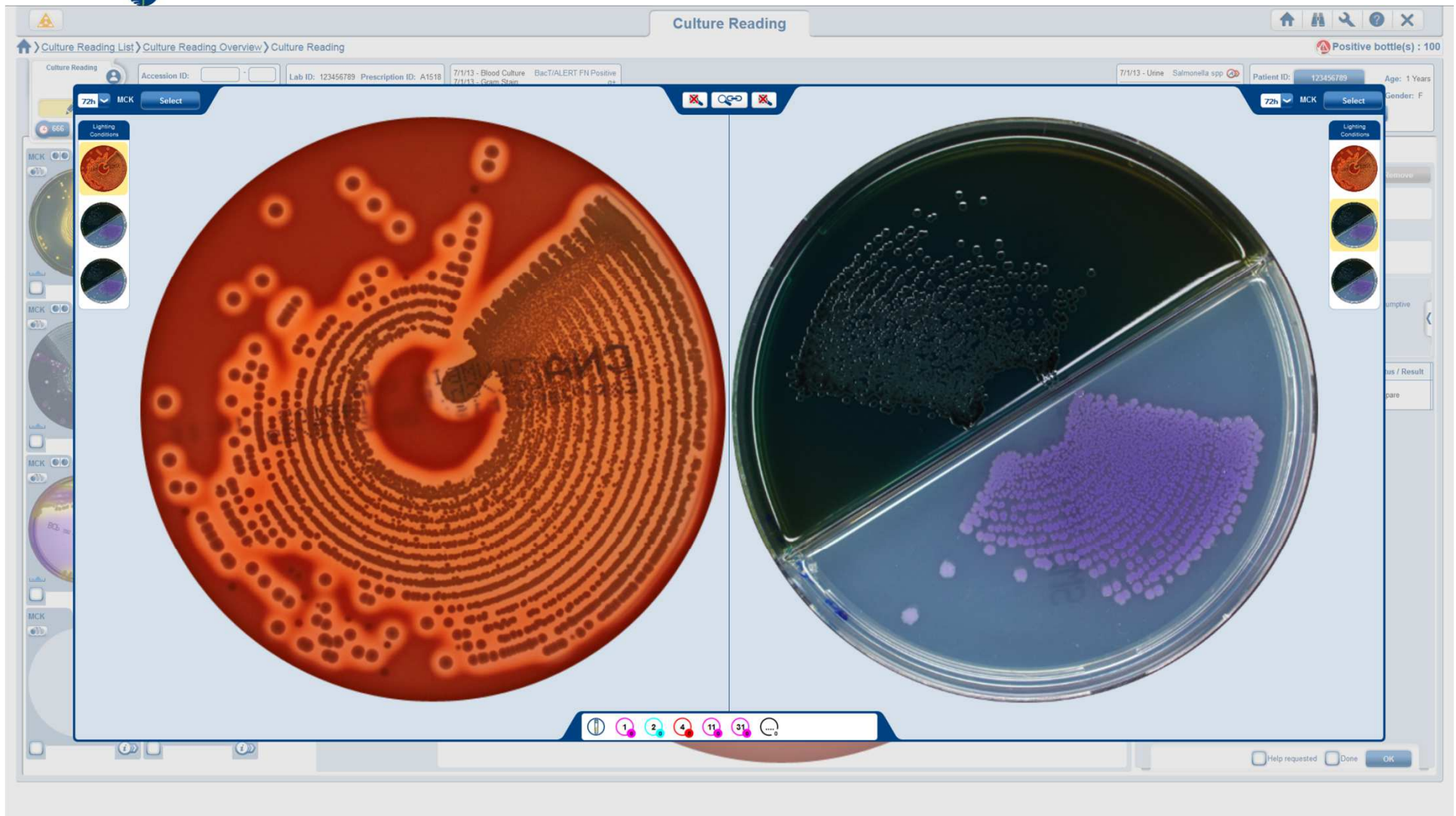
Lighting Conditions Potentially contaminated



Help requested Done OK

- EIZO CG 276 : 16/9 , 27'' (2540 x 1440) w. sRGB gamut
- Default display 1000 x 1000 up to full resolution (in zoom mode) with a « bluish background »

## Culture Reading : Clinical Diagnostics Screen (II)



- EIZO CG 276 : 16/9 , 27" (2540 x 1440) w. sRGB gamut
- Default display 1000 x 1000 up to full resolution (in zoom mode) with a « bluish background »

## ID / AST : Clinical Diagnostics Screen (II)

bioMérieux - PREVI Sightim Manual Picking Station - V1.0.0

Sample Preparation Configuration

Culture Summary Culture ID: **D13** Lab ID: ATTOS\_P1\_S13 Activity Bench Name: Blood

Specimen type: blood

Isolate	Tasks	Results
	VITEK 2 ID ID-YST	Quantity : many Shape : Cocci
	✓ VITEK 2 AST AST-GP	OXIDASE : Negative
1	✓ Purity plate C3_C, COS	Gram : Positive Beta-lactamase : Positive
	i Test GMP	Arrangement : Chained Manual input :

1/2



VITEK® MS

Enter Slide ID:

VITEK® 2

Details

Card Type: ID-YST

Organism ID: No organism ID

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15

X S+ 0

Cassette ID: cds

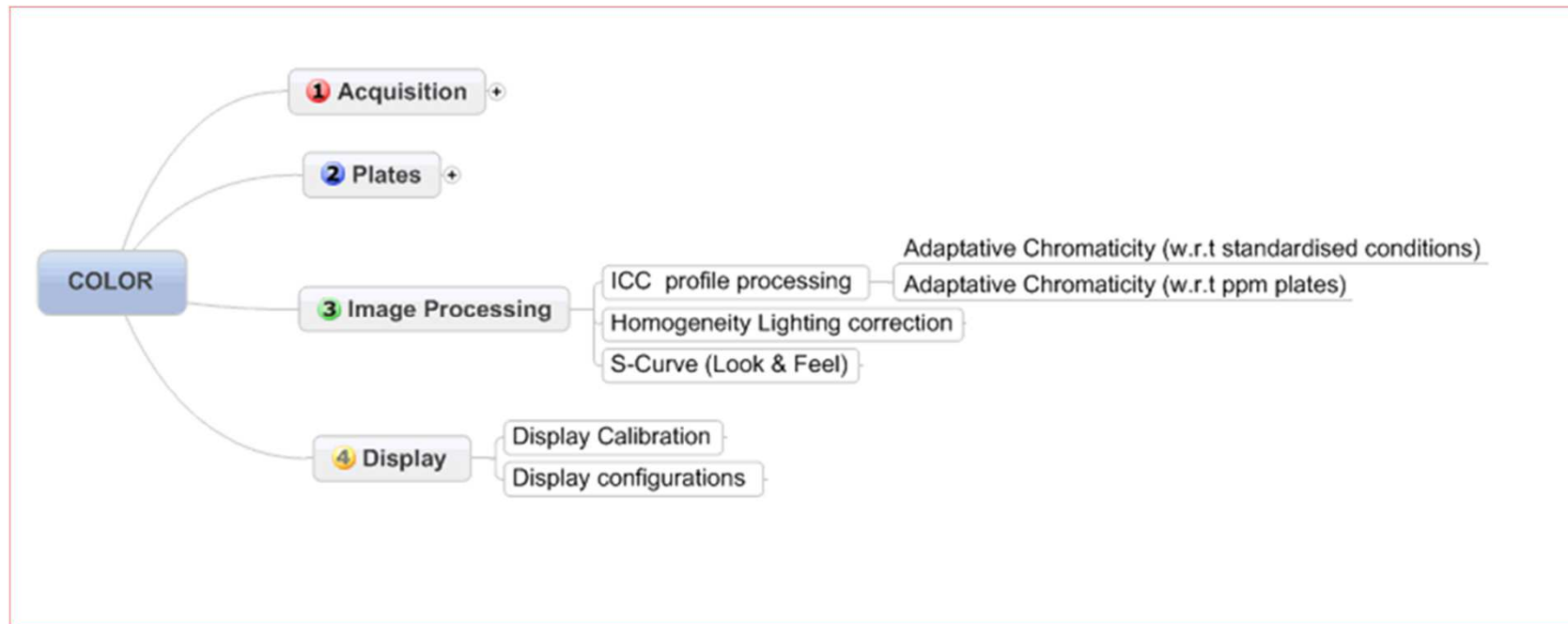
Purity Plate

Agar type	Env.	Culture ID
1/2 C3_C	ENV	IA00000110

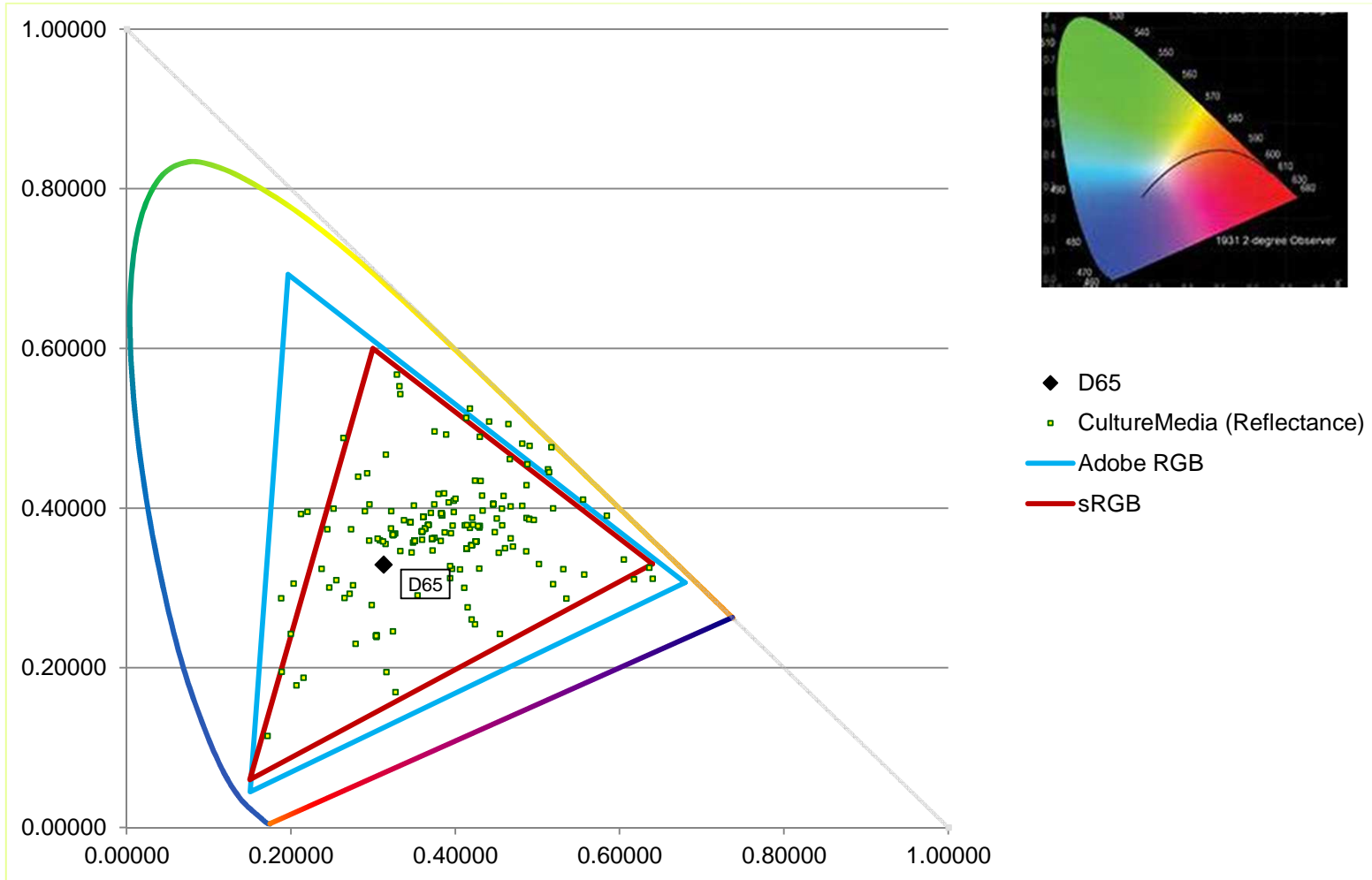
User: fse Bench Name: PREPSTATION1 3:32:41 PM

- ELO Medical 1519 Resolution : **16/9** → 1366 x 768
- Default display **350 x 350** (no zoom mode) with a « **bluish background** »

# Petri Plates Reading & Viewing



# Media Plates Reflectance & Existing Gamut



Can we define a **standardized** gamut w.r.t. to monitor characteristics ?

**Uncontrolled copy**



### Open Points :

1. How should the effect ambient light incorporated and standardized ?
2. How should the effect of the displayed background (chromatic adaptation) standardized with respect to viewing conditions
3. How should the **in/out gamut** of the display w.r.t. to petri optical properties (transmittance & reflectance) shall be standardized ?
4. How should (ie : image format, DICOM ?) output display profile shall be attached to the petri image data in order to display with the same manner on different displays ?

→ **Could mRGB or dRGB color spaces could include these considerations for petri images ?**



## Petri Plates Reading & Viewing

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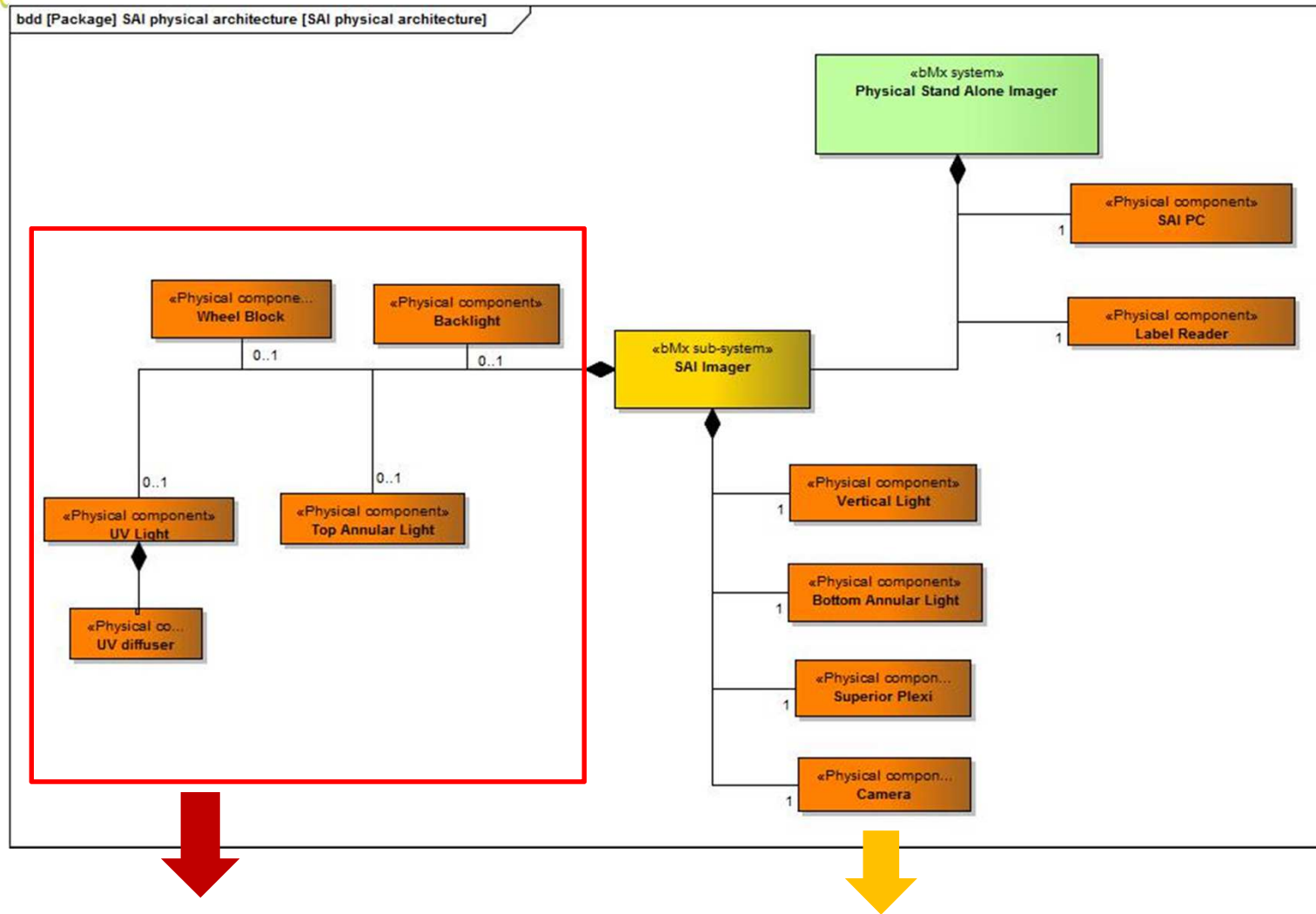
**THANK YOU FOR YOUR ATTENTION**

**[jeremie.pescatore@biomerieux.com](mailto:jeremie.pescatore@biomerieux.com)**

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# Imager : Physical Architecture



**TBC : No need for AST Imaging**

**Optics Change for AST imaging : square 120 - 140 mm**

# WSI Working Group

- Collaboration portal
  - Register an account at <https://nciphub.org>
  - Join the group [https://nciphub.org/groups/wsi\\_working\\_group](https://nciphub.org/groups/wsi_working_group)
- Resources have been added  
[https://nciphub.org/groups/wsi\\_working\\_group/resources](https://nciphub.org/groups/wsi_working_group/resources)
  - Editorial from WG organizers
    - “Evaluating Whole Slide Imaging: A Working Group Opportunity”
    - Motivation and Goals from organizers’ perspectives
  - Slides from Sept. meeting of ICC MIWG
- Survey being developed to collect input
  - WG organizational structure
  - Topics to be pursued
- Ideas for how to proceed